

1 **Influence of abattoir wastes on soil microbial** 2 **and physicochemical properties**

3 4 **ABSTRACT**

5 **Aim:** To assessed the Influence of abattoir wastes on soil microbial and physicochemical properties with regards
6 to its agronomical potential

7 **Design:** Soils impacted with abattoir wastes were collected from two each of Ikot Ekpene (IK1, IK2) and Eket
8 (EK1, EK2) Local Government Areas in Akwa Ibom State, Southern Nigeria. Soils far from the abattoir wastes
9 area were also obtained from the two area and used as control.

10 **Methodology:** Physicochemical, essential and trace metal levels, microbial studies of both abattoir and control
11 soils were carried out using standard analytical and microbiological methods.

12 **Results:** Studied abattoirs and the control soils were in the sandy-clay-loamy soil category with varied
13 quantities of sand, silt and clay. Physicochemical properties of studied abattoir soils were higher than in control.
14 Essential elements (K, Na, Ca, and Mg) and trace metals (Fe, Zn, Cd, Cu, Pb, Cr and Ni) levels were also higher
15 in abattoir soils than in control though were within permissible limit in soil except for Fe. Pollution status
16 calculations using empirical models indicated slight to moderate pollution of abattoir soils by most of the trace
17 metals studied. Microbial studies revealed total heterotrophic bacteria ranged from 6.41 ± 0.43 to 7.91 ± 0.58
18 \log_{10} CFU/g while fungal count ranged from 4.94 ± 0.26 to 5.79 ± 0.34 \log_{10} CFU/g. Among the four (4) locations,
19 IK2 had the highest heterotrophic bacterial densities of 7.91 ± 0.58 \log_{10} CFU/g while IK1 had the highest fungal
20 count of 5.79 ± 0.34 \log_{10} CFU/g. A total of six (6) bacteria (*Klebsiella*, *Micrococcus*, *Pseudomonas*, *Bacillus*,
21 *Escherichia* and *Enterobacter*) and two (2) fungi (*Aspergillus* and *Penicillium*) species were isolated.

22 **Conclusion:** Soil impacted with abattoir wastes is richer in plant nutrients and can be exploited for growing of
23 crops. But it is advised that routine checks be conducted to forestall trace metals accumulation above safe levels
24 in these soils.

25
26 **Keywords:** Microbial, physicochemical properties, abattoir waste, pathogens

27 **INTRODUCTION**

28 Studies have shown that environmental pollution and its attendant problems on land, air and water qualities are
29 severe now than before. There are several evidences to this fact ranging from soil fertility loss, depletion of
30 biodiversity, several health problems (those leading to metabolic disorder), ecological effect and others [1-3].

31 The major cause of these pollutions is the indiscriminate discharge of wastes into these natural habitats thereby
32 tampering with the natural workings of the environment. Solid and liquid wastes are usually disposed off on
33 open landfills, waterways, rivers and stream indiscriminately by most industries (small and large), and the entire
34 populace. These practices are very common in Nigeria till date since there are no well define management
35 protocols on solid waste disposal.

36 Abattoir waste is another emerging solid waste whose rate of generation is becoming alarming. Meat processing
37 is usually carried out in a specialized environment known as abattoir or slaughter house. According to [4]; it is a
38 place or building where animals are killed and processed for their meat. Several activities are involved in the
39 operation including receiving and holding of livestock, slaughter carcass, dressing of animals, chilling of carcass
40 products, carcass boning and packaging, drying of animal skins [5]. However, in Nigeria, meat processing
41 activities are mostly carried out in unsuitable places or building by butchers who have little or no idea of
42 sanitary principles. These activities are usually accompanied by the generation of large amount of wastes like

43 blood, fat, organic and inorganic solids, salts which are hitherto discharged into soils and water bodies around
44 the abattoir premises [6 – 7].

45 Abattoir wastes have complex composition and can be very detrimental to any environment where they are
46 discharged. Various organs of cattle like muscles, blood, liver and kidney have been reported to contain trace
47 metals, faeces of livestock consist of mucus, bacteria, cellulose fibre, paunch manure which is very acidic in
48 nature and others [8 – 11]. Additional reports have been made on the effect of abattoir wastes on soil including
49 increase concentration of trace metals, increase population of decomposers, loss of aesthetic value, excessive
50 soil nutrient enrichment and increase toxin accumulation, as well as large accumulation of sulphides,
51 mercaptans, amines and organic acids [3, 11, 12 -15].

52 In Akwa Ibom State, most abattoirs have farmlands surrounding them and these farmlands are usually cultivated
53 by local farmers living in the vicinity of these abattoirs. These soils are used to plant crops mostly vegetables
54 and legumes which are consumed by humans due to good crop yields from these abattoir soils without any
55 assessment of its sanitary nature. Therefore this study seeks to investigate the influence of abattoir wastes on the
56 physicochemical properties, trace metal levels and diversity of microorganisms in such soils.

57 **Materials and Method**

58 *Sample collection, treatment and analysis*

59 Top soil samples were obtained from four different abattoir soils, two each from Ikot Ekpene (IK1, IK2), and
60 Eket Local Government Area (EK1, EK2) respectively in Akwa Ibom State, Southern Nigeria. At each Local
61 Government Area, top soil not close to a slaughter house or abattoir waste was also collected as Control samples
62 (IKC, EKC). The abattoir soils were collected after clearing off waste materials from the soil within the vicinity
63 of these abattoirs and were done in January 2018. Soil samples were collected by using Soil Auger to obtain soil
64 from the depth of 0 – 20 cm. A total of six (6) soil samples were collected for this study. Soil samples for
65 microbial studies were placed in sterile polythene bags and transported to the laboratory for immediate analysis.
66 For chemical analysis, samples and Control were air dried for three days to drive off any liquids, ground and
67 sieved using a 2mm mesh. One gram of the sieved samples and Control was mixed with Aqua Regia
68 (HCl/HNO₃ 3:1) and digested on a hot plate. The filtrate obtained was used for the determination of total Fe, Zn,
69 Cd, Cu, Pb, Cr, Mg, and Ni levels using Agilent 710 Inductively Coupled Plasma Optical Emission
70 Spectrometer (ICP-OES). Na, K and Ca were determined using flame emission spectrophotometer (Model
71 381&391) at different wavelengths

72 73 *Determination of textural class and physicochemical properties*

74
75 Particle size distribution through the physical analytical test was carried out using the hydrometer methods as
76 describe by [16]. Textures class of the abattoir soils and Control were determined using United States
77 Department of Agriculture (USDA) textural triangle, while Bulk density was determined by gravimetric method
78 as described by [17]. The pH of studied abattoir soils and Control was determined in a 1: 2.5 (v/v) soil/water
79 suspension as described by [18]. Electrical conductivity was measured using portable meters after calibration
80 with standard solutions. Salinity was determined following the method of [19]. Moisture content was
81 determined gravimetrically after drying the soils in an oven (Gallenkamp OV-330) at 105 °C until a constant
82 weight was obtained. Total organic matter content was measured by wet oxidation methods of Walkley and
83 Black reported by [20]. Total organic carbon and cation exchange capacity were determined using the methods
84 of [21] and [22] respectively. Total petroleum hydrocarbon content, nitrogen and phosphorus were done by
85 methods describe by [23 – 25] respectively in their previous works.

86 87 *Microbiological Analysis of the Abattoir soil Samples*

88 The microbiological analysis of the soil samples were carried out according to the methods described by [26].
89 Bacterial and Fungal Counts were enumerated using pour plate method as described by [27]. The bacteria
90 culture plates were incubated at 37°C for 48 hours, the fungal plates were incubated at room temperature (28 ±
91 2°C) for 4 days. The emerging colonies were enumerated using the Quebec colony counter and recorded as
92 colony forming unit per gramme of sample (CFU/g).The emerging colonies after the incubation period were

93 discretely isolated and sub-cultured repeatedly on freshly prepared Nutrient agar for bacteria and Sabouraud
 94 Dextrose agar for fungi to obtain pure isolates. The pure isolates were maintained on agar slants and stored at
 95 4°C for further use. The bacterial isolates were characterized based on their cultural and morphological
 96 attributes as well as their responses to standard biochemical test as described by [28] and identified as described
 97 in Bergey’s manual of determinative bacteriology [29]. Fungal isolates were characterized on the basis of their
 98 cultural attributes, and identified by consulting various taxonomic books and monographs available on various
 99 groups of fungi [30].

100 *Determination of Pollution status of trace metals in studied abattoir soils.*

101

102 *Metal pollution index (MPI)*

103

104 Metal pollution index (MPI) usually indicates the relationship between metal in studied soil and in reference soil
 105 (Control). MPI was calculated in this work using equation (1) given below

106

107
$$MPI = \frac{\text{Concentration of metal in studied soil}}{\text{Reference soil (control)}} \text{----- (1)}$$

108 The classifications of MPI according to [31] are given in Table 1 below

109 Table 1: Classification of metal pollution index in soil and their implications

MPI	Significance	Remarks
< 0.1	Very slight contamination	No negative effect on soil, plant and environment
0.10 -0.25	Slight contamination	No negative effect on soil, plant and environment
0.26 – 0.50	Moderate contamination	No negative effect on soil, plant and environment
0.51 – 0.75	Severe contamination	No negative effect on soil, plant and environment
0.76 – 1.00	Very severe contamination	No negative effect on soil, plant and environment
1.10 – 2.00	Slight pollution	Will pose negative effect on soil, plant and environment
2.10 – 4.00	Moderate pollution	Will pose negative effect on soil, plant and environment
4.10 – 8.00	Severe pollution	Will pose negative effect on soil, plant and environment
8.10 – 16.00	Very severe pollution	Will pose negative effect on soil, plant and environment
> 16.00	Excessive pollution	Will pose negative effect on soil, plant and environment

110

111 *Degree of contamination (Cdeg)*

112 Degree of contamination of each location within the studied abattoir soils and Control was calculated using
 113 equation (2)

114
$$Cdeg = \sum MPI \text{-----(2)}$$

115 where MPI denotes the sum of metal pollution index for all the elements at a particular location. The different
 116 classifications of Cdeg according to [32] are as follows: Cdeg < 8 = low degree of contamination, 8 < Cdeg < 16
 117 = moderate degree of contamination, 16 < Cdeg < 32 = considerable degree of contamination and 32 < Cdeg =
 118 very high degree of contamination.

119

120 *Enrichment factor (EF)*

121

122 Enrichment factor and percent enrichment of each metal were determined using equations (3) and (4) below

123
$$EF = \frac{\left(\frac{M}{Fe}\right) D}{\left(\frac{M}{Fe}\right) C} \text{----- (3)}$$

124 where D = Abattoir soils studied; C = Control; M = metal studied; Fe in the equation is used for normalization
 125 [33]. EF values close to one indicate natural origin; values less than 1.0 suggest a possible mobilization or
 126 depletion of metals, while EF > 1.0 indicates that the element is of anthropogenic source [34].

127 *Geoaccumulation index (Igeo)*

128 Geo-accumulation index (Igeo) of trace metals in studied soil was determined using equation (5) below

$$I_{geo} = \text{Log } 2(C_n/1.5B_n) \text{ ----- (5)}$$

129 where C_n is the measured concentration of metal in studied abattoir soils, B_n is the concentration of metal in
 130 Control site while 1.5 is a constant to allow for fluctuations of a given metal in the environment as well as small
 131 anthropogenic influences [35]. The different classes of geo-accumulation index as proposed by [36] are as
 132 follows: $I_{geo} < 0$ = unpolluted, $0 - 1$ = unpolluted to moderately polluted, $1 - 2$ = moderately polluted, $2 - 3$ =
 133 moderately to strongly polluted, $3 - 4$ = strongly polluted, $4 - 5$ strongly to extremely and $I_{geo} > 5$ = extremely.
 134

135 *Pollution load index (PLI)*

136

137 Pollution load index (PLI) of metals in a particular location was obtained using equation (6).

$$PLI = (\text{MPI}_{Fe} \times \text{MPI}_{Pb} \times \text{MPI}_{Cd} \times \text{MPI}_{Zn} \times \text{MPI}_{Cu} \times \text{MPI}_{Cr} \times \text{MPI}_{Ni})^{\frac{1}{7}} \text{ ----- (6)}$$

138

139 Where MPI represents metal pollution index for the metals at each location. The different categories of PLI as
 140 proposed by [37] are as follows: No pollution ($PLI < 1$), moderate pollution ($1 < PLI < 2$), heavy pollution ($2 <$
 141 $PLI < 3$) and extremely heavy pollution ($3 < PLI$)

142 **Results and Discussion**

143 Results for bulk density (gcm^{-3}), pH, and textural characteristics (quantity of sand, silt and clay) of abattoir and
 144 Control soils are presented in Table 2

145 Table 2: Quantity (g/kg) of sand, silt and clay, textural class, bulk density and pH of abattoir soils and
 146 Control from Akwa Ibom State, Southern Nigeria

Parameters	Sampling points					
	IK1	IK2	IKC	EK1	EK2	EKC
Sand (%)	58	55	48	56	52	50
Silt (%)	12	8	15	10	14	17
Clay (%)	23	22	18	25	20	24
Texture	SCL	SCL	SCL	SCL	SCL	SCL
Bulk density (gcm^{-3})	1.42	1.28	1.14	1.38	1.36	1.17
Soil pH*	5.30	5.18	4.60	4.95	4.78	4.74

147 pH* (1:2.5 soil: water ratio); SCL – Sandy-clay-loamy soil; IK1-Ikot Ekpene abattoir soil 1; IK2-Ikot Ekpene
 148 abattoir soil 2; IKC-Ikot Ekpene control soil; EK1-Eket abattoir soil 1; EK2 - Eket abattoir soil 2; EKC - Eket
 149 control soil

150 The results indicated varied textural characteristics, bulk density and soil pH among the studied abattoir
 151 soils and Controls. For the two abattoir soils, the percentage of sand, silt and clay in the soil samples were
 152 within the range of 50 – 58% (sand), 8 -15% (silt) and 20 -25% (clay); while their Controls were 48 – 50%
 153 (sand), 15 - 17% (silt) and 18 - 24% (clay). The percentages of sand, silt and clay obtained for abattoir soils in
 154 this work agrees with 52 – 59% (sand), 10 -16% (silt) and 25 -30% (clay) reported by [19] for abattoir soils
 155 from Port Harcourt, River State, Southern Nigeria but disagrees with 76 – 83% (sand), 1.5 – 2.0 % (silt) and 13
 156 -23% (clay) reported by [38] for abattoir soils from Calabar, Cross River State. All the abattoir soils studied and
 157 their Controls fell in the sandy – clay – loam (SCL) class of soil, with higher percentage of sand, followed by
 158 clay and then silt. Soil texture parameter was measure so as to reveal the physical properties of the soil such as
 159 water retention capacities, permeability, easy or toughness of tillage of the soil studied among other things.
 160 From these findings, the abattoir soils under investigation were seen to have potential of holding more water
 161 within the particle because of the high percentage of clay ³⁹.

162 Results for bulk density (gcm^{-3}) and pH of abattoir soils and their Controls are presented in Table 2.
 163 The results indicate that both parameters varied differently among the abattoir soils and the Control with bulk
 164 densities having ranges of 1.28 – 1.42 for the two abattoir soils and 1.14 -1.17 for the Control. The result also
 165 reveals that the bulk densities of the abattoir soils were higher than those of the Control. The variations maybe
 166 as a result of differences in soil texture and organic matter content of abattoir soils and the Control. Also,
 167 Abattoir soil recorded greater percentages of sand than the Control soil and soils with higher percentage of sand

168 is usually more prone to high bulk density [17]. The range of bulk density obtained in this work is in agreement
 169 with 1.16 – 1.81 gcm⁻³ reported by [40] for abattoir soils from Abeokuta, South western Nigeria, but in contrast
 170 with 1.50 -1.65 gcm⁻³ reported by [41] for abattoir soils from Makurdi, Benue State, Nigeria. However, the
 171 values for bulk densities obtained in this work are considered suitable for crop production and also within the
 172 critical range [42 -43]

173 The pH values for the two abattoir soils and their Controls recorded in this work were 4.78 – 5.30 and
 174 4.60 - 4.74 respectively as indicated in Table 2. This parameter was determined because pH (acidity or
 175 alkalinity) plays a great role in determining the availability of nutrients in soil to plant and the type of organism
 176 found in the soil [44]. The results indicated that all the soils (abattoir and Control) studied were acidic in nature,
 177 with their Control soils showing higher acidities than the abattoir soils. However, the obtained ranges reported
 178 in this work are lower than 6.22 – 7.44 reported by [45] but are consistent with 4.99 – 6.73 obtained by [5] in
 179 abattoir soils though with slight differences. Also, the observation of higher pH obtained for abattoir soils than
 180 in the Control soil obtained in this work is in line with the findings of [46]. This could be attributed to
 181 biodegradable waste materials in studied abattoir soils which may lead to reduced anaerobic activities in these
 182 soils [47]. Consequently, pH of soils impacted by abattoir wastes could be affected considerably. Although there
 183 is no acceptable standard for pH for an ideal soil for planting as it depends upon the type of crops, researches
 184 have shown that most minerals and nutrients are best available to plants in soil with a pH of between 6.5 -6.8
 185 [48 – 49].

186
 187 Table 3: Physicochemical parameters of selected abattoir and Control soils

Parameters	Sampling points					
	IK1	IK2	IKC	EK1	EK2	EKC
Temperature (°C)	30.45	32.10	27.33	31.67	30.98	28.49
EC (µS/cm)	40.60	38.62	19.56	42.18	44.05	20.86
Salinity (mgkg ⁻¹)	18.00	22.00	10.50	15.00	26.06	13.15
Moisture content (%)	10.35	12.86	3.78	11.70	10.06	4.05
TOM (%)	8.65	7.94	4.37	7.27	8.42	4.64
TOC (%)	13.76	12.92	5.67	14.31	15.05	6.82
CEC (Cmol/kg)	28.63	26.80	20.84	25.11	26.95	21.67
TPH (mgkg ⁻¹)	8.06	7.15	2.61	11.72	13.63	4.52
Nitrogen (%)	0.04	0.06	0.01	0.09	0.10	0.02
Phosphorus (mgkg ⁻¹)	2.21	3.18	0.38	2.19	1.84	1.09

188
 189 *EC – Electrical conductivity; TOM – Total organic matter, TOC –Total organic carbon, CEC – Cation
 190 exchange capacity, TPH – Total petroleum hydrocarbons; IK1-Ikot Ekpene abattoir soil 1; IK2-Ikot Ekpene
 191 abattoir soil 2; IKC-Ikot Ekpene control soil; EK1-Eket abattoir soil 1; EK2 - Eket abattoir soil 2; EKC - Eket
 192 control soil

193
 194 Results for the physicochemical properties of selected abattoir soils and their Controls are presented in
 195 Table 3. Temperature (°C) ranges for the two abattoir soils studied and their Controls were 30.45 – 32.10 and
 196 27.33 – 28.49 respectively. The results also indicate that abattoir soils recorded varied temperatures and were
 197 higher than those of their respective Controls. The reasons for the differences in the temperatures among the
 198 abattoir soils may be attributed to factors such as variation in water content of the abattoir soils, soil relief and
 199 cover [19]. Temperature range for the abattoir soils obtained in this work is lower than (33.60 – 35.30) °C
 200 reported by [19] for abattoir soils from Port Harcourt, Rivers State, but higher than (18.80 – 21.43) °C reported
 201 by⁵⁰ for abattoir soils from Delta State, Nigeria. Temperature range for the Control soils recorded in this work is
 202 in agreement with 27.33 – 29.00°C reported by [26] for soils from different part of Southern Nigeria.

203 Levels of electrical conductivities (EC) in µS/Cm varied from 38.62 to 40.60 for abattoir soils obtained
 204 from IK, 42.18 to 44.05 for abattoir soils from EK and 19.56 -20.86 for the Control soils (Table 3). Ranges
 205 obtained for EC in studied abattoir soils are higher than 2.03 – 2.54 µS/Cm reported by [51] but lower than
 206 60.00 – 110.00 µS/Cm obtained by [45] in abattoir soils. Electrical Conductivity levels in studied abattoir soils
 207 were higher than values obtained at the Control soils, which is indicative of negative impact of abattoir wastes
 208 on studied soils. The findings of higher EC in abattoir soils than in Control are consistence with the report of

209 [52]. This could be attributed to the low cation exchange capacity (CEC) of Control soil and variations in rates
210 at which metallic salts and organic matter complexes are formed [53 - 54]. Hence, EC of abattoir waste-
211 impacted soils could be significantly affected by the wastes. However, EC values recorded in abattoir soils in
212 this study is within the permissible limit (below $100\mu\text{S}/\text{Cm}$) stipulated by [55].

213 Ranges for salt content (mgkg^{-1}) of abattoir and Control soils as presented in Table 2 were 15.00 -
214 26.06 and 10.50 – 13.15 respectively. This result indicates that soil content of abattoir soils were higher than
215 those of the Control soils. Although, salt content in soil are caused by natural factors such as weathering,
216 continuous irrigation or pouring of wastewater (after washing of animal parts) on soil can also increase salt
217 content of soil. This is so because almost all water contains some dissolve salts. Range for salinity of abattoir
218 soils obtained in this study is lower than 29.00 – 59.00 mgkg^{-1} reported by [19] and (475.05 – 667.88 mgkg^{-1}) by
219 [45] for abattoir soils from Port Harcourt, River State, Southern Nigeria. However, the abattoir soils recorded
220 salinity values that are within permissible limit (200 mgkg^{-1}) established for soil. The Low values of salinity
221 recorded in abattoir soil is advantageous, since high salinity in soil usually leads to poor plant growth and lower
222 soil microbial activity caused by osmotic stress and toxic ions [15].

223 Moisture content in % varied from 10.35 to 12.86 for abattoir soils obtained from IK, 10.06 to 11.70
224 for abattoir soils from EK and 3.78 to 4.05 for the Control soils (Table 3). The variations in these values
225 especially between the abattoir soils and the Control may be as a result of the effect of the abattoir effluent on
226 soil. Ranges obtained for moisture content in studied abattoir soils are higher than 7.03 – 9.54% reported by [56]
227 but lower than 19.01 – 21.07% obtained by [19] and 17.91 – 19.50% obtained by [40] in abattoir soils. The
228 findings of higher moisture content in abattoir soils than in Control are consistency with the report of [52]. This
229 observation can be explained by the fact that in ruminants, the first stomach or paunch contains undigested
230 materials or paunch manure. The paunch manure which is disposed on the soil could have a moisture content of
231 about 88% [10].

232 Results in Table 3 indicate ranges for total organic matter (TOM) for IK, EK abattoir soil and Control
233 soil as 7.94 – 8.65%, 7.27 – 8.42% and 4.37 – 4.64 respectively. These ranges are higher than 0.69 – 7.42%
234 reported by [3] for abattoir soils, and is inconsistent with values obtained by [57] but lower than 5.57 – 24.13%
235 obtained by [5]. Values of OM obtained in studied abattoir soils were also higher than values at Control which is
236 consistent with report by [58 – 59]. This disparity may be attributed to the absence of biodegradable wastes at
237 Control site thereby indicating that; waste materials from abattoir may increase the OM of soil significantly.
238 Also, the faeces of livestock have been observed to consist of undigested food which hitherto will increase the
239 OM content of abattoir soil [10]. However, soil organic matter usually act as a “storehouse or reservoir” for
240 most metals hence it can influence their availability in soil either positively or negatively [46].
241

242 Total organic carbon (TOC) results indicated ranges of 12.92 – 15.05% for the abattoir soils and 5.67 -
243 6.82% for the Control soils. The Control soils recorded lower TOC than the abattoir soils. This may be due to
244 high organic matter content of the abattoir soil. This observation corroborates with the reports of [3] and [60] in
245 their respective studies. Ranges obtained for abattoir soils in this study are lower than 6.1 – 7.6% reported by
246 [38] for abattoir soil from Calabar Metropolis, Cross River State Nigeria, but higher than 12.68 – 30.02%
247 reported by [19]. The differences in the reported values of total organic carbon and those earlier reported for
248 abattoir soils may be due to the rate of decomposition and composting of animal wastes such as dung, body part,
249 blood, bones etc [3]. Organic carbon content in soil plays a vital role in soil development, fertility, and moisture
250 availability in soil.

251 Cation exchange capacity (CEC) of studied abattoir soils varied from 26.80 to 28.63 Cmolkg^{-1} for IK
252 abattoir soils, 25.11 to 26.95 Cmolkg^{-1} for EK abattoir soils, and 20.84 to 21.67 Cmolkg^{-1} for the Control soils
253 (Table 3). The result also shows that the respective abattoir soils recorded higher CEC than the Control soils.
254 This is may be due to higher total organic matter content of the abattoir soils than in the Control soils including
255 the clay content of the soil [61]. Higher content of CEC in abattoir soils than in the Control soils obtained in this
256 study is in agreement with the report of [62]. Also, the obtained ranges of CEC in abattoir soils are higher than

257 12.54 - 16.84 Cmolkg^{-1} reported by [57] in abattoir soils. CEC is a measure of the soil's ability to hold
258 positively charged ions. It is very important to plant as it influences soil structure stability, nutrient availability,
259 soil hydrogen concentrations (pH), and the soil's reaction to fertilizers and other ameliorants [63]. Although
260 most crops do well in soil with low CEC, but vegetables and other productive food crops like vegetables are
261 perform best in soil with moderate to high CEC [64]. This study thus reveals the impact of abattoir wastes on the
262 CEC of soils impacted by wastes generated from abattoir activities.

263

264 Results presented in Table 3 reveals that total petroleum hydrocarbons (TPH) content of the abattoir
265 soils ranged from 7.15 - 8.06 mgkg^{-1} for abattoir soil samples from IK, 11.72 – 13.63 mgkg^{-1} for abattoir soils
266 from EK and 2.61 – 4.52 mgkg^{-1} for the Control soils. For the abattoir soils, ranges obtained in this study is
267 within the range (11.37 -27.68 mgkg^{-1}) reported by [19] for abattoir soils from Port Harcourt, River State,
268 Nigeria. EK abattoir soils recorded higher TPH content than IK abattoir soils as shown in the Table 3. This is
269 because EK hosts some oil companies like Exxon Mobil PLC, while there is no such in IK. For the Control
270 soils, the range obtained in this study is lower than 3400 – 6800 mgkg^{-1} reported by [65], for soils from Owaze
271 in Abia State, and 581.02 mgkg^{-1} reported by [61]. The large variation in the reported values of TPH content for
272 the Control soils in this study and those earlier reported is as a result of lesser frequency of crude oil spill in the
273 Control site used for this study. Areas of frequent crude oil spillage are expected to have higher TPH values than
274 those of sparsely crude oil spillage. However, the lower concentrations of TPH in both abattoir and Control soils
275 in this study is still a source of concern as TPH have been reported as a contaminant in any environment due to
276 its toxicity to humans and other environmental receptors [66].

277

278 Total nitrogen content (%) (both as NH_4^+ - N and NO_3^- - N) and phosphorus (mgkg^{-1}) of the abattoir
279 soils and Control soils are presented in Table 3. For Nitrogen, ranges are 0.04 – 0.06; 0.09 – 0.10 and 0.01 –
280 0.02 for IK, EK and Control soils respectively, while 2.21 – 3.18, 1.84 – 2.19 and 0.38 - 1.09 for IK, EK and the
281 Control soils respectively. The variations between nitrogen and phosphorus contents among the abattoir and
282 Control soils obtained in this study may be as a result of varied amounts of nitrogenous compounds in the
283 abattoir impacted soils and the Control. The results also show that the abattoir soils recorded higher content of
284 Nitrogen than the Control soils. This finding agrees the report of [38] who also reported higher nitrogen content
285 for abattoir soils from Calabar, Cross River State than in the Control. Nitrogen content in abattoir soil obtained
286 in this study is lower than 0.48 – 3.01% reported by [19] and 0.18 - 0.65% reported by [38] for abattoir soils, but
287 higher than (0.008 -0.009%) reported by [67] for abattoir soils from Yola metropolis, Adamawa State, North
288 Eastern Nigeria. For the Control soil, the obtained range is lower than 0.08 – 0.09% reported by [38]. For
289 phosphorus, both abattoir soils recorded higher phosphorus content than the Control soils. This result also agrees
290 with the report of [38]. This could be explained by the higher pH (less acidity) values and higher organic matter
291 of the abattoir soils as indicated in Table 2. Range of phosphorus recorded in this study is in agreement with
292 2.46 – 3.61 mgkg^{-1} reported by [67], though lower than 0.005 – 0.007 mgkg^{-1} reported by [26] for abattoir soils
293 obtained from Sokoto State, Nigeria. Both nitrogen and phosphorus contents of abattoir soils obtained in this
294 study all fell within the permissible limits (nitrogen < 40% and phosphorus < 40 mgkg^{-1}) stipulated by [55] for
295 soils. Although, nitrogen and phosphorus are needed in soil by plants since the first is a building block of
296 protein, nucleic acid and other cellular constituents which are essential to all forms of life, the later is a
297 component of the complex nucleic acid structure of plants, which regulates protein synthesis in plants [68].
298 However, excess nitrogen and phosphorus in soil usually causes plants to mature too rapidly in addition to
299 reducing Zn, Cu and Fe availability in soil. Consequently, the moderate levels of nitrogen and phosphorus in the
abattoir soils under investigation is plausible in lieu of its usage for planting of crops [45].

300

301 Results for essential elements and trace metal levels of abattoir soils and Control soils are presented in
302 Table 4. The results indicated significant variations in the levels of all essential and trace metals investigated for
303 the abattoir and Control soils. Results for potassium (K) indicated the following ranges: 0.81 -0.93 Cmol/kg for
304 abattoir soils from IK, 0.68 – 0.74 Cmol/kg for EK and 0.17 – 0.26 Cmol/kg for the Control. The reason for this
305 variation may be as a result of the high moisture content of the abattoir soils as indicated in Table 3. Levels of
306 potassium in the Control soils were lower than those of the abattoir soils and this finding corroborates with the
307 report of [57] who also reported similar findings. Ranges of potassium obtained in this study for the abattoir
308 soils are consistent with the reports of [69] and [57] in their respective studies. Potassium has many different
roles in the soil relative to plants. It is involve in photosynthesis as it regulate the opening and closing of

309 stomata, regulate carbon(IV)oxide, triggers activation of enzymes and its essential for production of adenosine
 310 triphosphate (ATP) [64]. For other exchangeable bases (Sodium, calcium and magnesium), abattoir waste had
 311 significant influences on their levels in the studied abattoir soils.

312 From the results presented in Table 4, the abattoir soils recorded highest value of Na, Ca and Mg than
 313 in Control soils. In the case of Na, ranges were 0.42 – 0.51 mol/kg for samples from IK, and 0.53 -0.67 mol/kg
 314 for samples from EK. Ca and Mg recorded ranges of 1.83 – 2.04 mol/kg and 2.08 – 3.92 mol/kg for IK, while
 315 EK were 1.65 – 1.83 and 2.19 – 2.67 mol/kg. Ranges of Na obtained in this study for abattoir soils is higher than
 316 0.1 – 0.12 mol/kg reported by[38], but lower than 2.24 – 2.47 mol/kg reported by [67] for abattoir soils obtained
 317 from Yola Metropolis, Adamawa State, Nigeria. Ca and Mg ranges obtained for abattoir soils in this study are
 318 higher than 0.48 – 0.50 mol/kg Ca and 0.51 – 0.77 ml/kg Mg reported by [67] but lower than 12.6 – 15.6 mol/kg
 319 Ca and 4.06 – 9.80 mol/kg reported by [38]. The Control soil samples recorded lower levels of Na, Ca and Mg
 320 than the studied abattoir soils. These results agree with the reports of [57] and [38] although at variance with the
 321 report of [67], who reported lower Ca level in abattoir soils than in the Control soils. Several factors affect the
 322 levels of exchangeable bases in soil and this includes soil texture, organic matter content, CEC and moisture
 323 content of the soil [64]. Exchangeable bases such as Na, Ca and Mg are important in soil because they are
 324 involve in translocation of carbohydrates and nutrient within plants, cell growth, are component of chlorophyll
 325 for photosynthesis, protein synthesis and energy transfer within plants. Although, there are no limits to the
 326 amount of these bases in soil, higher levels of Na usually causes dispersion of fine particle of soils into pores
 327 thereby reducing water penetration and blocking plant root access. Higher levels of Ca in soil reduces uptake of
 328 other cation nutrients [57].

329 Table 4: Essential elements and trace metals levels of selected abattoir and Control soils

	Sampling Points					
	IK1	IK2	IKC	EK1	EK2	EKC
Essential elements						
K (mol/kg)	0.93	0.81	0.26	0.74	0.68	0.17
Na (mol/kg)	0.42	0.51	0.16	0.67	0.53	0.32
Ca (mol/kg)	1.83	2.04	1.23	1.65	1.83	0.98
Mg (mol/kg)	2.08	3.92	0.83	2.67	2.19	1.03
Trace metals						
Fe (mg/kg)	643.45	604.76	548.10	611.04	665.10	562.82
Zn (mg/kg)	19.23	21.05	15.09	24.13	18.56	12.15
Cd (mg/kg)	0.35	0.47	0.17	0.46	0.52	0.23
Cu (mg/kg)	16.82	14.94	4.94	20.92	16.30	3.56
Pb (mg/kg)	0.73	0.66	0.32	1.01	0.89	0.37
Cr (mg/kg)	0.32	0.26	0.05	0.18	0.22	0.08
Ni(mg/kg)	9.73	11.47	6.19	10.21	8.84	4.23

330 *IK1-Ikot Ekpene abattoir soil 1; IK2-Ikot Ekpene abattoir soil 2; IKC-Ikot Ekpene control soil; EK1-Eket
 331 abattoir soil 1; EK2 - Eket abattoir soil 2; EKC - Eket control soil

332 Results in Tables 4 indicate that, Fe varied between (604.76 – 643.45mgkg⁻¹) for IK abattoir soils,
 333 (611.04 – 665.10mgkg⁻¹) for EK abattoir soils, and (548.10 – 562.82mgkg⁻¹) for the Control soils. Levels of Fe
 334 obtained for both abattoir soils in this study is lower than 2569.00 – 4130.00mg/kg reported by Yahaya *et al* ³,
 335 but higher 59.36 – 81.70mgkg⁻¹ obtained by [70]. Also, from the results, abattoir soils recorded higher levels of
 336 Fe than the Control soils and this finding is consistent with the reports of [57]. This is indicative of additional
 337 source of Fe in studied abattoir wastes-impacted soils. The highest level of Fe was obtained at EK2 abattoir soil,
 338 while the lowest level was recorded in sample obtained from IK2. Results obtained in this study revealed direct
 339 relationship between activities at abattoir and Fe accumulation in studied abattoir soils. However; levels of Fe in
 340 both studied abattoir soils and Control are higher than 400.00mgkg⁻¹ recommended by [71] for Nigerian soils.
 341 This confirms that Nigerian soils have elevated levels of Fe as clearly shown by their reddish nature.
 342 Nevertheless; the availability of Fe in soil for plant uptake may not be guaranteed as Fe oxides (the major form
 343 of Fe in soil) are highly insoluble in soil [72].

344 Results obtained showed that concentrations of Zn in studied abattoir soils varied between (19.23 –
345 21.05mgkg⁻¹) for IK abattoir soils, (18.56 – 24.13mgkg⁻¹) for EK abattoir soils, and (12.15 – 15.09 mgkg⁻¹) for
346 the Control soils. Ranges of Zn obtained in this study are lower than 1.302 – 5.2362 mgkg⁻¹ reported by [5] in
347 abattoir soils, but lower than 50.91 – 92.50 mgkg⁻¹ obtained by [3] and 171.93 mgkg⁻¹ obtained by [40]. Levels
348 of Zn reported in studied abattoir soils were relatively higher than values obtained at the Control site. This is in
349 agreement with the report of [57] who also reported a higher level of Zn in abattoir soils than in Control soils.
350 However; the obtained ranges are lower than 140.0 mgkg⁻¹ limit by [73] for Nigerian soils. Nevertheless, since
351 toxicity of metal may not be identified by total metal concentration alone its availability may not be established.
352 Also, lower levels of Zn obtained in this study for Zn when compared to the permissible limit in soil is plausible
353 because Zn though very essential in soil, it is needed by plant in trace amount.

354 For Cd, results indicated the following ranges: (0.35 – 0.47mgkg⁻¹) for IK abattoir soils, (0.46 -
355 0.52mgkg⁻¹) for EK abattoir soils, and (0.17 – 0.23mgkg⁻¹) for Control soils. For the abattoir soils, Cd showed
356 highest abundance in abattoir soil from EK2 and least in abattoir soil from IK1, while Control soils recorded
357 lower levels of Cd than the abattoir soils studied. Ranges for Cd obtained in this study agrees with 0.43 – 0.71
358 mgkg⁻¹ obtained by [45] for abattoir soils from Obiaakpor Area, River State, Nigeria, but higher than 0.25
359 mgkg⁻¹ reported by [40]. However, levels of Cd obtained in this study were below the permissible limit (2.0 -3.0
360 mgkg⁻¹ stipulated for soil by[73]. This shows that studied abattoir soils were not overloaded with Cd even
361 though other parameters may need to be evaluated to really ascertain the status of the abattoir soils as it relates
362 to Cd.

363 Results in Tables 4 indicate ranges for Cu as 14.94 – 16.82 mgkg⁻¹, 16.30 – 20.92 mgkg⁻¹ and 3.56 –
364 4.94 mgkg⁻¹ for IK, EK abattoir soils and Control soils respectively. Ranges of Cu obtained for abattoir soils are
365 higher than 0.05 - 1.7 mgkg⁻¹ reported by [74] in abattoir soils within Umuahia, Nigeria but lower than 36.46 –
366 40.60 mgkg⁻¹ obtained by [58] in Abeokuta, Nigeria. The highest concentration of Cu was recorded in samples
367 from EK1 abattoir soil, while the lowest Cu level was also obtained in abattoir soil from IK1. Concentrations of
368 Cu in studied abattoir soils were higher than values obtained at the Control Soils. This indicated availability of
369 Cu-containing waste materials in studied abattoir waste-impacted soils. This is in agreement with the findings
370 by [45] in abattoir soils. The obtained Cu values are also lower than 36.0mgkg⁻¹ stipulated by [73] for Nigerian
371 soils. Nonetheless, bioavailability and toxicity of Cu could not be confirmed based on total concentration alone.

372 Results obtained for total Pb in studied abattoir soils indicated a range of 0.66 – 0.73 mgkg⁻¹ and 0.89 –
373 1.01 mgkg⁻¹ for IK and EK abattoir soils (Tables 4). Levels of Pb obtained in studied abattoir soils are lower
374 than 7.17 – 12.50 mgkg⁻¹ reported by [45] in abattoir soils within Obio Akpor, Port Harcourt, River State
375 Nigeria but higher than 0.18 – 0.83 mgkg⁻¹ obtained by [5] except for IK abattoir soils. Highest level of Pb was
376 reported in samples from EK1 while lowest Pb concentration was obtained in IK2 abattoir soil. Concentrations
377 of Pb in studied soils were higher than values obtained in the Control site revealing negative impact of abattoir
378 wastes on Pb levels in studied soils. However; levels of Pb obtained were below 85.00mgkg⁻¹ recommended by
379 [73] for soil in Nigeria.

380 Levels of Cr and Ni in studied abattoir soils (IK and EK) ranged from 0.26 – 0.32 mgkg⁻¹ and 0.18 -
381 0.22 mgkg⁻¹ for Cr and 9.73 – 11.47 mgkg⁻¹ and 8.84 – 10.21 mgkg⁻¹ for Ni (Tables 4). The ranges for Cr are
382 lower than 4.25 – 5.86 mgkg⁻¹ reported by [45] but higher than 0.0717 – 0.1358mgkg⁻¹ obtained by [5] in
383 abattoir soils. For Ni, Levels of Ni obtained are higher than 2.160 – 4.690 mgkg⁻¹ reported by [70] but lower
384 than 33.50 – 107.13mgkg⁻¹ recorded for Ni in abattoir soils by [3]. The highest Cr level was obtained in abattoir
385 soil from IK1, while the lowest concentration was in EK1 abattoir soil. For Ni, highest Ni level was obtained in
386 abattoir soil from IK2, while the lowest level was obtained in EK2 abattoir soil. Levels of Cr and Ni in abattoir
387 soils were higher than levels obtained in soil from the Control site and are in agreement with findings reported
388 by[45] in abattoir soils. However, values of Cr and Ni obtained in studied abattoir soils are much lower than
389 100.0 mgkg⁻¹ Cr and 35.0 mgkg⁻¹ Ni limits in soil by [73]. Nevertheless, pollution status of these metals may not
390 be ascertained using information from total concentration, but lower levels of Cr and Ni obtained in this study
391 are significant especially since abattoir soils studied are already used by farmers in planting crops.
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393 *Pollution status of trace metals in studied abattoir soils*

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To ascertain the pollution status of the studied abattoir soils, metal pollution index (MPI), degree of contamination (Cdeg), trace metals enrichment factor (EF), geo-accumulation index (Igeo) and metal pollution load (MPL) were evaluated. Metal pollution index (MPI) was used to differentiate between contamination and pollution levels in studied abattoir soils. It denotes the ratio between metal level in studied abattoir soils and reference value obtained in Control [75]. The different categories of MPI as indicated in Table 4 shows that Fe recorded MPI values ranging from 1.09 – 1.53 for all abattoir soils under investigation indicating slight pollution of the studied abattoir soils by Fe. Consequently, negative impact on soil, plants and the environment are predicted in and around studied abattoir soils. However, Fe is an essential element with very low bioavailability factor in studied soils hence; the effect may not be alarming. Results in Tables 4 also indicates that MPI values for Zn was in slight pollution (1- 2) category (Table 1), while Cd, Cu, Pb, Cr, and Ni were in moderate pollution (2.1 – 4.0) category except for abattoir soils from EK1 and EK2 that were in severe pollution (4.1 -8.0) category for Cu and IK1 and IK2 for Cr. This means that Zn, Cd, Cu, Pb, Cr and Ni have slightly polluted the studied abattoir soils and is expected to affect the soil, plants and the studied environment negatively.

Enrichment factors (EF) of metals were calculated for the abattoir soils using the continental crust average where Fe was used as reference element for normalization (Table 5). Fe exhibited an EF value of 1.00 in all the abattoir soils studied indicating that a greater proportion of Fe may have emanated from natural soil forming processes [34]. EF for Zn, Cd, Cu, Pb, Cr and Ni for all the abattoir soils studied were greater than 1.0 indicating that these trace metals are from anthropogenic source [34]. Results for the geo-accumulation index (Igeo) of trace metals in studied abattoir soils are presented in Tables 4. Results obtained showed the following ranges: 0.26 – 0.39, 0.40 – 0.55, 0.61 – 1.18, 0.41 – 0.56, 0.45 – 1.28 and 0.31 – 0.48 for Zn, Cd, Cu, Pb, Cr and Ni respectively for all the abattoir soils studied. From these results, the metals were in the 0 – 1 class (unpolluted – moderately polluted) following the classifications for geo-accumulation index by³⁶. Cu In abattoir soil from EK1 and Cr in abattoir soil from IK1 were in the 1 – 2 class (moderately polluted)

Degree of contamination (Cdeg) was determined to assess the extent of contamination of the four (4) studied abattoir soils and the results are presented in Figure 1. From the results, Cdeg values were 18.10, 17.38, 18.33 and 17.15 for IK1, IK2, EK1, and EK2 respectively. The varied Cdeg reported in this study by the abattoir soils may be attributed to the volume of abattoir wastes and abattoir activities in each of these sites. From the Cdeg results, it can therefore be deduced that the abattoir soils were considerably contaminated (16 < Cdeg < 32) based on the model predicted by [32].

Results obtained for pollution load index (PLI) for the four abattoir soils examined are indicated in Figure 1. PLI values were found to range from 2.17 for abattoir soil from IK1 to 2.33 for abattoir soil from EK1. Thus, PLI values of all the studied abattoir soils were within the heavy pollution (2 < PLI < 3) category according to [37]. These PLI values obtained in this study further confirm findings by degree of contamination of studied abattoir soils. This work has therefore revealed the negative impact of abattoir wastes on underlying soils with regards to metal accumulation.

Table 5: Metal pollution index (MPI), enrichment factor (EF) and geo –accumulation index (Igeo) of trace metals in studied abattoir soils

	Fe	Zn	Cd	Cu	Pb	Cr	Ni
MPI							
IK1	1.17	1.20	2.08	3.40	2.28	6.40	1.57
IK2	1.10	1.39	2.76	3.02	2.06	5.20	1.85
EK1	1.09	1.98	2.00	5.87	2.73	2.25	2.41
EK2	1.53	1.53	2.26	4.58	2.41	2.75	2.09
EF							
IK1	-	1.09	1.75	2.90	1.94	5.45	1.33
IK2	-	1.26	2.51	2.74	1.87	4.71	1.68
EK1	-	1.83	1.84	5.41	2.51	2.07	2.33
EK2	-	1.29	1.01	3.87	2.04	2.33	1.77

Igeo							
IK1	-	0.26	0.41	0.68	0.46	1.28	0.31
IK2	-	0.28	0.55	0.61	0.41	1.04	0.37
EK1	-	0.39	0.40	1.18	0.56	0.45	0.48
EK2	-	0.31	0.45	0.92	0.48	0.55	0.41

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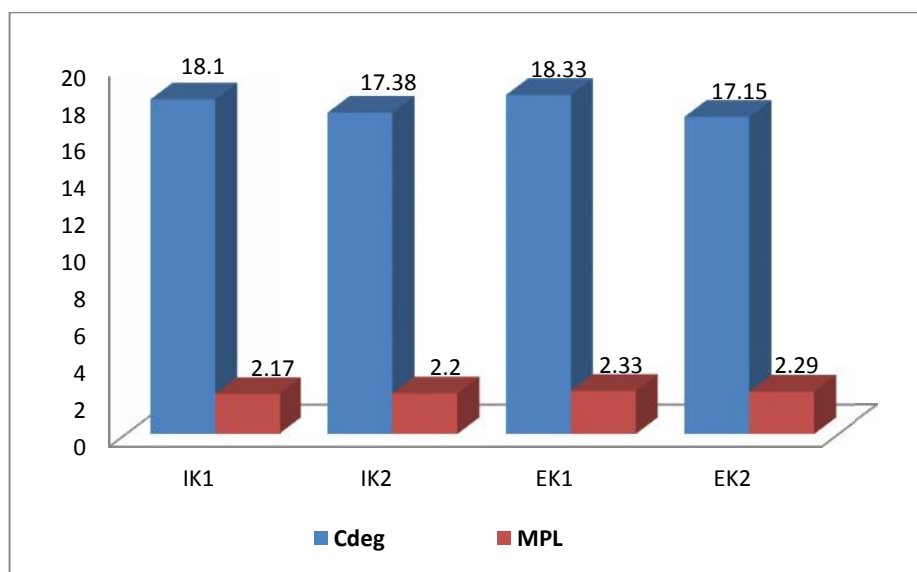
438 *MPI – Metal pollution index; EF – Enrichment factor; Igeo – Geo-accumulation index; IK1-Ikot Ekpene

439 abattoir soil 1; IK2-Ikot Ekpene abattoir soil 2; IKC-Ikot Ekpene control soil; EK1-Eket abattoir soil 1; EK2 -

440 Eket abattoir soil 2; EKC - Eket control soil

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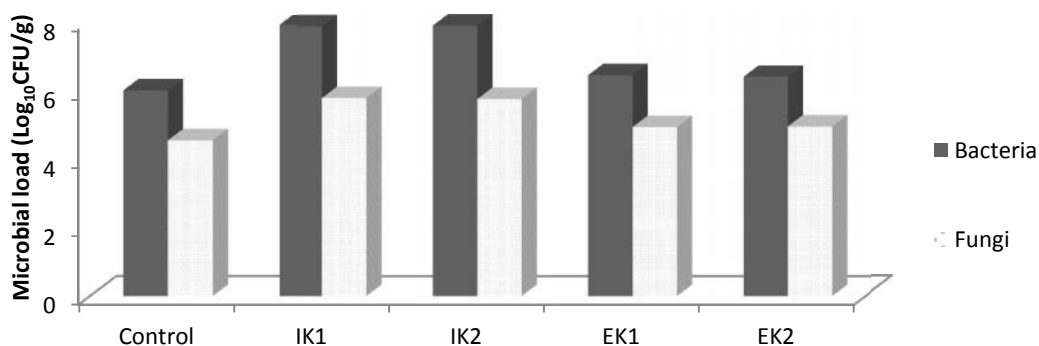
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445 Figure 1: Degree of contamination (Cdeg) and pollution load index (MPL) of trace metals in studied

446 abattoir soils

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Control= Uncontaminated Soil

IK1,IK2,EK1,EK2= Abattoir wastewater contaminated soil samples obtained from Ikot Ekpene(IK1,IK2) and Eket(EK1,EK2) region of Akwa Ibom State

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450 Figure 2: Microbial Load (CFU/g) of abattoir soils obtained from Ikot Ekpene and Eket
 451 region of Akwa Ibom State.

452 *Microbial loads of abattoir soils*

453 Figure 2 shows the microbial load (log₁₀CFU/g) of soil samples. The result shows the density of the culturable
 454 bacterial community present in 1g of samples obtained from the four (4) abattoir locations ranges from 6.41
 455 ±0.43 to 7.91 ±0.58 log₁₀CFU/g for total heterotrophic bacterial count and 4.94 ± 0.26 to 5.79 ± 0.3 log₁₀CFU/g
 456 for fungal count. Among the four (4) abattoir soils, IK2 had the highest heterotrophic bacterial densities of 7.91
 457 ± 0.58 log₁₀CFU/g while IK1 had the highest fungal count of 5.79 ±0.34 log₁₀CFU/g.

458 *Cultural and Biochemical Characteristics of microbial Isolates*

459 The cultural and biochemical characteristics of the microbial isolates are presented in Table 6 and 7. Six (6)
 460 species of bacteria were obtained from the soil samples using the aerobic culture techniques. These were
 461 *Klebsiella*, *Micrococcus*, *Pseudomonas*, *Bacillus*, *Escherichia* and *Enterobacter* species. While the two (2)
 462 fungal species isolated were *Aspergillus* and *Penicillium* species

Table 6: Morphological and Biochemical characteristics Bacterial of Isolates

Isolates	G.R	Shape	Motility	Catalase	Starch hydrolysis	Oxidase	Indole	Citrate	MR	VP	Glucose	Lactose	Mannitol	Probable Organism
1	-	R	-	+	-	-	+	+	-	+	AG	AG	-	<i>Klebsiella pneumoniae</i>
2	+	S	-	+	-	-	-	-	-	-	-	-	-	<i>Micrococcus luteus</i>
3	-	R	+	+	-	-	-	+	-	+	AG	AG	-	<i>Enterobacter cloacae</i>
4	+	R	-	+	+	-	+	+	-	-	-	-	-	<i>Bacillus polymyxa</i>
1.	+	R	-	+	+	-	-	-	-	-	-	-	-	<i>Bacillus subtilis</i>
2.	-	R	+	+	-	+	-	+	-	-	A	-	A	<i>Pseudomonas aeruginosa</i>
3.	-	R	+	+	-	-	+	-	+	-	AG	AG	-	<i>Escherichia coli</i>

Key: G.R = Gram Staining; + = Positive; - = Negative; R = Rod; S = Spherical; A = Acid only;
 AG = Acid and Gas produced

463

464 Table 7: Colonial and morphological characteristics of fungal (mould) isolates

Isolates	Colonial Morphology	Somatic cell type	Type of Hyphae	Asexual spores	Special Reproductive Structure	Conidia Head	Vesicle shape	Probable Fungi
1	Compact white with dark basal colour	Filamentous	Septate mycelium	Globose conidia	Conidiospore	Globose	Subglobose	<i>Aspergillus niger</i>
2	Whitish, yellowish to grey mycelium	Filamentous	Septate mycelium	Globose conidia	Conidiospore	Sub-globose to ellipsoidal	-	<i>Penicillium frequentans</i>

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468 *Occurrence and Distribution of microbial isolates within the samples and location*

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470 The distribution of microbial isolates within the soil samples is shown in Table 8 and 9. *Micrococcus luteus*,
471 *Pseudomonas aeruginosa* and *Bacillus subtilis* 5(100%) had the highest frequency of occurrence, while
472 *Enterobactercloacae* and *Klebsiellapneumoniae* had the least 2(40%). For the fungal isolates, *Aspergillusniger*
473 5(100%) had the highest frequency of occurrence, while *Penicillium frequentans* had the least 3(40%) frequency
474 of occurrence. Also, samples taken from Ikot Ekpene region were more contaminated than those taken from
475 Eket abattoir soils

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482 Table 8: Occurrence and distribution of bacterial isolates in abattoir and Control soil samples

Isolates	Sample Points					% Occurrence
	Control	IK1	IK2	EK1	EK2	
<i>Klebsiella pneumoniae</i>	-	+	+	-	-	2(40%)
<i>Micrococcus luteus</i>	+	+	+	+	+	5(100%)
<i>Enterobacter cloacae</i>	-	+	+	-	-	2(40%)
<i>Pseudomonas aeruginosa</i>	+	+	+	+	+	5(100%)
<i>Escherichia coli</i>	-	+	+	+	+	4(80%)
<i>Bacillus polymyxa</i>	+	+	+	-	-	3(60%)
<i>Bacillus subtilis</i>	+	+	+	+	+	5(100%)

483

484 *Key: + = Present; - = Absent

485 Table 9: Occurrence and distribution of fungal isolates in abattoir and Control soil samples

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Isolates	Sample Points					% Occurrence
	Control	IK1	IK2	EK1	EK2	
<i>Aspergillusniger</i>	+	+	+	+	+	5(100%)
<i>Penicilliumfrequentans</i>	+	+	+	-	-	3(60%)

487

488 *Key: + = Present; - = Absent

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490 The significant ($p=0.05$) increase in microbial loads encountered in the contaminated soil against that
 491 of the control soil is not surprising and it could be directly linked to the impacted abattoir wastes disposed on the
 492 studied land. This is because abattoir waste may contain many growth factors that could serve as an easily
 493 utilizable source of nutrient and encourage rapid multiplication by microorganisms. This result is in agreement
 494 with the report of [56] and [26] who independently reported similar increase in microbial load of soil samples
 495 contaminated with abattoir effluents. Eket (EK1 and EK2) abattoir soils were found to harbor less number and
 496 species of microorganisms. This could be because the region is well known for oil exploration activities and is
 497 expected to harbor more of microorganisms that could survive in soil contaminated with hydrocarbon. The
 498 presence and abundance of species of *Bacillus* and *Micrococcus*, *Aspergillus* and *Penicillium* observed in both
 499 the abattoir and Control soils is not surprising as these organisms are indigenous to soil environment and are
 500 known to persist and thrive especially during carbon and nitrogen sources influx in the soil [76]. However,
 501 presence of *Pseudomonas aeruginosa*, *E. coli*, *Enterobacter cloacae* and *Klebsiella pneumoniae* in the abattoir
 502 soil may be attributable to the high load of animal excreta in the abattoir wastes since these microorganisms are
 503 well known flora of fresh beef. Their presence in the abattoir soil samples is indicative of recent faecal pollution
 504 as they are mostly indicator organisms. Similar findings were reported by [76]. Most of the fungal isolates were
 505 also soil-inhabiting microorganisms as well as common spoilage organisms associated with beef industry [76-
 506 77]. The presence of these organisms is a pointer to possible pollution and may have an effect on the soil
 507 ecological balance.

508 Conclusion

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510 The results of this study have shown the physicochemical characteristics, total essential and trace metal
 511 levels, total heterotrophic bacterial and fungal loads of abattoir and Control soils from Ikot Ekpene and Eket
 512 Local Government Areas of Akwa Ibom State, Nigeria. Metal pollution index (MPI), enrichment factor (EF),
 513 geo-accumulation index (Igeo), degree of contamination (Cdeg) and pollution load index (PLI) of trace metals
 514 have also been calculated using empirical pollution models. Essential elements and trace metal levels were
 515 higher in abattoir soils than in Control though were within permissible limit in soil except for Fe. Also,
 516 microbial results revealed a significant increase in the number and varieties of microorganisms most of which
 517 may be pathogenic, but are more often than not indicators of recent faecal pollution in the soil impacted with
 518 abattoir wastes. This study, therefore, concludes that soil impacted with abattoir wastes is richer in plant
 519 nutrients and can be exploited for growing of crops. But it is advised that routine checks be conducted to
 520 forestall trace metals accumulation above safe levels in these soils.

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522 References

523

- 524 1. Nwuche, C. O., and Ugorji, E. O. (2010). Effect of co-existing plant species on soil
 525 microbial activity under heavy metal stress. *International Journal of Environmental
 526 Science and Technology*, 7: 697 – 704.
- 527 2. Daniel, L. A. and Braide, S. A. (2004). The effect of oil spillage on a cultivated wet-land of
 528 the Niger Delta. *Journal of Environmental Science*, 2: 153 – 158
- 529 3. Yahaya, M.I., Mohammad, S. and Abdullahi, B. K. (2009). Seasonal variations of heavy
 530 metals concentration in abattoir dumping site soil in Nigeria. *J. Appl. Sci. Environ. Manage.*,
 531 13(4): 9 – 13.
- 532 4. Hornby, A. S. (2006). *Oxford Advanced Learner's Dictionary of current English*, 7th edition, p. 1381
- 533 5. Ubwa, S. T., Atoo, G. H., Offem, J. O., Abah, J. and Asemave, K. (2013). Effect of Activities at the
 534 Gboko Abattoir on Some Physical Properties and Heavy Metals Levels of Surrounding Soil.
 535 *International Journal of Chemistry*, 5(1): 49 – 57.
- 536 6. Red Meat Abattoir Association (2010). Waste Management-Red Meat Abattoir. Retrieved
 537 from SON (2003). Standard Organization of Nigeria Safe Drinking Water Regulation.
 538
- 539
- 540

- 541 [http://www.docstoc.com/docs/103302144/Waste-Management- %EE%9F% A6-Red-Meat-](http://www.docstoc.com/docs/103302144/Waste-Management-%E9%A6-Red-Meat-)
542 Abattoirs.
543
- 544 7. Steffen, Roberts, Kirsten Inc. (1989). Water and waste-water management in the red meat industry (P.
545 36).WRC Report No. 145 TT41/89. WRC, Pretoria.
546
- 547 8. Kruslin E, Hodel CM, Schurgast H (1999). Progress in diagnosis of Chronic Toxic metal
548 Poisoning by Hair analysis. *Toxicol. Lett.* 88:84.
549
- 550 9. Jukna, C., Jukna, V. and Suigzdaite, (2006). Determination of Heavy metals in Viscera and
551 Muscles of cattle. *Bulg. J. Vet. Med.*, 9(1):35 – 41.
552
- 553 10. Ezeoha, S. L. and Ugwuishiwu, B. O. (2011). Status of Abattoir Wastes Research in Nigeria. *Nigerian*
554 *Journal of Technology*, 30(2): 143-148.
555
- 556 11. Nwachukwu, M. I., Akinde, S. B., Udujih, O. S. and Nwachukwu, I. O. (2011). Effect of abattoir
557 wastes on the population of proteolytic and lipolytic bacteria in a Recipient Water Body (Otamiri
558 River). *Global Research Journal of Science*, 1: 40 - 42.
559
- 560 12. Magaji, J. Y. and Chup, C.D. (2012). The effects of abattoir waste on water quality in Gwagwalada-
561 Abuja, Nigeria. *Ethiopian Journal of Environmental Studies and Management*, 54 (Suppl.2): 542 -549.
562
- 563 13. Raymond, C.L. (1977), *Pollution Control for Agriculture*. New York: Academic Press Inc.
564
- 565 14. Mohammed, S. and Musa, J. J. (2012). Impact of Abattoir Effluent on River Landzu, Bida, Nigeria.
566 *Journal of Chemical, Biological and Physical Sciences*, 2(1), 132-136.
567
- 568 15. Chen, T., Liu, X., Li, X., Zhao, K., Zhang, J., Xu, J., Shi, J. and Dahlgren, R. A. (2009). Heavy metal
569 sources identification and sampling uncertainty analysis in a field-scale vegetable soil of Hang Zhou,
570 China. *Environmental Pollution*, 157: 1003–1010.
571
- 572 16. Ibitoye, A. A. (2006). Laboratory manual on basic soil analysis. Foadave Nigeria Limited, 16
573 -36.
574
- 575 17. Olofunmi, O. E., and Alli, A. A. (2016). To determine variability of soil physical and
576 chemical properties of a typical land disposal catfish effluent in South Western Nigeria.
577 *International Journal of Agricultural Science*, 6 (3): 963 – 968
578
- 579 18. Van-Reeuwijk, L. P. (1993). *Procedures for soil analysis*. Wageningen: Technical Paper 9,
580 ISRIC; pp. 16–18.
- 581 19. Edori, O. S., and Iyama, W. A. (2017). Assessment of physicochemical parameters of soils
582 from selected abattoirs in Port Harcourt, Rivers State, Nigeria. *Journal of Environmental*
583 *Analytical Chemistry*, 4 (2): 194 -201.
- 584 20. Ebong, G. A., Offiong, O. E. and Ekpo, B. O. (2014). Seasonal variations in trace metal levels,
585 speciation and physicochemical determinants of metal availability in dumpsite soils within Akwa Ibom
586 State, Nigeria. *Chemistry and Ecology*, 2014: 1- 15.
587
- 588 21. Al-Khashman, O. A. and Shawabkeh, R. A. (2006). Metals distribution in soils around the cement
589 factory in southern Jordan. *Environmental Pollution*, 140: 387–394.
590
- 591 22. Fagbote, E. O. and Olanipekun, E. O. (2010). Speciation of heavy metals in sediment of Agbabu
592 Bitumen deposit area, Nigeria. *Journal of Applied Science and Environmental Management*, 14 (4): 47
593 – 51.
594

- 595 23. Khan, R. S., Kumar, N. J., and Patel, G. H (2013). Physicochemical properties, heavy metals
596 content and fungal characterization of an old gasoline contaminated soil site in Anand,
597 Gujarat, India. *Environ Exp Biol*, 11: 137 -143.
598
- 599 24. Okoye, C. O. B., and Agbo, K. E. (2011). Dispersion pattern of trace metals in soils
600 surrounding solid waste dump in Nsukka. *J. Chem Soc Nigeria*, 36: 112 – 119.
- 601 25. Allen, S. E., Grinshaw, H. W., Pancinson, J.A., and Quarmby, L. (1974). Chemical methods
602 of analyzing ecological materials. Blackwell Scientific Publications, Oxford, London, UK.
- 603 26. Rabah, A. B., Oyeleke, S. B., Manga, S. B., Hassan, L. G. and Ijah, U. J. (2010).
604 Microbiological and Physico-Chemical Assessment of Soil Contaminated with
605 Abattoir Effluents in Sokoto Metropolis, Nigeria. *Science World Journal*, 5(3): 1-4.
606
- 607 27. Cappuccino, J. G. and Sherman, N. (2002). Techniques for Isolation of Pure Culture. In:
608 *Microbiology: A Laboratory Manual*. 6th ed. Singapore : Pearson Education, Inc., p.544
609
- 610 28. Cheesbrough, M. (2006). District Laboratory Practice in Tropical Countries Part 2. UK:
611 Cambridge University Press, pp. 45-70.
612
- 613 29. Brenner, D. J.; Noel, R. K. and Staley, J. T. (1982). *Bergey's Manual of Systematic*
614 *Bacteriology*. 2nd ed., New York: Springer, p. 304.
615
- 616 30. Aneja, K. R. (2003). Experiments in Microbiology, Plant Pathology and Biotechnology. 4th
617 Ed., New Delhi: New age International Limited Publishers, p. 282.
618
- 619 31. Lacatusu, R. (2000). Appraising levels of soil contamination and pollution with heavy metals in:
620 Heinike H.J., Eckselman W., Thomasson, A.J., Jones, R.J.A, Montanarella L and Buckeley B (eds.).
621 *Land information systems for planning the sustainable use of land resources*. European Soil Bureau
622 Research Report No. 4. Office of Official Publication of the European Communities, Luxembourg, pp.
623 393 – 402.
624
- 625 32. Hakanson, L. (1980). Ecological Risk Index for Aquatic Pollution Control, a Sedimentological
626 Approach. *Water Research*, 14 (8): 975-1001.
627
- 628 33. Rubio, B., Nombela, M. A. and Vilas, F. (2000). Geochemistry of major and trace elements in
629 sediments of the Ria de Vigo (NW Spain) an assessment of metal pollution, *Marine Pollution Bulletin*,
630 40 (11): 968 – 980.
631
- 632 34. Zsefer, P., Glasby, G. P., Sefer, K., Pempkowiak, J., and Kaliszan, R. (1996). Heavy-metal
633 pollution in superficial soils from the southern Baltic Sea off Poland. *Journal of*
634 *Environmental Science & Health*, 31A, 2723–2754.
635
- 636 35. Lu, X., Wang, L., Lei, K., Huang, J. and Zhai, Y. (2009). Contamination assessment of copper, lead,
637 zinc, manganese and nickel in street dust of Baoji, NW China, *Journal of Hazardous Materials*, 161:
638 1058-1062.
639
- 640 36. Huu, H. H., Rudy, S and Damme, A. V. (2010). Distribution and contamination status of heavy metals
641 in estuarine sediments near Cau Ong Harbor, Ha Long Bay, Vietnam. *Geologica Belgica*, 13 (1-2): 36
642 – 47.
- 643
- 644 37. Tomilson, D. C., Wilson, J. G., Harris, C. R. and Jeffrey, D. W. (1980). Problems in assessment of
645 heavy metals in 20 estuaries and formation of pollution index. *Helgol Meeresanlter*, 33: 566-575.
646

- 647 38. Ediene, V. F., Iren, O. B., and Idiong, M. M (2016).Effect of abattoir effluents on the
648 physicochemical properties of surrounding soils in Calabar Metropolis. *International Journal*
649 *of Advance Research*, 4 (8): 37 – 41.
650
- 651 39. Brady, N. C., and Well, R. R. (1999). The nature and properties of soil. 12th edition Delhi,
652 Pearson Educational Publishers: p.213
653
- 654 40. Olayinka, O. O., Akande, O. O., Bamgbose, K., and Adetunji, M. T. (2017). Physicochemical
655 characteristics and heavy metal levels in soil samples obtained from selected anthropogenic
656 sites in Abeokuta, Nigeria. *J. Appl. Sci. Environ. Manage.*, 21(5): 883 – 891.
657
- 658 41. Chibuzor. O. J., Nwakonobi, T. U., and Itodo, I. N. (2017). Influence of physic-chemical characteristic
659 of soils on heavy metal contamination in Makurdi, Benue State. *International Journal of Environmental*
660 *Science, Toxicology and Food Technology*, 11: 84 -92
- 661 42. Franzmeier DP, Lemme GD, Miles RJ (1995). Organic Carbon in soils of North Central United States Soil
662 Science Society America Journal. 49: 702 – 708. Rich, K. and Von, I. 2010. Utah State University, USA,
663 Extension and Agriculture Department: Guidelines for Soil Quality, pp. 60.
664
- 665 43. FAO. (1979). Soil Survey investigation for irrigation soils. New York: John Wiley and sons Inc.
666
- 667 44. Arias, M. E., Gonzalez-Perez, J. A.,and Ball, A.S (2005). Soil health: A new challenge for
668 microbiologist and chemists. *Int Microbiol.* 8: 13 -21
669
- 670 45. Chukwu, U. J. and Anuchi, S.O. (2016). Impact of Abattoir Wastes on the Physicochemical Properties
671 of Soils within Port Harcourt Metropolis. *The International Journal of Engineering and Science*, 5(6):
672 17 -21.
673
- 674 46. Agbaire, P. O. and Emoyan, O. O. (2012). Bioaccumulation of heavy metals by earthworm (*Lumbricus*
675 *terrestris*) and associated soils in domestic dumbsites in Abraka, Delta State, Nigeria. *International*
676 *Journal of Plant, Animal and Environmental Sciences*, 2(3): 210-217
677
- 678 47. Odu, C.T.I., Nwoboshi, L.C. and Esuruoso, O.F. (1985) Environmental studies (soil and
679 vegetation) of the Nigerian Agip oil company operation areas. In: Proceedings of an
680 international seminar on petroleum industry and the Nigerian environment, pp. 274–283.
681 NNPC, Lagos Nigeria.
682
- 683 48. Thomson, C. J., Marschner, H., and Rombeld, V. (1993). Effect of nitrogen fertilizer form on
684 pH of the bulk soil and rhizosphere, and on the growth, phosphorus, and micronutrients
685 uptake of bean. *Journal of Plant Nutrition*, 16 (3): 493-506
686
- 687 49. Osakia, M., Watanabe, T., and Tadano, T. (1997). Beneficial effect of aluminium on growth
688 of plants adapted to low pH soil. *Soil Science and Plant Nutrition*, 43 (3): 551 – 563
689
- 690 50. Osakwe, S. A (2016). Contribution of abattoir activities in Delta State, Nigeria, to the soil
691 properties of their surrounding environment. *J Chem Biol Phys Sci* 6: 982 – 991.
692
- 693 51. Akan, J. C., Abdulrahman, F. I., Sodipo, O. A. & Lange, A. G. (2010). Physicochemical parameters in
694 soil and vegetable samples from Gongulon agricultural site, Maiduguri, Borno State, Nigeria. *Journal*
695 *of American Science*, 6 (12): 75 – 87.
696
- 697 52. Onweremadu, E. U. (2008). Physico-chemical charaterisation of a farmland affected by wastewater in
698 relation to heavy metals. *Journal of Zhejiang University Science A.*, 9 (3): 366-372.

- 700 53. Chaudhari, P. R., Ahire, D. V. & Ahire, V. D. (2012). Correlation between physico-chemical properties
701 and available nutrients in sandy loam soils of Haridwar. *Journal of Chemical, Biological and Physical*
702 *Sciences*, 2 (3): 1493-1500.
703
- 704 54. Nafarnda, W. D., Ajayi, I. E., Shawulu, J. C. Kawe, J. C., Omeiza, O. K., Sani, N. A., Tenuche, O. Z.
705 Dantong, D. D., and Tags, S. Z. (2006). Bacteriological quality of Abattoir effluents discharged into
706 water bodies in Abuja, Nigeria. *ISRN Vet. Sci.*, article ID515689.
707
- 708 55. WHO/FAO/IAEA. World Health Organization. Switzerland: Geneva. (1996). Trace Elements in
709 Human Nutrition and Health.
710
- 711 56. Adesemoye, A. O., Opere, B. O. and Makinde, S. C. O. (2006). Microbial Content of abattoir waste
712 water and its contaminated soil in Lagos, Nigeria. *African Journal of Biotechnology*, 5(20): 1963-
713 1968.
714
- 715 57. Neboh, H. A., Iiusanya, O. A., Ezekoye, C. C., and Orji, F. A. (2013). Assessment of Ijebu-Igbo
716 abattoir effluent and its impact on the ecology of the receiving soil and river. *IOSR Journal of*
717 *Environmental Science and Food Technology*, 7(5): 61-67.
718
- 719 58. Owagboriaye, F. O., Dedeke, G. A. and Ademolu, K. O. (2016). Glutathione-S-transferase production
720 in earthworm (Annelida: Eudrilidae) as a tool for heavy metal pollution assessment in abattoir soil.
721 *Biol. Trop. (Int. J. Trop. Biol.)*, 64(2): 779-789.
722
- 723 59. Oyedele, D. J., Gasu, M. B., Awotoye, O. O. (2008). Changes in soil properties and plant
724 uptake of heavy metals on selected municipal solid waste dumpsites in Ibe-Ife, Nigeria.
725 *African Journal of Environmental Science and Technology*, 3 (5): 107 – 115.
726
- 727 60. Allorge, D. O. (1992). Abattoir design management and effluent disposal in Nigera.
728 University of Ibadan Press. p. 254
729
- 730 61. Okoro, D., Oviasogie, P. O., and Oviasogie, F. E. (2011). Soil quality assessment 33 months
731 after crude oil spillage and clean-up. *Chemical Speciation and Bioavailability*, 23: 1-6
732 DOI:10.3184/095422911X12963991543492.
733
- 734 62. Iwegbue, C. M. A., Egobueze, F. and Opuene, K. (2006). Preliminary assessment of heavy metals
735 levels of soils of an oil field in the Niger Delta. Nigeria. *International Journal of Environmental*
736 *Science and Technology*, 3 (2): 167-172.
737
- 738 63. Hazelton, P. A. and Murphy, B. W (2007). Interpreting soil test results: What do all the
739 numbers mean?. CSIRO Publishing: Melbourne
740
- 741 64. Rayment, G. E., and Higginson, F. R. (1992). Cation Exchange Capacity in Australian
742 Laboratory Handbook of soil and water chemical methods . Inkata Press: Melbourne.
743
- 744 65. Osu, C. I. and Okereke, V. C. (2015). Heavy metal accumulation from abattoir wastes on soils and
745 some edible vegetables in selected areas in Umuahia metropolis. *International Journal of Current*
746 *Microbiology and Applied Sciences*, 4(6): 1127 – 1132
747
- 748 66. Onianwa, P. C., and Essien (1999). Petroleum hydrocarbon levels in sediment of stream and
749 river within Ibadan City, Nigeria. *Bull Chem Soc Ethiopia*, 13: 82 – 85.
750
- 751 67. Abubakar, G. A. (2014). Impact of abattoir effluent on soil chemical properties in Yola,
752 Adamawa State, Nigeria. *International Journal of Sustainable Agricultural Research*, 1: 100-
753 107
754
- 755 68. Holland, M. D. G., Barton, A. D., and Morph, S. T. (1989). Land evaluation for agricultural
756 recommendations for Cross River National Park. Oban Division. Prepared by Odwki in
757 Collaboration with INNFF.

- 758
759
760
761
762
763
764
765
766
767
768
69. Sumayya, B. U., Usman, B. U., Aisha, U., Shahida, A., Mohammad, A., Yakubu, M. S. and Zainab, M. (2013). Determination of Physiochemical Qualities of Abattoir Effluent on Soil and Water in Gandu, Sokoto State. *IOSR Journal of Environmental Science, Toxicology and Food Technology*, 4(4): 47-50.
70. Simeon, E. O. and Friday, K. (2017). Index Models Assessment of Heavy Metal Pollution in Soils within Selected Abattoirs in Port Harcourt, Rivers State, Nigeria. *Singapore Journal of Scientific Research*, 7: 9 – 15.
71. FEPA(1997). *Guidelines and standard for environmental impact assessment in Nigeria*. pp. 87– 95.
- 769
770
771
772
773
774
775
776
777
778
779
780
781
782
783
784
785
786
787
788
789
790
72. Federal Environmental Protection Agency (FEPA), (1999). National Guidelines and Standards for Soil Quality in Nigeria. FEPA, Rivers State Ministry of Environment and Natural Resources, Port Harcourt.
73. Shittu, O. S. and Ogunwale, J. A. (2010). Total and Extractable Iron and Manganese in Soils Developed on Charnockite in Ekiti State, Nigeria. *International Journal of Water and Soil Resources Research*, 1(1–3): 83 – 93.
74. Osu, C. I. and Okereke, V. C. (2015). Heavy metal accumulation from abattoir wastes on soils and some edible vegetables in selected areas in Umuahia metropolis. *International Journal of Current Microbiology and Applied Sciences*, 4(6): 1127 – 1132
75. Hong, A. H., Ling, L. P. and Selaman, O. S. (2014). Environmental Burden of Heavy Metal Contamination Levels in Soil from Sewage Irrigation Area of Geriyo Catchment, Nigeria. *Civil and Environmental Research*, 6(10): 118 – 124.
76. Atlas, R. M., and Bartha, R. (2007). *Microbial Ecology: Fundamentals and Applications*, Benjamin/Cummings Publishing Company Inc, India. pp. 34 - 39
77. Alonge, D. O. (1991). *Textbook of Meat Hygiene in the Tropics*. Farm Coe Press, Ibadan, Nigeria