

4 **PARALLEL MECHANISMS BETWEEN PLACENTAL**
5 **AMYLOIDOSIS/PREECLAMPSIA AND NEURODEGENERATIVE DISEASES**
6

7 **Placental Prion like proteins in Preclampsia-a mini review**
8

9 **ABSTRACT**

10 This short review summarizes recent studies on placenta-preeclampsia (PE) in the
11 mother and/or intrauterine growth restriction (IUGR) in the child. The ideas raised here
12 are framed within a paradigm that favors the opening of new research lines in these
13 themes and are focused on the outlining of early investigation and/or an adequate
14 treatment for mothers who develop the pathology. Thus, this review focuses on those
15 studies that categorize PE in the group of pathologies defined as "conformational
16 diseases", as a consequence of the misfolding of proteins due to endoplasmic reticulum
17 ER stress. In this particular case, the ER stress that develops in the syncytiotrophoblast
18 (ST) because of the oxidative stress caused in the placenta by the hypoxia that occurs as
19 a consequence of the failure in the remodeling of endometrial arteries. This leads to an
20 increased ST syncytiotrophoblast apoptosis with detachment of misfolded proteins into
21 the maternal circulation, which in turn would be primarily responsible for the signs of
22 PE preeclampsia in the mother: proteinuria, edema, and hypertension. The review also
23 analyzes the PE preeclampsia-prions-placenta relationship, since the normal cell-surface
24 protein PrPc is normally present in the plasma membrane of ST syncytiotrophoblast ,
25 but appears to be increased in cases of PE preeclampsia . However, although
26 neurodegenerative disorders resulting from conformational changes in the prion protein
27 from its normal cellular form, PrPc, to the infectious scrapie isoform, PrPSc are well
28 known, limited information is available on Pr Pc and PrPSc in the ST, hence review on
29 these proteins gains more attention in normal and pathological placenta.
30

31 **Placenta and preeclampsia**

32 The passage of nutrients from the maternal blood to the fetus is mediated by the
33 placenta, so the normal fetal metabolism and growth require of an adequate exchange
34 across this organ [1]. The trophoblast is the epithelium that covers the placental fetal
35 villous tree and during development differentiates into two layers: the
36 syncytiotrophoblast (ST) and the cytotrophoblast (CT). The former is externally located
37 and contains many nuclei and a continuous cytoplasm, forming a syncytium. The latter
38 consists of a monolayer of ovoid cells immediately underlying the ST. Both structures
39 contribute to the formation of the villi and ultimately the placenta. Villous CT fuse in
40 order to form the ST layer that contributes to the metabolic exchange of gas and
41 nutrients, as well as to the process of waste elimination [2, 3]. Apoptosis of the
42 trophoblast has been observed to naturally occur in placentas of normal human
43 pregnancies but, as expected, placentas from women with preeclampsia (PE) or
44 intrauterine growth retardation (IUGR) show enhanced apoptosis when compared with
45 placentas from normal pregnancies [4].

46 PE is a systemic pregnancy syndrome that affects about 3-5% of all pregnancies
47 [5]. This pathology is an important contributor to maternal and perinatal morbidity and
48 mortality worldwide. Because there is no cure other than delivery, PE is the leading
49 cause of iatrogenic preterm birth. Despite to be of unknown etiology, it is currently

50 accepted that this pathology originates in the placenta [6] due to the fact that the
51 maternal symptoms (high blood pressure and proteinuria) disappear once the organ has
52 been expelled after delivery [7, 8].

53

54 **Oxidative stress**

55 To date, PE has been related to the process of hypoxia due to
56 ischemia/reperfusion experienced by the placenta as a consequence of extravillous
57 trophoblast failure in the process of endometrial spiral arteries remodeling. The
58 involvement of oxidative stress (OS) in the early placental hypoxia development has
59 been previously proposed in the mechanism of the syndrome [1, 5, 8, 9]. Due to the fact
60 that the human fetal-placental vasculature lacks autonomic innervation, it is reasonable
61 to assume that autocrine and/or paracrine agents such as the NO radical may play an
62 important role in the regulation of fetal-placental blood flows [9].

63 On the other hand, OS constitutes a unifying mechanism of injury involved in
64 many types of disease. It occurs when there is an imbalance between the production of
65 ROS and the ability of the biological system to readily detoxify these reactive oxidative
66 species (ROS) or the tissues cannot easily repair the resulting damage [10]. In PE it has
67 been shown that enhanced ROS generation leads to a decrease in the NO bioavailability
68 [11]. Increased generation of superoxide anion by the placenta leads to increased
69 peroxynitrite production, resulting in further oxidative stress and endothelial
70 dysfunction in PE patients [8]. Additionally, it has been well established that NO
71 disrupts the mitochondrial respiratory chain in a dose dependent manner, causing
72 changes in the mitochondrial Ca^{2+} flux that induce ER Stress in pluripotent stem cells
73 [12]. Taking all of these evidences into account, it is plausible to assume that OS
74 developed in the placenta by the exaggerated generation of ROS would trigger ER stress
75 in the organ, which in turn will increase the apoptosis of the ST.

76

77 **Placental ER stress and Amyloidosis**

78 In the last few years, a number of studies suggesting that PE could be triggered
79 by disorders in the folding of proteins in the ER of the ST, which results in amyloid
80 deposits in this organelle [13-16] have been published. In light of this evidence, the
81 accumulation of misfolded protein in the ER lumen has been defined as 'ER Stress' [17-
82 20].

83 In addition, ER stress has recently been identified as a major regulator of cell
84 homeostasis through its involvement in post-translational protein modification and
85 folding, as well as its capacity to activate the unfolded protein response (UPR) which
86 aims to restore the homeostatic balance within the ER [21]. If this cannot be achieved,
87 the cell apoptotic machinery becomes consequently activated. The initial intent of the
88 UPR is to adapt the cell to the changing environment, and reestablish normal ER
89 function. These adaptive mechanisms involve transcriptional programs that induce
90 expression of genes that enhance the protein folding capacity of the ER, and promote
91 ER-associated protein degradation to remove misfolded proteins [17]. Persistent protein
92 misfolding initiates apoptotic cascades [21] that are known to play fundamental roles in
93 the pathogenesis of multiple human diseases, including diabetes, atherosclerosis, PE and
94 neurodegenerative diseases [14-16,22,23], all of which have been defined as
95 "conformational diseases".

96 The ER stress due to misfolded proteins in the ST increases placental apoptosis
97 in this epithelial layer [16, 18]. Moreover, due to the fact that the ST establishes direct
98 contact with the maternal blood, the apoptotic process produces detachment of the
99 syncytial infolding proteins, accumulated due to ER stress, to the maternal blood.

100 Consequently, these particles will be mainly responsible for the development of PE
101 symptoms in the mother. Recently, **we** found that the Amyloid A (AA) was present in
102 the ST of PE and IURG placentas, and that the degree of apoptosis of the CT regulates
103 the amyloidosis destiny of the AA in the ST [16]. In brief, in PE cases the misfolded
104 proteins are expelled to the maternal blood. On the contrary, in the IURG cases they are
105 deposited on the basal lamina of the trophoblast, without being expelled from the
106 placenta, but also altering the mother/fetus metabolic exchange, thus producing IUGR.
107 Moreover, Hitomi et al., [24] suggested that activation of ER-resident caspase-12
108 indirectly activates cytoplasmic caspase-3 and might be important in ER stress-induced
109 neuronal apoptosis as a consequence of the presence of misfolded proteins. This is in
110 agreement with **our** the placental study of ~~Boseo et al.~~, [16] which showed the presence
111 of active caspase 3 in the CT of PE placentas with AA amyloidoses, but not in the CT of
112 normal placentas.

113 It has also been reported that caspase-12-deficient mouse cortical neurons were
114 defective in apoptosis induced by amyloid-beta protein, but not by trophic factor
115 deprivation [25]. Thus, caspase-12 mediates an ER-specific apoptosis pathway and may
116 contribute to amyloid-beta neurotoxicity. This idea is in concordance with Fu et al.,
117 [26] who found significantly higher caspase 12 activity in placentas of early or late
118 severe PE. It is important to note that ER stress apoptosis can be induced by other
119 various pathological conditions that alter the ER function. In the same line of evidence,
120 Wang et al., [27] experimentally induced ER stress and apoptosis in placentas of
121 pregnant rats exposed to lead, which was accompanied by an increase in the caspase-12
122 mRNA expression, and Xu et al., [28] found an increase in the early expression of ER
123 stress markers, followed by increased activity of caspase 12 in placental trophoblast
124 exposed *in vivo* and *in vitro* to *T. gondii*, followed by an increased apoptosis of the
125 exposed trophoblasts. Similar results were found by Wang et al., [29] in neural stem
126 cells exposed to this parasite. It should be emphasized that in the last three
127 investigations no studies were carried out in order to evaluate the presence of
128 misfolding proteins in the placentas, which would have allowed amyloidosis to be
129 discarded. It is also important to note that in a case control study where pregnant
130 women suspected of *T. gondii* infection were treated with spiramycin, a macrolide
131 antibiotic administered before 18 weeks of pregnancy in order to reduce the rate of
132 transmission of the parasite to the fetus, reported a reduced incidence of pregnancy-
133 induced hypertension [30]. On the basis of these results, the association of *T. gondii*
134 infection with hypertension disease during pregnancy needs to be further investigated.

135 In another line of evidence, transthyretin (TTR) is a homotetrameric serum and
136 cerebrospinal fluid protein. The TTR dissociation forms monomer misfolding, a variant
137 of TTR that results in familial amyloid polyneuropathy, familial amyloid
138 cardiomyopathy, or familial central nervous system amyloidosis [31]. TTR is also a
139 carrier protein for thyroxin and retinol binding protein, which are secreted by
140 trophoblast. McKinnon et al., [32] and Mortimer et al., [33] have reported that human
141 placenta secretes TTR into the maternal and fetal circulations and that placental TTR
142 secreted into the maternal placental circulation can be taken up by the trophoblasts and
143 translocated to the fetal circulation, thus conforming a TTR shuttle system. This may
144 have important implications for maternal-fetal transfer of thyroid hormones,
145 retinol/retinol binding protein and xenobiotics, all of which bind to TTR. Additionally,
146 Fruscalzo et al., [34] demonstrated that TTR is dysregulated in cases of IUGR and
147 severe early onset PE, and Kalkunte, et al., [13] showed the presence of amyloid
148 aggregates of TTR in PE placentas, as well as in the serum of these patients.

149 Taken together, all these evidences allow us to postulate that by effect of the OS
150 the placenta develops ER stress in the ST and CT, which leads to the accumulation of
151 misfolded proteins and, if the quantity greatly increases, this will finally activate the
152 UPR with the consequent increase of ST apoptosis and therefore the release of the
153 misfolded proteins into the maternal blood, which in turn will trigger the symptoms of
154 PE in the mother.

155

156 **Normal cellular prion protein form in placenta**

157 The study of this prion protein was initiated due to its involvement in a number
158 of related neurodegenerative disorders seen in various species (bovine spongiform
159 encephalopathy in cattle, scrapie in sheep and Creutzfeldt–Jakob disease in humans).
160 The name ‘prion’ (for Proteinaceous Infectious) was coined as the infectious agent of
161 these diseases was found to be significantly constituted by proteins [35]. A protein with
162 identical sequence was found to be expressed in significant quantities in the brains of
163 non-diseased animals. Hence, a consensus was reached that the protein existed in two
164 distinct forms: the normal cellular prion protein form (PrPc) and the diseased or scrapie
165 form (PrPSc). However, recent evidence suggests that the scrapie form of the protein
166 may be sufficient by itself for transmission of the disease [36]. Transmissible
167 spongiform encephalopathies (TSE) or prion diseases are characterized by the
168 deposition of PrPc in the structurally altered PrPsc form. While PrPc configuration is
169 primarily α -helix and susceptible to proteolysis, PrPSc instead forms fibrillar aggregates
170 containing a high percentage of β -sheet and is rather resistant to proteolytic digestion
171 [37]. TSE condition is accompanied by physiological symptoms similar to those of
172 aging which, in turn, have been shown to be affected by divalent metal ions [38,39].
173 Over the past three decades, the role of metal ions in TSE has attracted considerable
174 attention particularly since 1970s, when Cu^{2+} chelator-induced histopathological
175 changes were documented to be similar to scrapie [40]. Metal ions have been implicated
176 as potential pathogenic candidates owing to their properties of being free-radical
177 generators and their association with metalloenzymes such as superoxide dismutases
178 (SODs), **redox enzymes important for cellular resistance to oxidative stress** [41].
179 Pathological features of TSE resemble neuronal and brain tissue loss as is observed in
180 the case of free radical-mediated oxidative damage [42].

181 On the other hand, PrPc, a copper-binding glycoposphatidylinositol-anchored
182 protein whose function is to protect the cells against oxidative stress and to prevent the
183 apoptosis it is expressed in the plasma membrane of neural and not neural tissues [43-
184 46]. A number of roles in neuroprotection, cellular homeostasis, response to oxidative
185 stress, cell proliferation and differentiation, synaptic function and signal transduction
186 have been proposed for PrPc [43,46]. Additionally, it has been shown that the abnormal
187 isoform of PrPSc is able to induce further PrPc \rightarrow PrPSc transition, accumulating in
188 infected brains and forming amyloid plaques involved in prion diseases such as TSE, a
189 disease with neuronal death and gliosis, producing extensive and sponge-like tissue
190 vacuolization [37,38,47]. Additionally, Hetz et al., [48] demonstrate that prion diseases
191 characterized by accumulation of the misfolded protease-resistant form of the prion
192 (PrPSc) produce neuronal death by apoptosis that also correlated with caspase 12
193 activation in neural mouse cells treated with PrPSc. Furthermore, it has also been
194 reported that the hypoxia-inducible factor-1 alpha (HIF-1 α), which appears to be a
195 master regulator of the cellular response to hypoxia [49], regulates PrPc expression in
196 order to protect against neuron cell damage [50]. In correlation with this, a variety of
197 studies have shown that women with PE are characterized by persistently elevated
198 placental HIF-1 α levels that promote enhanced transcription of genes encoding the

199 soluble antiangiogenic protein fms-like tyrosine kinase-1 (sFlt-1), the soluble
200 antiangiogenic factor endoglin (sEngs) and endothelin-1 (ET-1), a powerful
201 vasoconstrictor known to contribute to this pregnancy pathology [51-55]. Moreover,
202 Donadio et al., [56] and Alfaidy et al., [57] reported that PrPc is highly expressed in the
203 human placenta, especially in CT and ST, and Hwang et al., [58] found that the
204 immunohistochemical expression of PrPc was increased in CT and ST of PE placentas
205 versus those from the controls. Additionally, Brown et al., [59] and Brown and
206 Besinger [60] demonstrated in mouse neurons that PrPc may directly or indirectly
207 regulate the activity of Cu/Zn superoxide dismutase (Cu/Zn SOD). In this context, our
208 group found a decreased activity of Cu/Zn SOD in PE placentas versus normal
209 placentas with an increased of F2-isoprostanes, a lipid peroxidation indicator [61].
210 Furthermore, Klamt et al., [47] found a decreased activity of SOD in liver, heart,
211 hippocampus and cerebellus in PrPc knockout and wild-type mice and an oxidative
212 damage in proteins and lipids. In addition, Anantharam et al., [46] found that PrPc
213 plays a proapoptotic role during ER stress.

214 On the bases of the above arguments, we consider of the essential interest to
215 carry out new research aimed at investigating the possible presence of PrPsc in ST and
216 CT in cases of severe PE and eclampsia. This, due to the fact that poorly folded
217 proteins form amyloid precipitates, and because in PE, our group found a decrease in
218 the activity of the antioxidant enzyme SOD [61] which is regulated by PrPc [60]. It is
219 noteworthy that in the cases of pregnant mothers who develop eclampsia, the maximum
220 expression of PE, the maternal endothelial damage can lead to severe intracranial
221 (intracerebral and subarachnoid) hemorrhage and cerebral venous thrombosis, preceded
222 by visual hallucinations and the final appearance of convulsions and coma [62].

223 We would like to hypothetically propose that the presence of PrPSc in the ST
224 and CT of the placenta of these mothers could be related to the increase of apoptosis in
225 these cells and also with the significant maternal endothelial damage observed, since the
226 release of PrPSc into the maternal blood would allow these misfolded proteins reach the
227 blood-brain barrier. Therefore, it would be essential to perform brain biopsies of women
228 who have died from eclampsia for the determination of amyloidosis and/or PrPSc [63].
229 Finally, it is important to note that in sheep placentas exposed naturally to PrPSc, the
230 presence of PrPSc in the trophoblast has been shown by immunohistochemistry and/or
231 ELISA essays [64].

232 CONCLUSIONS

233 This review concludes that the latest studies show evidence of a relationship
234 between the presence of placental amyloidosis and PE. The amyloidosis condition may
235 be either due to an increased ER stress in the trophoblast, or to an increase in the
236 caspase 12 activity in the ER of these cells and an increase in caspase 3 activity in the
237 CT, whose cells define the fate of the misfolding proteins in the ST.

238 This review also emphasize the presence of the normal prion PrPc in the plasma
239 membrane of the CT and ST, whose presence has been noticed to increase in cases of
240 PE. However, it is important to analyze what type of response will occur if *in vitro*
241 BeWo cells are subjected to exposure of the prion isoform PrPSc.

243 PROJECTIONS

244 Future studies are required in order to elucidate the functional role of increased
245 amyloidosis and PrPc in the placenta of PE pregnancy, and to establish whether the

246 determinations of amyloids [14] or PrPc [59] in urine or serum of this women could be
247 used to prevent or predict this pregnancy pathology.

248

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