

Previously Unacknowledged Potential Factors in Catastrophic Bee and Insect Die-off Arising from Coal Fly Ash Geoengineering

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ABSTRACT

Aims: We investigate previously unacknowledged potentially major contributory factors in global catastrophic bee and insect die-off that arise from the use of aerosolized coal fly ash (CFA) for covert weather and climate manipulation. We also present forensic evidence that CFA is the primary material used in atmospheric aerosol geoengineering operations.

Methods: We conducted extensive literature research and additionally utilised inductively coupled plasma mass spectrometry.

Results: The primary components of CFA, silicon, aluminium, and iron, consisting in part of magnetite (Fe_3O_4), all have important potential toxicities to insects. Many of the trace elements in CFA are injurious to insects; several of them (e.g., arsenic, mercury, and cadmium) are used as insecticides. Toxic particulates and heavy metals in CFA contaminate air, water, and soil and thus impact the entire biosphere. Components of CFA, including aluminium extractable in a chemically-mobile form, have been shown to adversely affect insects in terrestrial, aquatic, and aerial environments. Both the primary and trace elements in CFA have been found on, in, and around insects and the plants they feed on in polluted regions around the world. Magnetite from CFA may potentially disrupt insect magnetoreception. Chlorine and certain other constituents of aerosolized CFA potentially destroy atmospheric ozone thus exposing insects to elevated mutagenicity and lethality levels of UV-B and UV-C solar radiation.

Conclusions: It is necessary to expose and halt atmospheric aerosol geoengineering to prevent further gross contamination of the biosphere. As insect populations decline, bird populations will decline, and ultimately so will animal populations, including humans. The gradual return of insects when the aerial spraying is stopped will be the best evidence that aerosolized CFA is, in fact, a leading cause of the current drastic decline in insect population and diversity.

Keywords: Insect die-off; biodiversity; geoengineering; coal fly ash; aluminium toxicity; colony collapse; magnetite.

1. INTRODUCTION

There is public awareness and concern [1] about the population decline of the Western honey bee,

Apis mellifera, the principal agricultural pollinator worldwide [2]. Bumble bee populations (*Bombus* sp.), secondary but nevertheless important pollinators, are also in decline in North America and Europe [3-5]. Evans et al. [6] investigated 61

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quantified variables, such as pesticide levels and pathogen loads in *Apis mellifera* and reported: "no single measure emerged as a most-likely cause of colony collapse disorder". As noted by Watanabe [7] there is "no smoking gun."

A recent study documented the alarming decline, 75% reduction, in insect populations (biomass) in protected areas of Germany just over the past three decades [8]. This dramatic loss of insect abundance and diversity has profound ramifications for the world-wide food web and ecosystems. In that study, neither climate change nor land use could be linked to this frightening decrease in insects, although agricultural practices and pesticide use could not be excluded as contributing factors. Like Western honey bee decline, there is no readily identifiable cause, no 'smoking gun'.

Biodiversity declines have been reported elsewhere in other species. For example, Brooks et al. [9] in the UK reported over a 15 year period that three-quarters of the carabid beetle species investigated had declined substantially. Similar declines were reported for British common macro-moths [10] and butterflies [11]. In the last 40 years, there has been a 45% decline in invertebrates, a decline that includes all of the major insect Orders [12]. No readily identifiable cause of these declines has emerged.

These investigations clearly implicate a large-scale cause of insect die-off, and point to an

urgent need to discover the actual underlying cause(s) of this insect decline. However, it is presumed that deliberately aerosolized coal fly ash (CFA), a global and toxic by-product of coal combustion, potentially represents a major contributor to the worldwide die-off of insects.

When coal is burned, primarily by electric utilities, the heavy ash settles, while the light ash, CFA, formed in the gases above the burner, would exit smokestacks if not trapped and sequestered as required by modern regulations. Coal fly ash is one of the largest industrial waste-product streams throughout the world. Disposal of CFA is problematic; it is often simply dumped into surface impoundments or placed into landfills which cause concerns for ground water contamination and environmental pollution [13,14]. However, in many countries including the United States, a significant percentage of coal fly ash is recycled into the structural fill and such products as concrete [15]. Coal fly ash is also utilised in soil additives and fertiliser [16].

Reports are available to show that CFA is consistent with its use as the primary material aerosolized for covert, jet-emplaced climate manipulation operations (Fig. 1) [17,18]. CFA forms as particles ranging from <0.1 μm to 50 μm in width and therefore requires little further processing for use as a climate-altering aerosol. Sprayed into the atmosphere, these particles reflect some sunlight, but they also absorb energy which is transferred to the atmosphere via molecular collisions. The particles also block



Fig. 1. Jet-emplaced weather/climate manipulation particulate trails. (Photographers with permission) Clockwise from upper left: Karnak, Egypt (author JMH); London, England (Ian Baldwin); Geneva, Switzerland (Beatrice Wright); Chattanooga, TN, USA (David Tulis); San Diego, CA, USA (author JMH); Jaipur, India (author JMH)

heat from leaving Earth's surface. The aerosolized particles inhibit rainfall by keeping water droplets from coalescing to fall as rain; the effect is to cause drought, but eventually, the atmosphere becomes so burdened with moisture that storms occur with rain falling in deluges. This covert aerial spraying worsens global warming and totally disrupts natural weather patterns [19].

In the present investigation, efforts are made to describe and provide evidence that aerosolized CFA yields toxic elements that contaminate the environment and potentially become major contributors of insect die-offs. These include, specifically the consequences of toxins extractions from CFA into rainwater, and the effects of CFA particulate-components on insect viability. Further, the harmful consequences of

enhanced UV-B and UV-C solar radiation that concomitantly arise from atmospheric ozone reduction by aerosolized CFA are discussed.

2. METHODS

In the face of the obvious aerial particulate spraying, there is, however, a concerted effort to deceive the public and the scientific community of its existence and its adverse consequences on human and environmental health [20]. For the following reasons, CFA is a likely material for use in global-scale geoengineering operations: (1) It is a major industrial waste product; (2) It is produced in the size needed without much additional processing; and, (3) Its production

facilities are in place, out of sight, and utilize railroad transport.

The methods for demonstrating that the aerosolized particulates are consistent with CFA are twofold: (1) Showing that the relative amounts of elements dissolved in rainwater are similar the relative amounts of elements of CFA extracted into water during laboratory leach studies [21]; and, (2) Showing that the relative amounts of elements brought down by snow, in a manner analogous to the technique of co-precipitation [17], are similar to the relative amounts of elements found in CFA [21]. Measurements, previously published and newly presented here, are by inductively coupled plasma mass spectrometry.

3. RESULTS AND DISCUSSION

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Since at least the beginning of the 21st century and even before, numerous citizens from around the world have witnessed aerial spraying of particulate trails across the sky [24]. Without reliable information available about chemical composition and potential health-risks of the aerosol substance being sprayed, concerned citizens took post-spraying rainwater samples to commercial laboratories for analyses. Typically, only aluminium analysis was requested; sometimes barium was also requested and, occasionally, strontium as well. When the aerial spraying became an obvious near-daily activity in San Diego (USA), one of us (JMH) began a series of investigations to ascertain the composition of the aerosolized particles. Standard protocols for certified laboratory water analyses require filtration to remove particulate matter before measurements; thus it is evident that the rainwater had leached those three elements from some readily leachable particulate matter before it fell to the ground.

By expressing data as ratios to some common element, such as barium, provides a means to eliminate the consequences of various amounts of dilution. Comparison of those analytical results, expressed as ratios relative to barium, to corresponding experimental water-leach analyses of a likely aerosolise-substance, coal fly ash (CFA), provided the first scientific evidence that CFA is the main particulate-pollutant substance used for ongoing tropospheric geoengineering [22].

To understand the chemical process involved, consider by analogy the hypothetical example of

finely powdered tea leaves being sprayed into the troposphere. Atmospheric moisture would “brew” the tea, extract tannin and other chemicals, which would come down as rain, with chemical signatures of tea; the rain would be tea, albeit very weak tea. Coal fly ash (CFA) forms principally by condensation in the hot combustion gases in the flue above the combustion chamber of coal-powered electric utilities in circumstances, unlike those typically encountered in nature, and consists of a disequilibrium assemblage of typically anhydrous matter [23]. Water is capable of quickly extracting numerous toxic elements from CFA [22]. When CFA is sprayed into the troposphere, atmospheric water extracts numerous toxic elements by leaching, which are brought down dissolved in rainwater and provide a chemical signature of the CFA. The more elements measured in rainwater, the more precise and unique the signature becomes. This is a significant signature as common windblown sands and soils are not readily and quickly leached by rainwater.

Fig. 2 shows a comparison of rainwater analyses with ranges of CFA laboratory leach data. Except for the Bangor, Maine (USA) data, the remainder of the data has been published and is reproduced with permission [18]. Dilution is a variable factor that can be compensated in analytical comparisons by using ratios. Dilution, however, in many instances causes the less abundant elements to be below the detection limits for commercial analytical laboratories. The Bangor, Maine (USA) data, shown in Fig. 2, is particularly significant as the dilution factor was low and important trace element analyses as requested were able to be determined.

From [18] with permission. Rainwater data from 2011 in Bangor, ME, USA, courtesy of Russ Tanner, is newly added.

Fig. 3, reproduced with permission from [18], shows analyses of aerosolized particulates brought down by snow, the residue from evaporation and the residue trapped upon underlying snow mold as the snow melted, compared with the range of corresponding CFA analyses. This figure and Fig. 2 demonstrate the range of toxic elements that contaminate the environment consistent with CFA being the main aerosolize particulates used in climate manipulation.

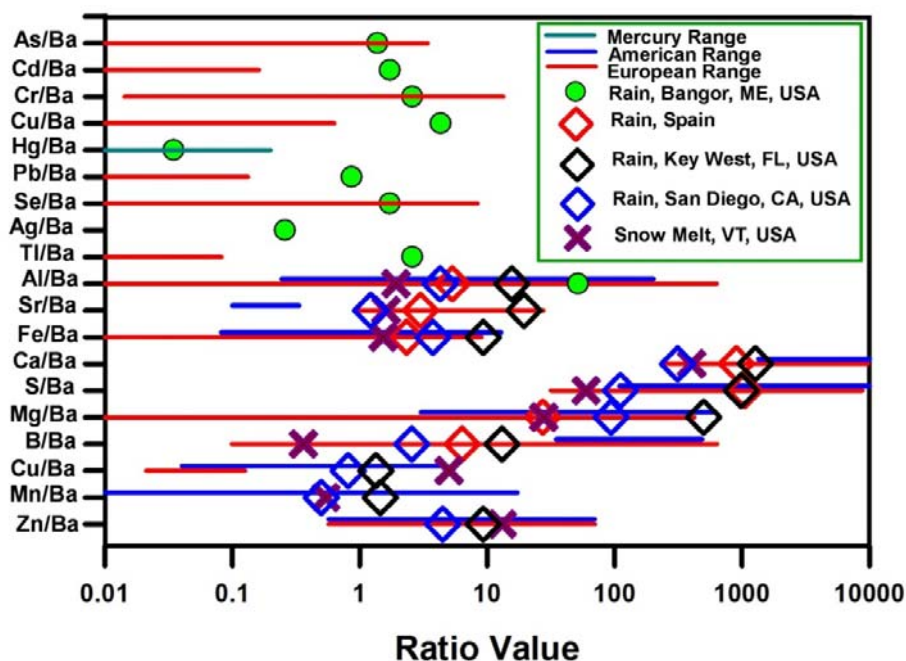


Fig. 2. Element ratios measured in filtered post-spraying rainwater and snow

The elemental composition of CFA is variable, dependent upon the compositions of the parent coals and the coal-burner dynamics. In Figs. 2 and 3, the ranges of CFA elemental compositions of European CFA samples are indicated by red lines, the ranges of American CFA elemental compositions by blue lines.

The aerosolized CFA mixes with the air we breathe and settles to Earth, hence the need for near-daily spraying. Consequently, CFA employed for climate manipulation/intervention grossly contaminates the biosphere with particulate toxic CFA and with toxins extracted from the CFA into rainwater [17,18,22].

The main elements in CFA are oxides of silicon, aluminum, iron, and calcium, with lesser amounts of magnesium, sulfur, sodium and potassium. Primary components of CFA are aluminosilicates and an iron-bearing (magnetic) fraction that contains magnetite, Fe_3O_4 . Coal fly ash is principally composed of spherical particles, including aluminosilicate and magnetite spherules [23]. The spherical configurations are due to surface tension of the melts during condensation and agglomeration in the hot gas above the coal burner [18]. Among the many trace elements originally present in coal that occur in CFA include arsenic (As), barium (Ba), beryllium (Be), cadmium (Cd), chromium (Cr),

lead (Pb), manganese (Mn), mercury (Hg), nickel (Ni), phosphorus (P), selenium (Se), strontium (Sr), thallium (Tl), titanium (Ti), vanadium (V), and zinc (Zn). Small amounts of organic material and even the radionuclides uranium (U), thorium (Th) and their radioactive daughter products are found in CFA [21,25].

Early studies of the adverse effects of air pollution on insects focused on volatile emissions including fluoride-containing gases, sulfur (SO_2), nitrogen oxides, and ozone [26]. It is now recognized that sustained exposure to particulate matter (PM) in air pollution is a major global cause of morbidity and mortality [27]. Coal fly ash is one of the main sources of anthropogenic particulate matter pollution on a world-wide basis [28]. Tropospheric aerosol geoengineering (TAG) operations, increasing in scope and intensity in recent years, represent a deliberate form of CFA-PM air pollution that also contaminates soil and water. This kind of particulate pollution can affect insects through respiration, ingestion, and direct contact. The particulate material in CFA, including metals and metalloids, are difficult for organisms to regulate, and are toxic to arthropods in various concentrations and by different modes of action [29].

Pollution caused by CFA can affect insects by bottom-up (e.g. soil or host plant quality) or top-

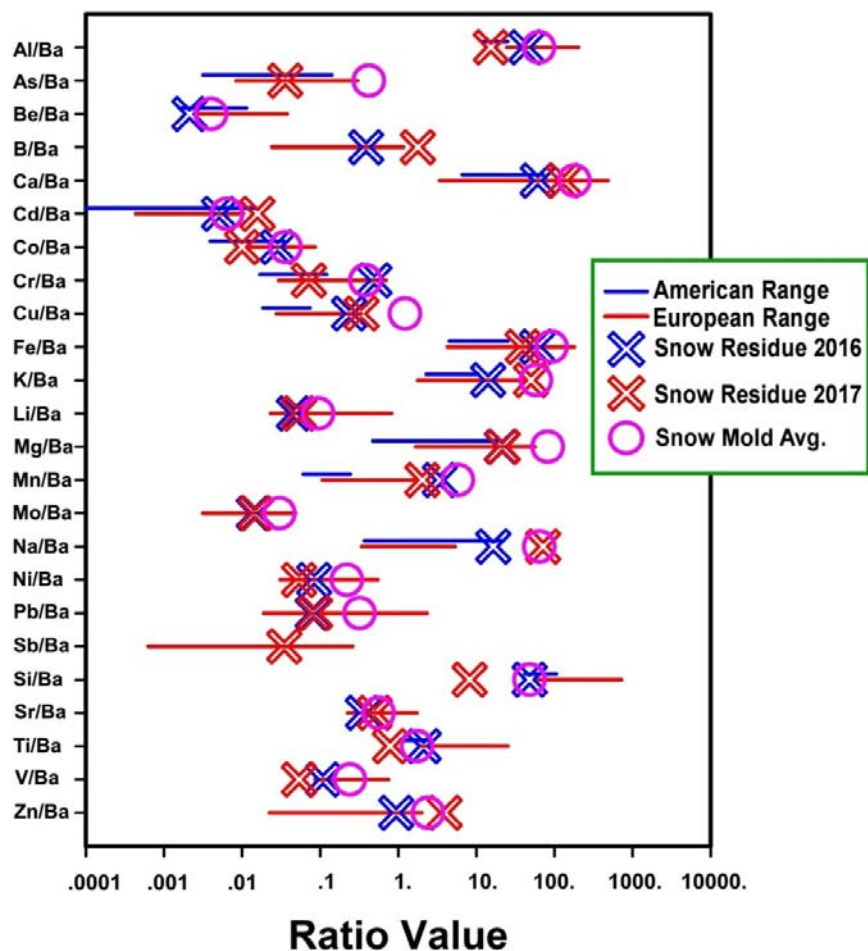


Fig. 3. Element ratios measured in post-spraying snow residue after evaporation and in snow mold found beneath melting snow. From [18] with permission.

down (e.g. direct contact or effects on predators or pathogens). A comprehensive review showed the fitness of insect herbivores was usually impacted by bottom-up factors. Fewer studies have been carried out by top-down factors, but it has been shown that air pollution does affect insect population dynamics by differential effects on herbivores and their natural predators [30]. Pollutants often bioaccumulate in predatory insects. Airborne pollution particles coat leaves and plants, affecting plant chemistry, photosynthesis, and thereby nutrition for herbivores. Contamination of soil allows for plant uptake of many elements that in turn are consumed by herbivores [31]. Coal fly ash added to fertilizer or soil can lead to potentially toxic accumulations of elements including arsenic [32].

The primary component elements of CFA, Si, Al, and Fe all have toxic effects upon insects. Deposition of Si in plant tissue provides a barrier

against insect probing, feeding, and penetration into plant tissue [33]. Silicon-bearing components remove the waxy coat of insects that preserves moisture, thus killing them by desiccation [34].

Moisture is capable of extracting aluminum from CFA in a chemically-mobile form [21]. Aluminum is usually not found in the natural world in chemically-mobile form thus there is an absence of defense mechanisms; aluminum is a non-essential metal with no biologic function. Aluminum is found in insecticides like aluminum phosphide, a highly toxic material used for grain preservation. Aluminum has been found to be toxic (causing deformities) in caddisfly larvae, with an enhanced effect in acid conditions [35]. In-vitro studies show aluminum toxicity in *Drosophila* flies [36]. Ingested aluminum is detrimental to foraging and other behaviors in bees [37].

As in other organisms, insects must balance opposing properties of ionic iron, that of an essential nutrient and a potent toxin. Iron must be acquired as a catalyst for oxidative metabolism, but it must be tightly regulated to avoid destructive oxidative reactions [38]. Ionic iron is one of the most reactive of all atmospheric pollutants. A biological effect common to many ambient air pollution particles is the disruption of iron homeostasis in cells and tissues [39]. Iron is known to play a catalytic role in the generation of oxygen free radicals in vitro. Houseflies fed ferrous chloride in their drinking water had shortened life spans with evidence of oxidative stress [40]. Iron accumulates in insects causing lipid peroxidation and eliciting an antioxidant response [41].

There is currently more direct evidence of pollution damage to insects from the main components of CFA. Exley et al. [42] reported that Bumble bee pupae from both urban and rural areas were found to be heavily contaminated with aluminum. This aluminum content was higher than levels considered harmful to humans and was associated with smaller Bumble bee pupae. High levels of aluminum and other elements found in coal fly ash (Cd, Co, Cr, Cu, Mn, Se, Sr, Ti and V) have been measured in honey bees from polluted areas [43,44]. High levels of aluminum, iron and multiple other trace elements including As, Pb, and Ba have been detected in bee pollen collected from polluted areas [45-47]. Bee pollen is a mixture of flower pollen, the bee's own secretions, and some nectar. It can be assumed that bees acquire significant amounts of metals and metalloid pollution from a "bottom-up" mechanism by ingestion of contaminated plant products and drinking water sources. In the case of bee pollen this material is brought back to the hive on the insects' legs and is one of their primary nutrition sources [45].

In addition to bees, other insects have been evaluated as bioindicators of heavy metal pollution, including those trace elements in CFA. In Pakistan, significant levels of Cd, Cu, Cr, Zn, and Ni were detected in a *libellulid* dragonfly, an *acridid* grasshopper, and a *nymphalid* butterfly. The highest levels of these elements were found near polluted industrial areas, and the lowest values (but still present) at a site far from industrial activity [48]. Accumulation of Cd, Co, Cu, Fe, Mn, Ni, and Pb were documented in grasshoppers (Orthoptera, Acrididae) that were collected near a copper mine in Bulgaria. Cadmium and lead were heavily concentrated in

grasshoppers at the most contaminated sites [48]. Concentrations of Pb > Cd > Hg were found in food plants and grasshoppers collected from a mountain grassland 1200 m above sea level in Greece, suggesting an anthropogenic source of pollution transported in the atmosphere [50].

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As bioindicators of pollution, honey bees are also used as samplers of airborne particulate matter. As reported by Negri et al. [51], honey bees foraging in polluted areas collect many inorganic pollution particles, mostly concentrated in the forewings, the head area, and back legs. These anthropogenic particles, ranging from 500 nm to 10 μ m in diameter, display a sub-spherical morphology and have been characterized by EDX as either Fe-rich particles or aluminosilicates. Lead and barium (both found in CFA) were also detected adhering to the body of the honey bee [51].

Coal fly ash is a rich source of nanoparticle-sized pollution. Nano and bio-nanoparticles are increasingly being studied and employed for insect control. Aluminum, silicon, zinc, and titanium nanoparticles (all components of CFA) are being developed for crop pest management [52]. For example, nanoaluminum dust can be engineered through modified synthesis to target different insect species [53]. Chemically fabricated nano-iron is being developed as an effective pesticide. It has been shown that iron and iron oxide nanoparticles are highly toxic to *Culex quinquefasciatus*, the Southern House Mosquito [54].

Recently spherical magnetite pollution nanoparticles like those in CFA, and distinct from biogenic magnetite particles, were found abundant in the brain tissue of humans with dementia [55]. Many insects (e.g. bees, ants, termites) contain biogenic magnetite and employ it for magnetoreception [56-58]. Honey bees, for example, use magnetite-based magnetoreception to detect the Earth's magnetic field by means of magnetoreceptor iron granules located in their abdomen [57]. It is therefore likely that exogenous magnetic pollution particles can disrupt these functions.

Magnetic measurements of deposited atmospheric dust serve as an additional parameter in assessing environmental pollution. Samples of this particulate atmospheric pollution contain magnetite of spherical shape, consistent with particles in the magnetic-magnetite fraction of coal fly ash [59]. Both biogenic and exogenous magnetite particles are known to be exquisitely

sensitive to external electromagnetic fields [60]. Insects are continually exposed to radio-frequency electromagnetic fields at different frequencies. The range of frequencies used in wireless communication systems will soon increase from 6 GHz to 120 GHz (5G). It has now been reported that insects absorb radiofrequency electromagnetic power as a function of frequency from 2 GHz to 120 GHz [61]. There is growing evidence that exposure to cell phone radiation induces stress, and can produce both behavioral and biochemical changes in worker honey bees [62].

Thermal (coal-fired) power plants (TPP's) have a long history of adverse environmental impacts due to their emissions of particulate matter, organic, and inorganic pollutants. Honey bees from apiaries foraging near TPP's accumulate high quantities of the primary (Al/Fe) constituents and trace elements (e.g. Cr, Ba, Cu, Li, and Ni) found in coal fly ash compared to bees from apiaries in rural areas [63]. Declines in honey bee populations due to pesticides have been studied, but the role of soil-borne pollutants on honey bee survival wasn't examined until recently. In regard to the soil-borne pollutant, selenium (Se), pollen collected by bees from plants growing in coal fly ash from TPP's contained 14 mg Se per kg [64]. In an urban but less polluted area of Poland, honey bee foragers collected from stationary hives contained 7.03 mg of Se per kg [65]. It was later shown that selenium in excessive amounts adversely affects honey bee behavior and survival. Bees foraging on nectar containing high levels of selenium (particularly selenate) suffer direct toxicity and population reduction from this soil-borne pollutant [66].

Coal fly ash itself has been used as a pesticide, with activity against many types of insects [16]. Many of the trace elements in CFA are quite toxic to insects. Before the development of organic/synthetic pesticides, inorganic chemicals and elements including arsenic, mercury, cadmium and boron were used as insecticides. Arsenic, cadmium, mercury and lead have no useful function in living organisms and may be toxic at any dose [67]. An insect model used to assess mercury toxicity found that mercury induces oxidative stress in insects just as it does it in vertebrates [68]. Cadmium chloride (CdCl_2),

mercuric chloride (HgCl_2), and methylmercuric chloride (MeHgCl) all produced marked toxicity including cell death in *Aedes albopictus* (mosquito) cells with $\text{MeHgCl} > \text{HgCl}_2 > \text{CdCl}_2$ [65]. We have shown that climate manipulation using aerosolized coal fly ash is likely a previously undisclosed and world-wide source of mercury contamination in the biosphere [18].

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Contamination of water in lakes, rivers, and other bodies of water by chemical pollutants is one of the most important threats to all wildlife including insects. Toxic elements of CFA readily leach into water where they concentrate in aquatic plants and insects. Selenium, one such element, is an essential trace nutrient, but it is toxic in higher amounts. The development and survival of insect herbivores can be affected by even low to moderate concentrations of selenium acquired from pollution in plants [70]. Elevated levels of copper, zinc, iron, manganese, lead, cobalt, and cadmium have been detected in water and aquatic insect body samples from polluted sites [71]. These pollutants have been shown to cause both oxidative stress and genotoxicity (e.g. chromosomal breaks/damage) in aquatic infections. Even small amounts of heavy metals can change the physiochemical characteristics of water and dramatically affect the metabolism of insects [71].

Another major contributor to the world-wide insect die-off is the thus-far widely unacknowledged but independently confirmed elevated level of short-wave ultraviolet UV-B and UV-C radiation penetrating to earth's surface [72-75]. We have proposed that this increase in deadly UV-B and UV-C radiation is partially caused by geoengineering utilizing CFA, which places ozone depleting chemicals (e.g. chlorine) high into the atmosphere [75]. The mutagenicity and lethality action spectra of sunlight exhibits two maxima, both in the UV-B and UV-C region [76]. Insects are very sensitive to changes in UV-B irradiance, and solar UV-B has a large direct and indirect (plant mediated) effect on arthropods [77]. It was recently shown that UV-B influences and disrupts the metamorphosis of insects [78]. UV-C radiation (100-290 nm) is well-known to be lethal to insects [79].

Table 1 presents a brief instructional overview of toxic effects from CFA constituents.

Table 1. Brief instructional overview of toxic effects from constituents of Coal Fly Ash (CFA)

Primary components of Coal Fly Ash (CFA):

Silicon (Si) – Deposition in plants creates a barrier for insect feeding/probing, and penetration into plant tissue. Silicon-bearing components remove/destroy waxy coat of insects causing desiccation.

Aluminum (Al) – CFA is the chief source of chemically-mobile aluminum. Aluminum, which has no biological function in insects, is used in insecticides (Al-phosphide). Aluminum toxicities include deformities and adverse changes in behavior/foraging (bees). Anthropogenic CFA aluminosilicate particles ‘coat’ insects including bees.

Iron (Fe) – Ionic iron is one of most reactive atmospheric pollutants. Biologically, iron excess causes oxidative stress and lipid peroxidation. Magnetite (Fe_3O_4) pollution particles ‘coat’ insects, and are exquisitely sensitive to external electromagnetic fields; they may interfere with magnetoreception in insects.

Nanoparticles (abundant in CFA) – Nanoparticles in CFA are reasonably assumed to be detrimental to insects as chemically fabricated Al, Si, and Fe nanoparticles are being developed for insect control.

Trace elements in Coal Fly Ash (CFA):

Arsenic (As), cadmium (Cd), mercury (Hg), and boron (B) have been used as insecticides. Arsenic, cadmium, mercury, and lead (Pb) have no known useful function in living organisms and may be toxic at any dose.

Selenium (Se) has been shown to concentrate in plants grown in CFA and accumulate with toxicity in insects (e.g. bees) foraging/feeding on those plants. Selenium in excess has been shown to be toxic to many organisms in the aquatic environment, including insects.

4. CONCLUSION

Coal fly ash, including its use in covert (undisclosed) climate engineering operations, is a previously unrecognized prime suspect in the world-wide decline of insects. CFA is a global source of pollution known to be toxic to insects that contaminates air, water, and soil. In fact, we suggest that of the many threats to insects, i.e. habitat loss/degradation, pesticides, foreign species and disease, atmospheric geoengineering, especially utilizing CFA, may well be not only the most dire, but the most neglected and unrecognized cause of the catastrophic loss of insects on a world-wide basis. Previously published data and updated in this study are consistent with CFA being the main undisclosed particulate aerosol used in tropospheric geoengineering. Coal fly ash adversely affects insects in aerial, terrestrial, and aquatic environments. Coal fly ash is implicated in the dramatic decline of insects because its primary components (alumino-silicates and iron) and multiple trace elements, are found in, on, and around insects collected in polluted areas from around the world. It is imperative to confirm and expand these findings and look for the “fingerprint” of CFA in rainwater, insects, and their surroundings in areas far removed from industrial sites but impacted by CFA aerosol spraying. Atmospheric geoengineering using CFA likely contributes the increasing irradiance by UV-B and UV-C radiation which is deadly to insects.

To date there has been no statistically significant cause ascertained to account for the demise of insects [1-12]. The precautionary principle, which is proposed as a new guideline in environmental decision making [80], consists of four central components: (1) taking preventive action in the face of uncertainty; (2) shifting the burden of proof to the proponents of an activity (in this case the aerial particulate spraying); (3) exploring a wide range of alternatives to possibly harmful actions; and, (4) increasing public participation in decision making, which in the matter of widespread insect demise should rightly include scientists. In this spirit we have disclosed potential primary, yet previously unacknowledged, causes of the catastrophic decline of insects. It is necessary to expose and halt atmospheric aerosol geoengineering to prevent further gross contamination of the biosphere. The gradual return of insects when the aerial spraying is stopped will be the best evidence that aerosolized CFA is in fact a leading cause of the current drastic decline in insect population and diversity.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Cox-Foster D, VanEngelsdorp D. Saving the honey bee. *Scientific American*. 2009;300(4):40-7.

2. Council NR. Status of pollinators in North America: National Academies Press; 2007.
3. Grixti JC, Wong LT, Cameron SA, Favret C. Decline of bumble bees (*Bombus*) in the North American Midwest. *Biological conservation*. 2009;142(1):75-84.
4. Goulson D, Lye GC, Darvill B. Decline and conservation of bumble bees. *Annu Rev Entomol*. 2008;53:191-208.
5. Cameron SA, Lozier JD, Strange JP, Koch JB, Cordes N, Solter LF, et al. Patterns of widespread decline in North American bumble bees. *Proceedings of the National Academy of Sciences*. 2011;108(2):662-7.
6. Evans JD, Saegerman C, Mullin C, Haubruge E, Nguyen BK, Frazier M, et al. Colony collapse disorder: a descriptive study. *PLoS ONE*. 2009;4(8):e6481.
7. Watanabe ME. Colony collapse disorder: many suspects, no smoking gun. *BioScience*. 2008;58(5):384-8.
8. Hallmann CA, Sorg M, Jongejans E, Siepel H, Hofland N, Schwan H, et al. More than 75 percent decline over 27 years in total flying insect biomass in protected areas. *PLoS ONE*. 2017;12(10):e0185809.
9. Brooks DR, Bater JE, Clark SJ, Monteith DT, Andrews C, Corbett SJ, et al. Large carabid beetle declines in a United Kingdom monitoring network increases evidence for a widespread loss in insect biodiversity. *Journal of Applied Ecology*. 2012;49(5):1009-19.
10. Conrad KF, Warren MS, Fox R, Parsons MS, Woiwod IP. Rapid declines of common, widespread British moths provide evidence of an insect biodiversity crisis. *Biological conservation*. 2006;132(3):279-91.
11. Thomas J. Monitoring change in the abundance and distribution of insects using butterflies and other indicator groups. *Philosophical Transactions of the Royal Society of London B: Biological Sciences*. 2005;360(1454):339-57.
12. Dirzo R, Young HS, Galetti M, Ceballos G, Isaac NJ, Collen B. Defaunation in the Anthropocene. *Science*. 2014;345(6195):401-6.
13. Deonaraine A, Bartov G, Johnson TM, Ruhl L, Vengosh A, Hsu-Kim H. Environmental impacts of the Tennessee Valley Authority Kingston coal ash spill. 2. Effect of coal ash on methylmercury in historically contaminated river sediments. *Environmental Science & Technology*. 2013;47(4):2100-8.
14. Harkness JS, Sulkin B, Vengosh A. Evidence for coal ash ponds leaking in the southeastern United States. *Environmental Science & Technology*. 2016;50(12):6583-92.
15. Environmental Protection Agency EPA: <https://www.epa.gov/coalash/coal-ash-reuse> Accessed June 4, 2018.
16. Basu M, Pande M, Bhadoria PBS, Mahapatra SC. Potential fly-ash utilization in agriculture: A global review. *Progress in Natural Science*. 2009;19(10):1173-86.
17. Herndon JM, Whiteside M. Further evidence of coal fly ash utilization in tropospheric geoengineering: Implications on human and environmental health. *J Geog Environ Earth Sci Intern*. 2017;9(1):1-8.
18. Herndon JM, Whiteside M. Contamination of the biosphere with mercury: Another potential consequence of on-going climate manipulation using aerosolized coal fly ash. *J Geog Environ Earth Sci Intern*. 2017;13(1):1-11.
19. Herndon JM. Evidence of variable Earth-heat production, global non-anthropogenic climate change, and geoengineered global warming and polar melting. *J Geog Environ Earth Sci Intern*. 2017;10(1):16.
20. Shearer C, West M, Caldeira K, Davis SJ. Quantifying expert consensus against the existence of a secret large-scale atmospheric spraying program. *Environ Res Lett*. 2016;11(8):p. 084011.
21. Moreno N, Querol X, Andrés JM, Stanton K, Towler M, Nugteren H, et al. Physico-chemical characteristics of European pulverized coal combustion fly ashes. *Fuel*. 2005;84:1351-63.
22. Herndon JM. Aluminum poisoning of humanity and Earth's biota by clandestine geoengineering activity: implications for India. *Curr Sci*. 2015;108(12):2173-7.
23. Chen Y, Shah N, Huggins FE, Huffman GP. Transmission electron microscopy investigation of ultrafine coal fly ash particles. *Environ Science and Technolgy*. 2005;39(4):1144-51.
24. Thomas W. *Chemtrails Confirmed*. Carson City, Nevada (USA): Bridger House Publishers; 2004.
25. Fisher GL. Biomedically relevant chemical and physical properties of coal combustion products. *Environ Health Persp*. 1983;47:189-99.
26. Alstad D, Edmunds Jr G, Weinstein L. Effects of air pollutants on insect

- populations. Annual Review of Entomology. 1982;27(1):369-84.
27. Forouzanfar MH, Alexander L, Anderson HR, Bachman VF, Biryukov S, Brauer M, et al. Global, regional, and national comparative risk assessment of 79 behavioural, environmental and occupational, and metabolic risks or clusters of risks in 188 countries, 1990–2013: a systematic analysis for the Global Burden of Disease Study 2013. The lancet. 2015;386(10010):2287-323.
 28. State of Global Air <https://www.stateofglobalair.org> Accessed June 4, 2018.
 29. Jensen P, Trumble JT. Ecological consequences of bioavailability of metals and metalloids in insects. Recent Res Dev Entomol. 2003;42:1-17.
 30. Butler CD, Trumble JT. Effects of pollutants on bottom-up and top-down processes in insect–plant interactions. Environmental Pollution. 2008;156(1):1-10.
 31. Trumble JT, Vickerman DB. Pollution and Terrestrial Arthropods. Encyclopedia of Entomology: Springer; 2004. p. 1787-9.
 32. Brake S, Jensen R, Mattox J. Effects of coal fly ash amended soils on trace element uptake in plants. Environmental Geology. 2004;45(5):680-9.
 33. Calatayud P, Njuguna E, Juma G. Silica in Insect-Plant Interactions. Entomol Ornithol Herpetol. 2016;5:e125.
 34. Mucha-Pelzer T, Debnath N, Goswami A, Mewis I. Comparison of different silicas of natural origin as possible insecticides. Communications in agricultural and applied biological sciences. 2008;73(3):621-8.
 35. Vuori KM. Acid-induced acute toxicity of aluminium to three species of filter feeding caddis larvae (*Trichoptera, Arctopsychoidea and Hydropsychidae*). Freshwater Biology. 1996;35(1):179-88.
 36. Kijak E, Rosato E, Knapczyk K, Pyza E. *Drosophila melanogaster* as a model system of aluminum toxicity and aging. Insect science. 2014;21(2):189-202.
 37. Chicas-Mosier AM, Cooper BA, Melendez AM, Pérez M, Oskay D, Abramson CI. The effects of ingested aqueous aluminum on floral fidelity and foraging strategy in honey bees (*Apis mellifera*). Ecotoxicology and Environmental Safety. 2017;143:80-6.
 38. Nichol H, Law JH, Winzerling JJ. Iron metabolism in insects. Annual Review of Entomology. 2002;47(1):535-59.
 39. Ghio AJ, Cohen MD. Disruption of iron homeostasis as a mechanism of biologic effect by ambient air pollution particles. Inhalation Toxicology. 2005;17(13):709-16.
 40. Sohal R, Allen R, Farmer K, Newton R. Iron induces oxidative stress and may alter the rate of aging in the housefly, *Musca domestica*. Mechanisms of ageing and development. 1985;32(1):33-8.
 41. Ferrero A, Torreblanca A, Garcerá MD. Assessment of the effects of orally administered ferrous sulfate on *Oncopeltus fasciatus* (*Heteroptera: Lygaeidae*). Environ Sci Pollut Res. 2017;24(9):8551-61.
 42. Exley C, Rotheray E, Goulson D. Bumblebee pupae contain high levels of aluminum. PLoS ONE. 2015;10(6):e0127665.
 43. van der Steen JJ, de Kraker J, Grotenhuis T. Spatial and temporal variation of metal concentrations in adult honey bees (*Apis mellifera* L.). Environmental Monitoring and Assessment. 2012;184(7):4119-26.
 44. Zhelyazkova I. Honey bees–bioindicators for environmental quality. Bulg J Agric Sci. 2012;18(3):435-42.
 45. Altunatmaz SS, Tarhan D, Aksu F, Barutcu UB, Or ME. Mineral element and heavy metal (cadmium, lead and arsenic) levels of bee pollen in Turkey. Food Science and Technology (Campinas). 2017(AHEAD):0-.
 46. Kostić AŽ, Pešić MB, Mosić MD, Dojčinović BP, Natić MM, Trifković JĐ. Mineral content of bee pollen from Serbia/Sadržaj minerala u uzorcima pčelinjega peluda iz Srbije. Archives of Industrial Hygiene and Toxicology. 2015;66(4):251-8.
 47. Sattler JAG, De-Melo AAM, Nascimento KS, Mancini-Filho J, Sattler A, et al. Essential minerals and inorganic contaminants (barium, cadmium, lithium, lead and vanadium) in dried bee pollen produced in Rio Grande do Sul State, Brazil. Food Science and Technology (Campinas). 2016;36(3):505-9.
 48. Azam I, Afsheen S, Zia A, Javed M, Saeed R, Sarwar MK, et al. Evaluating insects as bioindicators of heavy metal contamination and accumulation near industrial area of Gujrat, Pakistan. BioMed Research International. 2015;2015.
 49. Karadjova I, Markova E. Metal accumulation in insects (*Orthoptera, Acrididae*) near a copper smelter and copper-flotation factory (Pirdop, Bulgaria). Biotechnology & Biotechnological Equipment. 2009;23(sup1):204-7.

50. Devkota B, Schmidt G. Accumulation of heavy metals in food plants and grasshoppers from the Taigetos Mountains, Greece. *Agriculture, ecosystems & environment*. 2000;78(1):85-91.
51. Negri I, Mavris C, Di Prisco G, Caprio E, Pellecchia M. Honey bees (*Apis mellifera*, L.) as active samplers of airborne particulate matter. *PLoS ONE*. 2015;10(7):e0132491.
52. Kitherian S. Nano and Bio-nanoparticles for Insect Control. *Res J Nanosci Nanotechnol*. 2017.
53. Buteler M, Sofie S, Weaver D, Driscoll D, Muretta J, Stadler T. Development of nanoalumina dust as insecticide against *Sitophilus oryzae* and *Rhyzopertha dominica*. *International journal of pest management*. 2015;61(1):80-9.
54. Murugan K, Dinesh D, Nataraj D, Subramaniam J, Amuthavalli P, Madhavan J, et al. Iron and iron oxide nanoparticles are highly toxic to *Culex quinquefasciatus* with little non-target effects on larvivorous fishes. *Environ Sci Pollut Res*. 2018;25(11):10504-14.
55. Maher BA, Ahmed IAM, Karloukovski V, MacLauren DA, Foulds PG, et al. Magnetite pollution nanoparticles in the human brain. *Proc Nat Acad Sci*. 2016;113(39):10797-801.
56. Acosta-Avalos DL, Wajnberg E, Oliveira PS, Leal I, Farina M, Esquivel DM. Isolation of magnetic nanoparticles from *Pachycondyla marginata* ants. *Journal of Experimental Biology*. 1999;202(19):2687-92.
57. Liang C-H, Chuang C-L, Jiang J-A, Yang E-C. Magnetic sensing through the abdomen of the honey bee. *Scientific Reports*. 2016;6:23657.
58. Maher BA. Magnetite biomineralization in termites. *Proceedings of the Royal Society of London B: Biological Sciences*. 1998;265(1397):733-7.
59. Petrovský E, Zbořil R, Grygar TM, Kotlík B, Novák J, Kapička A, et al. Magnetic particles in atmospheric particulate matter collected at sites with different level of air pollution. *Studia Geophysica et Geodaetica*. 2013;57(4):755-70.
60. Kirschvink JL. Microwave absorption by magnetite: A possible mechanism for coupling non-thermal levels of radiation to biological systems. *Bioelectromag*. 1996;17:187-94.
61. Thielens A, Bell D, Mortimore DB, Greco MK, Martens L, Joseph W. Exposure of Insects to Radio-Frequency Electromagnetic Fields from 2 to 120 GHz. *Scientific Reports*. 2018;8(1):3924.
62. Kumar NR, Sangwan S, Badotra P. Exposure to cell phone radiations produces biochemical changes in worker honey bees. *Toxicology international*. 2011;18(1):70.
63. Zarić N, Ilijević K, Stanisavljević L, Gržetić I. Metal concentrations around thermal power plants, rural and urban areas using honey bees (*Apis mellifera* L.) as bioindicators. *International journal of environmental science and technology*. 2016;13(2):413-22.
64. De Jong D, Morse RA, Gutenmann WH, Lisk DJ. Selenium in pollen gathered by bees foraging on fly ash-grown plants. *Bulletin of environmental contamination and toxicology*. 1977;18(4):442-4.
65. Roman A. Levels of Copper, Selenium, Lead, and Cadmium in Forager Bees. *Polish journal of environmental studies*. 2010;19(3).
66. Hladun KR, Smith BH, Mustard JA, Morton RR, Trumble JT. Selenium toxicity to honey bee (*Apis mellifera* L.) pollinators: effects on behaviors and survival. *PLoS ONE*. 2012;7(4):e34137.
67. Tchounwou PB, Yedjou CG, Patlolla AK, Sutton DJ. Heavy metal toxicity and the environment. *EXS*. 2012;101:133-64.
68. Zaman K, MacGill R, Johnson J, Ahmad S, Pardini R. An insect model for assessing mercury toxicity: effect of mercury on antioxidant enzyme activities of the housefly (*Musca domestica*) and the cabbage looper moth (*Trichoplusia ni*). *Archives of Environmental Contamination and Toxicology*. 1994;26(1):114-8.
69. Braeckman B, Raes H, Van Hoyer D. Heavy-metal toxicity in an insect cell line. Effects of cadmium chloride, mercuric chloride and methylmercuric chloride on cell viability and proliferation in *Aedes albopictus* cells. *Cell biology and toxicology*. 1997;13(6):389-97.
70. Trumble JT, Kund G, White K. Influence of form and quantity of selenium on the development and survival of an insect herbivore. *Environmental Pollution*. 1998;101(2):175-82.
71. Shonouda M, El-Samad L, Mokhamer H, Toto N. Use of oxidative stress and genotoxic biomarkers of aquatic beetles *Anacaena globulus* (Coleoptera:

- Hydrophilidae) as biomonitors of water pollution. *J Entomol.* 2016;13:122-31.
72. Cabrol NA, Feister U, Häder D-P, Piazena H, Grin EA, Klein A. Record solar UV irradiance in the tropical Andes. *Frontiers in Environmental Science.* 2014;2(19).
 73. Córdoba C, Muñoz J, Cachorro V, de Carcer IA, Cussó F, Jaque F. The detection of solar ultraviolet-C radiation using KCl:Eu²⁺ thermoluminescence dosimeters. *Journal of Physics D: Applied Physics.* 1997;30(21):3024.
 74. D'Antoni H, Rothschild L, Schultz C, Burgess S, Skiles J. Extreme environments in the forests of Ushuaia, Argentina. *Geophysical Research Letters.* 2007;34(22).
 75. Herndon JM, Hoisington RD, Whiteside M. Deadly ultraviolet UV-C and UV-B penetration to Earth's surface: Human and environmental health implications. *J Geog Environ Earth Sci Intn.* 2018;14(2):1-11.
 76. Ravanat J-L, Douki T, Cadet J. Direct and indirect effects of UV radiation on DNA and its components. *Journal of Photochemistry and Photobiology B: Biology.* 2001;63(1):88-102.
 77. Ballare CL, Caldwell MM, Flint SD, Robinson SA, Bornman JF. Effects of solar ultraviolet radiation on terrestrial ecosystems. Patterns, mechanisms, and interactions with climate change. *Photochemical & Photobiological Sciences.* 2011;10(2):226-41.
 78. Sang W, Yu L, He L, Ma W-H, Zhu Z-H, Zhu F, et al. UVB radiation delays *Tribolium castaneum* metamorphosis by influencing ecdysteroid metabolism. *PLoS ONE.* 2016;11(3):e0151831.
 79. Hori M, Shibuya K, Sato M, Saito Y. Lethal effects of short-wavelength visible light on insects. *Scientific Reports.* 2014;4:7383.
 80. Kriebel D, Tickner J, Epstein P, Lemons J, Levins R, Loechler EL, et al. The precautionary principle in environmental science. *Environ Health Perspec.* 2001;109(9):871-6.