Original Research Article

Assessement of phenology and morphological diversity of 3 species of Asteraceae :

Anacyclus clavatus*, *Chamaemelum fuscatum* and *Leucanthemum parthenium

Abstract

Aim : 3 species of Asteraceae : *Anacyclus clavatus*, *Chamaemelum fuscatum* and *Leucanthemum parthenium* that have a wide range of uses in medicine and in industry were characterized by inter-specific variations and phenological activities.

Study Design : Morphological characterization using 18 quantitative traits and phenology study like vegetatif study, flowering time, fruiting time and seed formation for two consecutive years.

Place and Duration of Study: Experimental plot at the Faculty of Sciences of Tunis, Tunisia- 2009-2010.

Methodology: Measurements of the 18 morphological characters were performed on 3 samples of *Anacyclus clavatus*, *Chamaemelum fuscatum* and *Leucanthemum parthenium* grown in the Faculty of Sciences of Tunis, for each species, we have studied 10 individuals. Different phenological stages : Vegetative period, flowering and Fruiting presented by the individuals of each species are studied.

Results : The phenological study show that the 3 species studied have distinct phenologies. The longest phenological cycle is observed for *Leucanthemum parthenium.* Results of morphology study showed significant differences to highly significant for the majority of the traits studied using variance analysis. The comparison of means reveals that *Anacyclus clavatus* and *Chamaemelum fuscatum* form a single group for most of the traits measured, while *Leucanthemum parthenium* is clearly distinct from these two species. In addition, the principal component analysis confirms the results of the variance analysis and the comparison of means.

Conclusion : The results of the phenological cycles follow-up show that the 3 species studied have distinct phenologies. The longest phenological cycle is observed for *Leucanthemum parthenium.* The morphological study reveals that *Anacyclus clavatus* and *Chamaemelum fuscatum* form a single group while *Leucanthemum parthenium* is clearly distinct from these two species.

Keywords *: Anacyclus clavatus ; Chamaemelum fuscatum ; Leucanthemum parthenium ; morphological ; phenology.*

1. Introduction

Phenological study is important in plant management and combating afforestation, honey analysis, floral biology, estimation of reproductivity and regeneration [1]. It is important also in understanding species interrelations and their interaction with the environment. Variations in phenophases among individuals of different species have been linked to environmental perturbations [2].

A clear understanding of phenological behavior on time of anthesis, time and duration of stigma receptivity, fertilization, mode of pollination, seed development is necessary for breeding programmes to obtain better traits [3]. Thus plant phenological study has great significance because it not only provides knowledge about the plant growth pattern but it also provides the idea on the effect of environment and selective pressure on flowering and fruiting behavior [4].

Evaluation and characterization through morphological parameters of different crop germplasm is therefore so much important for all plant breeders [5]. Therefore, it is important to make proper strategies for the collection and evaluation of germplasm sources which are locally used in different regions of the world and save them from being vanished [6]. To have a variety of better traits of any crop we need informations about its genetic diversity [7]. Thus, characterization and estimation of genetic diversity is an important step for the competent and successful maintenance and utilization of different crop germplasm [8].

Genetic diversity is an inherited variation among and between populations, created, activated and maintained by evolution [9]. Morphological traits provide a simple way of measuring genetic diversity while studying genotype performance under normal growing conditions, but are influenced by environmental factors ([10] ; [11]). Plants have the potential to response to the changed environments by changing their morphology and there for, the intra-specific variation in plant characteristics is usually regarded as the adaptive mechanism to different environments [12].

The Asteraceae is one of the largest families, comprising 25.000 species, It is known for its wide range of uses not only in medicine but also some plants are grown as ornamental plants such as

chamomile (*Leucanthemum parthenium*), others can provide different products: natural rubber, colorants, insecticides and spices [13].

A. clavatus (*Anacyclus clavatus*), belonging to the Asteraceae family and the genus *Anacyclus*, is an herbaceous, annual and spontaneous plant that is found almost everywhere in the Mediterranean region [14]. It's 20 to 50 cm tall, hairy, green or whitish-green, with an upright or ascending stem, woolly and rowdy whose branches are divorced. Leaves are bipinnatized, long to very narrow segments terminated by a small mucron [15]. The convex or somewhat conical receptacle carries triangular bracts, ovals in the shape of sequins. The inflorescences have two types of hermaphrodite flowers: the central flowers are yellow-colored and the peripheral flowers are tongued, long and white. They flourished from March to June [14].

The fruits in the form of achenes are small, very compressed cuneiform and of gray to beige color [15]. The number of chromosomes of this species is 2n = 18 [16]. It's a plant that grows on the edges of fields and roads and in the wastelands of the entire Mediterranean coast [15].

In Tunisia, it's is located in the north (Kroumirie, Oued Medjerda and Cap Bon), and in the center. The use of this species is very limited. The aerial part of *A. clavatus* is used as a powder against stomach and belly pains. It may also be one of the components of tobacco [17].

C. fuscatum (*Chamaemelum fuscatum*), belonging to the Asteraceae family, anthemidae tribe, *Anthemis* genus and Ormenis sub-section, is an annual, herbaceous, glabrous 30 cm rowing, ascending or upright.

The leaves are bipinnatized. The heads are heterogeneous with yellow disc and white ligules; their flowering is very early from November to April. The achene is very small, striated, tetragonal and brown to yellow in color. It's a very widespread plant on the banks of the seguias.

In Tunisia, *C. fuscatum* is found everywhere: in the north (Ain Drahim, Kef, etc.), in the center (Sousse, Enfidha, etc.) and in the South (Gabes, etc.). Internationally, It's located in the western Mediterranean basin of Spain, Greece and North Africa (Tunisia, Morocco and Algeria) [15]. The number of chromosomes of this species is 2n = 18 [18]. It's known for its anti-malaria property and its protective effect against cell damage [19].

L. parthenium (*Leucanthemum parthenium*) belonging to the Asteraceae family too, the Anthemidae tribe and the Asteroida subfamily [20] and the *Leucanthemum* genus. This chamomile is a very fragrant, perennial, rooted plant, with flowering stem erect without hair.

The leaves are deeply divided into 4 to 12 toothed segments. The internal tubular flowers are yellow and the ligulate external flowers are white. They flourish from June to August in European conditions [14] and from July to October in Iran [21]. The ripe fruits are brown, glandular and surmounted by a very short membranous crown.

L. parthenium is a medicinal plant used primarily for the prevention and reduction of migraine attacks frequency, against stomach aches and malaria [22]. It's also known for its properties: antiseptic, stomachic, antihysteric, vermifuge and insecticide. It's found spontaneously on the edges of roads and often in the vicinity of dwellings and it can also be grown in gardens as an ornamental plant. Internationally, *L. parthenium* is found almost all over Europe except the boreal zone and it is also found in South-Western Asia [14].

However, there is little information on the morphological diversity and the phenology of *Anacyclus clavatus*, *Chamaemelum fuscatum* and *Leucanthemum parthenium* and the potential of these species in breeding programs. The aim of this study is to assess the variations in morphology and phenology of *A. clavatus*, *C. fuscatum* and *L. parthenium*.

2. Materials and methods

2.1. Plant material

Three species of Asteraceae have been studied in this work: *Anacyclus clavatus*, *Chamaemelum fuscatum* and *Leucanthemum parthenium*. These species were grown on an experimental plot at the Faculty of Sciences of Tunis, Tunisia under uncontrolled conditions. The seeds used originate from Esbikha for *A. clavatus*, Haouz (Morocco) for *C. fuscatum* whereas the seeds of *L. parthenium* are available in the laboratory of Genetics and Bioresources of the Faculty of Sciences of Tunis.

2.2. Phenological characters

Different phenological stages presented by the individuals of each species are defined:

2.2.1.Vegetative period

This stage spreads from the planting to the beginning of flowering. This is the phase of vegetative growth.

2.2.2. Flowering

This is the period during which the flowers appear. The method of study is based essentially on the visual observation of the appearance of the flowers.

2.2.3. Fruiting

This phase is characterized by the formation of the fruit. It begins with the formation of the first seeds and ends with the general ripening of the seeds.

2.3. Morphological traits

In order to compare the various species studied, we describe the characters of their vegetative part: The type of branching, the stem, the structure and color of the leaves, the structure and color of the inflorescences and flowers, the structure and color of akene and the weight of 100 akene.

Measurements of the morphological characters were performed on three samples of *Anacyclus clavatus*, *Chamaemelum fuscatum* and *Leucanthemum parthenium* grown in the Faculty of Sciences of Tunis, for each species, we have studied 10 individuals. The 18 morphological quantitative traits were assessed to characterize and estimate genetic diversity among the 3 species studied, the quantitative traits measured were :

- Length of main axis in cm: LAP
- Average length of primary branch in cm: LMRP
- Average length of branch in cm: LMRS
- Average length of the tertiary branch in cm: LMRT
- Length of main root in cm: LRP
- Number of leaves per plant: NF
- Average diameter of the receptacle in cm: DMR
- Average number of leaflets per leaf: NMf

- Average length of the leaf rachis in cm: LMRF
- Number of inflorescence per plant: NI
- Number of primary branches: NRP
- Number of secondary branches: NRS
- Number of tertiary branches: NRT
- Average number of ligules per head: NML
- Number of ligules of the main axis head: NLCAP
- Length of the smallest branch in cm: LPR
- Length of the longest branch in cm: LLR
- Weight of 100 akene : P_{100} A

2.4. Data analysis

In order to evaluate morphological diversity and to establish relationships among studied species, several statistical procedures were conducted. Quantitative data were computed using the software XLSTAT version 2011 to perform analysis of variance, comparison of mean and to calculated the Pearson correlation coefficient. Principal component analysis (PCA) was also done using the software XLSTAT.

3. Results and discussion

3.1. Phenology study

3.1.1. Vegetative period

The vegetative period is characterized by a strictly herbaceous development and extends from seedling to full bloom. We divided this phase into 2 stages:

Stage of germination: it is characterized by the appearance of the primordial leaves. In all three species, the germination begins after 10 days.

Stage of foliage: Observation of the phenological spectrum reveals that this stage is the longest of the phenological cycle. This stage, which is characterized by the growth of the stems in length and by

the formation of the leaves, lasts 6 months for *Chamaemelum fuscatum* (Figure 1) and 7 months for *Anacyclus clavatus* (Figure 2). *Leucanthemum parthenium* is a perennial herb plant (Figure 3).

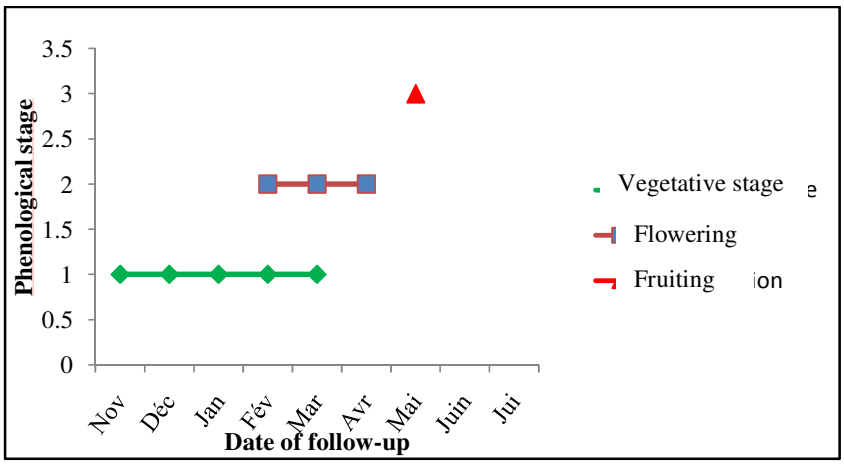

Fig.1. Phenological cycle of *Chamaemelum fuscatum*

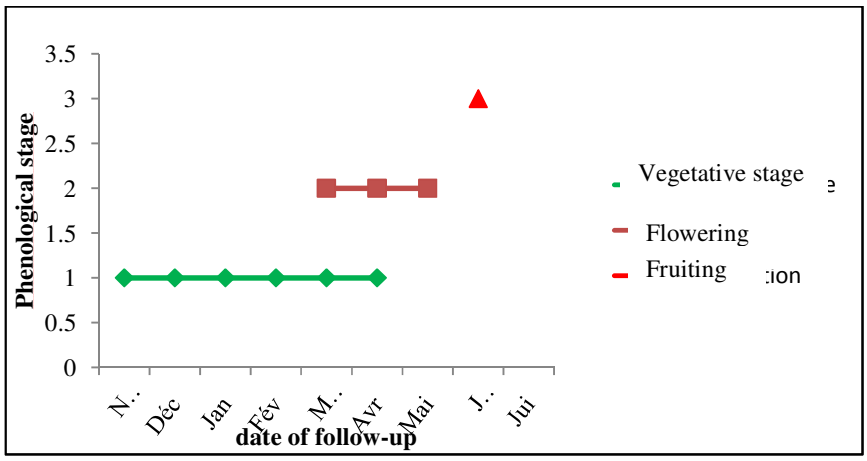

Fig. 2. Phenological cycle of *Anacyclus clavatus*

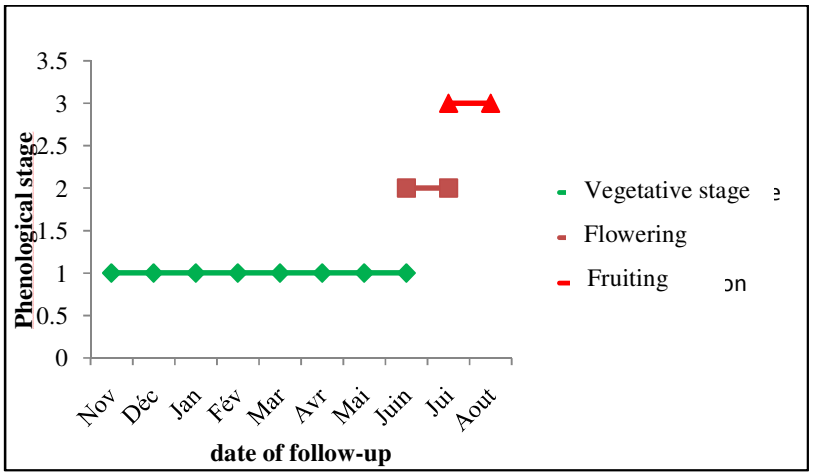

Fig.3. Phenological cycle of *Leucanthemum parthenium*

3.1.2. Flowering

Flowering is considered from the formation of the first flower until most flowers have evolved this period differs from one species to another:

For *Chamaemelum fuscatum*, the flowering period ranges from mid-February to the end of April (Figure 1).

For *Anacyclus clavatus*, this period extends from the end of March to mid-May (Figure 2).

For *Leucanthemum parthenium*, the first flower blooms in early June and full bloom is observed around mid-July (Figure 3).

Flowering appears to be highly favoured during the rainy season for *Anacyclus clavatus* and *Chamaemelum fuscatum*, only *Leucanthemum parthenium* flowers during the dry season.

We find that the species *Chamaemelum fuscatum* characterized by a very early flowering date has a spread flowering period. In addition, the species *Leucanthemum parthenium* characterized by a late flowering date has a relatively short flowering stage and this to escape the water stress.

3.1.3. Fruiting

It is the formation of fruit in the form of akene. We have noticed that the appearance of the first akene coincides with the peak of flowering, while the full fructification characterized for the 3 species by the change of color flowers in tubes from yellow to light gray and the fall of the white ligules is generally obtained after two weeks of the appearance of the first fruit (Figure 1, 2 and 3).

In fact, the study of [23] reveals that achenes of *A. clavatus* that germinated earlier produced plants with higher biomass and higher reproductive effort. In addition, this work show that the phenology of *Anacyclus clavatus* achene germination was the main factor affecting postdispersal life-history traits related to competitive ability and reproductive success.

In addition, the study of [24] showed a high phenological diversity for the four phenological patterns (buds, flowers, fruits and seeds) among fifteen leguminous plant species growing in Amritsar.

3.2. Morphology study

3.2.1. Study of vegetative part

It allows us to obtain, on the one hand, the resemblances, the objective of the typology of organs in the sense of a unity of organization and, on the other hand, the differences : the possible variations around the type : it's the comparative morphology. It seems useful to draw up a comparative table of the morphological characteristics of the 3 species studied (Table 1).

Table 1 : Main distinctive characteristics of 3 species studied.

Species	NR	Leafs	Flowers	Akenes	P_{100} A in mg
Anacyclus clavatus	T+5	Dark green Bipennatized	Flowers in yellow tubes and white ligulated flowers.	Beige.	45.23
Chamaemelum fuscatum	T+5	Green Bipennatized	Flowers in yellow tubes and white ligulated flowers.	Brown to yellow.	26.6
Leucanthemum parthenium	T+3	Greenish-yellowish divided into wide segments.	Flowers in yellow tubes and white ligulated flowers.	Brown.	0.99

NR: number of ramifications, **P_{100} A:** weight of 100 akenes.

The branching

Branching is the development of axillary buds in shoots. The number of branches is counted from the principal axis of the stem noted « T ». Two types of branching are found: a branching of type (T + 5) for the species : *Anacyclus clavatus* and *Chamaemelum fuscatum* at the mature plant stage and a Tertiary branching (T + 3) for the species *Leucanthemum parthenium*.

The stem

The main stem of *Anacyclus clavatus* and *Chamaemelum fuscatum* is often orthotropically developed. Plagiotropic development is sometimes observed. While their ramification has a plagiotropic development. The main stem and branch of *Leucanthemum parthenium* have a strictly orthotropic development.

Leaves

The leaves of *Anacyclus clavatus* are alternate, long, of average length equal to 3.19, short petiolated, bipennatized, acute at the tip and dark green at maturity. The leaves of *Chamaemelum fuscatum* are alternate, long, their mean length equal to 2.19, petiolate, bipennatized, containing a pointed end and green in color at maturity. The leaves of *Leucanthemum parthenium* are inserted in an alternate phyllotaxy. They are long, of average length equal to 4.36, petiolate, divided into narrow segments and yellowish-green at maturity.

The inflorescences and the flowers

The inflorescence of *Anacyclus clavatus*, *Chamaemelum fuscatum* and *Leucanthemum parthenium* is a flower head containing two types of flowers : yellow flowers tubulated in the center and white flowers ligated at the periphery. The flowers of the 3 species have the same floral biology, but show a difference in floral structure. Indeed, the liguled flowers of *Chamaemelum fuscatum* are long and beaked at the tip, while those of two other species are similar, they are short and more or less rounded.

The diameter of the receptacle varies from one species to another. It is 0.65 cm in *Leucanthemum parthenium*, 0.67 cm in *Chamaemelum fuscatum* and 1.56 cm in *Anacyclus clavatus*.

Fruit

The fruits differ from one species to another. The fruit of *Anacyclus clavatus* (Figure 4) is an indelible akene, beige at maturity, of rectilinear shape to flattened cone. This akene is surrounded by two membranous wings, clear, very thin, parchment and truncated at the apex. In the case of an akene without these wings, the fruit appears mottled and has four longitudinal ribs.

Figure 4 : Akene of *Anacyclus clavatus*

Figure 5 : Akene of *Chamaemelum fuscatum*

Figure 6 : Akene of *Leucanthemum parthenium*

The fruit of *Chamaemelum fuscatum* (Figure 5) is an indehiscent akene, very small, not marginated, flattened ovoid, raised by 3 ribs weak and finely striated. Their color is brown to yellow at maturity. The fruit of *Leucanthemum parthenium* (Figure 6) is an indehiscent akene, very small, brown at maturity, glandular and surmounted by a very short membranous crown and crenate.

Weight of 100 akenes

The mean weight of 100 akenes of *A. clavatus* is 45.23 mg, varies from 37.7 to 53.8 mg. For *C. fuscatum*, It is 26.63 mg and varies from 25.3 to 27.9 mg. An average weight of 9.96 mg was calculated in *L. parthenium*; For this species, the range of variation is 8.5 to 10.9 mg.

3.2.2. Analysis of morphological variability

The evaluation of a collection of genetic resources is commonly based on the simultaneous examination of many populations for various morphological characters. In this context, data on the different morphological traits measured were:

• An analysis of variance with one classification criterion followed by a comparison of means.

• An estimate of the degrees of association between the different traits studied by the Pearson correlation coefficient [25].

• A principal component analysis (PCA) based on the derivation of orthogonal variables [26].

3.2.2.1. Analysis of variance

The analysis of variance with one classification criterion (species effect) showed highly significant differences between the three species studied (Table 2) for the majority of the quantitative traits measured such as: Length of the longest branch (LLR), Length of the smallest branch (LPR), number of secondary branches (NRS), number of primary branches (NRP), mean leaf spine length (LMRF), average number of leaflets (NMf), mean diameter of the receptacle (DMR), length of the main root (LRP), mean length of the tertiary branch (LMRT), average length of secondary branch (LMRS), average length of primary branch (LMRP) and length of the main axis (LAP). This species effect is only significant for the number of leaves (NF). The difference between the three species is not significant for : The number of the principal axis head ligules (NLCAP), the average number of ligules per capitule (NML) and the number of tertiary branches (NRT). This result reflects a phenotypic heterogeneity between the 3 species studied, taking into account the measured parameters.

Table 2: Results of the variance analysis of the 17 morphological traits measured.

Characters	ddl	Average square	F_{obs}	Pr › F
LAP	2	3730,630	68,058	< 0,0001 **HS**
LMRP	2	982,641	26,382	< 0,0001 **HS**
LMRS	2	862,412	52,589	< 0,0001 **HS**
LMRT	2	360,894	26,359	< 0,0001 **HS**
LRP	2	40,961	11,73	0,000 **HS**
NF	2	338256,13	5,355	0,011 **S**

DMR	2	2,701	108,846	< 0,0001 **HS**
NMf	2	150,633	75,039	< 0,0001 **HS**
LMRF	2	11,796	36,769	< 0,0001 **HS**
NI	2	30601,433	2,983	0,068 **NS**
NRP	2	185,633	14,312	< 0,0001 **HS**
NRS	2	14770	15,244	< 0,0001 **HS**
NRT	2	4548,433	0,867	0,432 **NS**
NML	2	226,9	1,258	0,3 **NS**
NLCAP	2	0,7	1,086	0,352 **NS**
LPR	2	15,74	22,619	< 0,0001 **HS**
LLR	2	935,217	8,415	0,001 **HS**

F_{obs} : F observed ; **HS**: highly significant; **S**: significant ($P < 0.05$) ; **NS**: no significant ($P \geq 0.05$).

3.2.2.2. Comparison of means

According to the results obtained by analyzing the differences between the means with a 95% confidence interval, we distinguish 5 types of groups (Table 3). Comparison of means shows that *A. clavatus* and *C. fuscatum* are distinctly different from *L. parthenium* for : the length of the main axis (LAP), the mean length of the secondary branch (LMRP), the average length of the tertiary branch (LMRT), Root length (LR), number of leaves (NF), number of primary branches (NRP) and number of secondary branches (NRP).

A. clavatus is distinguished from *L. parthenium* and *C. fuscatum* for the mean diameter of the receptacle (DMR), the length of the smallest branch (LPR) and the length of the longest branch (LLR). In fact, the three species did not differ significantly in the mean diameter of the receptacle (DMR), the length of the smallest branch (LPR) and the length of the longest branch (LLR).

The parameters discriminating the three species are : the average length of the primary branch (LMRP), the mean number of leaflets per leaf (NMf) and the average length of the spine (LMRF). For the number of inflorescence per plant (NI), *Anacyclus clavatus* is not significantly different from *Chamaemelum fuscatum* or *Leucanthemum parthenium*. Therefore, *Anacyclus clavatus* and *Chamaemelum fuscatum* are much alike for more than half the morphological characters studied. Most of the highest averages of the morphological traits are observed in *Anacyclus clavatus*, while the majority of the lowest averages are observed in *Leucanthemum parthenium* (Table 3).

Table 3: Comparison of means of the 3 species studied.

Traits	*Anacyclus clavatus*	*Chamaemelum fuscatum*	*Leucanthemum parthenium*
LAP	19,8 B	20,71 B	53,7 A
LMRS	20,68 A	17,91 A	3,39 B
LMRT	12,12 A	12,5 A	1,91 B
LR	8,1 B	7,72 B	11,4 A
NF	629,5 A	524,5 A	271,7 B
NRP	11,4 B	11,9 B	19,1 A
NRS	39,6 A	29,6 B	100,6 A
DMR	1,56 A	0,67 B	0,65 B
LPR	3,21 A	1,4 B	0,8 B
LLR	46,69 A	29,97 B	29,91 B
NRT	53,7 A	37,3 A	79,6 A
NML	11,7 A	19,9 A	11,6 A
NLACP	13,3 A	13,4 A	12,9A
LMRP	36,12 A	24,34 B	16,42 C
NMf	15,6 A	10,9 B	7,9 C
LMRF	4,36 A	3,19 B	2,19 C
NI	116,5 A and B	82,4 B	190,6 A

3.2.2.3. The Matrix of correlation coefficients

The matrix of correlation coefficients between the characters studied (Table 4) shows: A positive correlation of the following traits: LMRP and LMRS correlate positively with each other and with all the parameters of LMRT, NF, DMR, NMF, LPR and LLR ; The parameters DMR and NMF correlate positively with each other and with LPR and LLR ; The character LAP is strongly correlated positively with the parameters LR, LMRF, NI, NRP and NRT ; LMRT correlates positively with NF and NMF ; LR correlates positively with the parameters NMF, LMRF, NI, NRP and NRS ; A positive correlation between the parameters NF and NMF, NRT and LLR ; A highly significant positive correlation between LMRF with NI, NRP and NRS ; NI correlates strongly with the parameters : NRP, NRS and NRT and weakly with LLR ; NRP is strongly correlated with NRS and weakly correlated with the characters NRT and LPR. A strong positive correlation is noted between NRS and NRT. A weak positive correlation is noted between NRT and LLR.

The LAP has a highly significant negative correlation with the parameters (LMRS, LMRT, NMF) and significant with the characters (LMRP, NF, DMR, LPR) ; LMRS and LMRT correlate negatively with LR, LMRF, NRP and NRS ; NMF is significant negatively correlated with NRP and NRS ; LR correlates positively and significantly only with NMF ; DMR is significant negatively correlated only with NRP (Table 4).

It is important to note that NLCAP and NML are not correlated with any of the other characters and that LMRP is the most positively correlated with the other traits (Table 4).

3.2.2.4. Principal component analysis

The graphical representation of the individuals dispersion of the 3 species studied reveals a more or less homogeneous grouping of the species studied forming 3 clear groups (Figure 7).

Indeed, there is a slight overlap between the two groups : *Anacyclus clavatus* and *Chamaemelum fuscatum*, whereas, *Leucanthemum parthenium* group seems very far from the others. These results confirm those of the variance analysis which showed a strong resemblance between *Anacyclus clavatus* and *Chamaemelum fuscatum*.

It is also observed that the individuals of the species *Chamaemelum fuscatum* occupy a rather restricted part of the plane and are located entirely in the negative part of the two axes F1 and F2. While, the individuals belonging to *Anacyclus clavatus* are scattered on the two axes (F1 and F2) with a trend towards the positive values of the F1 axis (Figure 7).

Furthermore, individuals of *Leucanthemum parthenium* are the best dispersed on the 2 axes (F1 and F2) with a tendency towards the negative values of F1 axis (Figure 7).

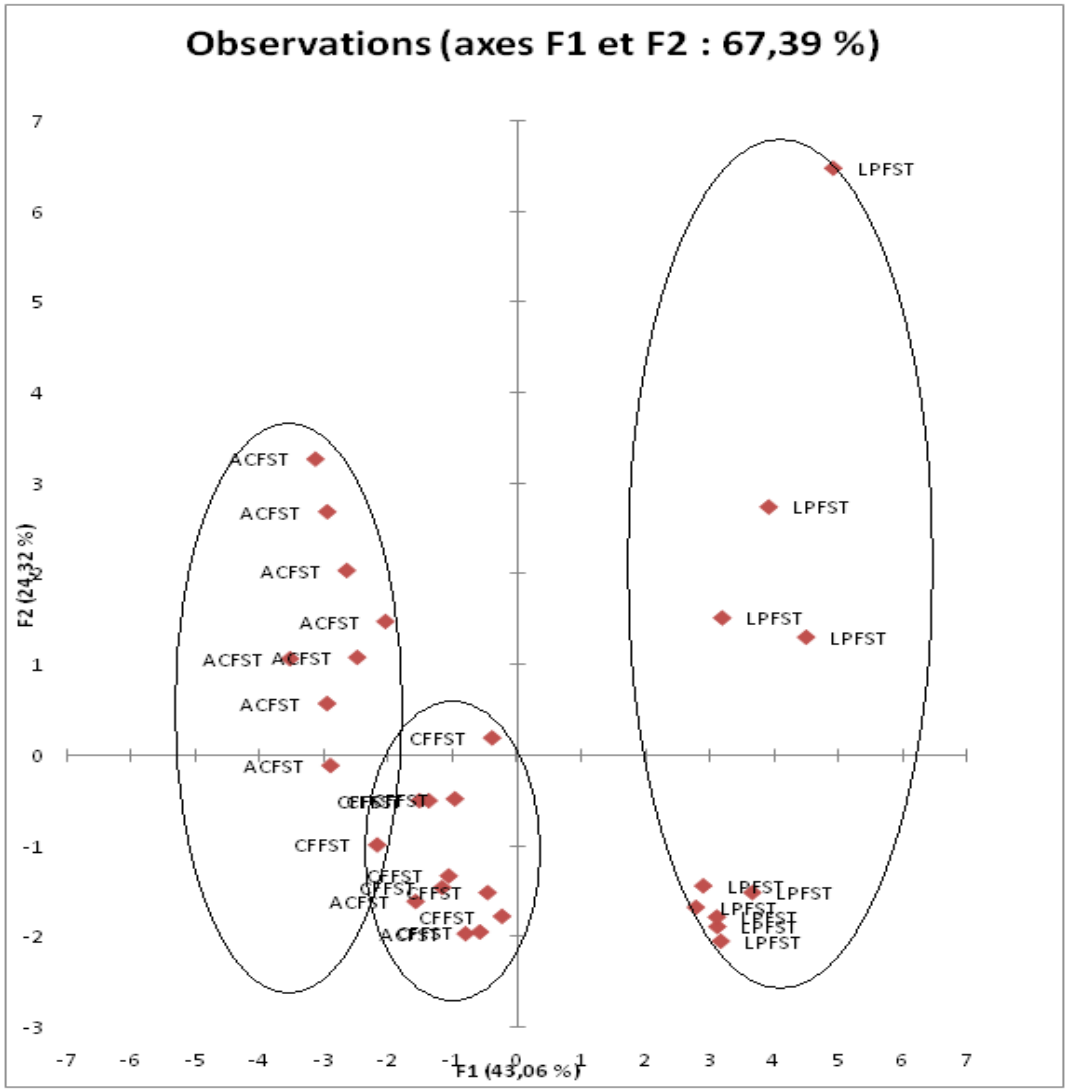

Figure 7 : Dispersal of the 3 species individuals in the plan defined by the axis 1 and 2 of the ACP.

Table 4 : Matrix of correlation coefficients of the different morphological parameters .

Traits	LAP	LMRP	LMRS	LMRT	LR	NF	DMR	NMf	LMRF	NI	NRP	NRS	NRT	NML	NLCAP	LPR	LLR
LAP	1																
LMRP	-0,536	1															
LMRS	-0,810	0,842	1														
LMRT	-0,766	0,707	0,918	1													
LR	0,607	-0,270	-0,572	-0,541	1												
NF	-0,388	0,797	0,763	0,764	-0,281	1											
DMR	-0,451	0,679	0,496	0,315	-0,290	0,271	1										
NMf	-0,670	0,691	0,662	0,522	-0,511	0,423	0,798	1									
LMRF	0,780	-0,266	-0,629	-0,677	0,451	-0,269	-0,048	-0,283	1								
NI	0,532	0,220	-0,123	-0,143	0,417	0,377	-0,176	-0,195	0,523	1							
NRP	0,803	-0,291	-0,579	-0,594	0,575	-0,135	-0,410	-0,572	0,673	0,628	1						
NRS	0,826	-0,119	-0,494	-0,461	0,603	0,014	-0,314	-0,455	0,701	0,872	0,774	1					
NRT	0,410	0,329	-0,007	-0,004	0,303	0,462	-0,104	-0,080	0,373	0,946	0,473	0,798	1				
NML	-0,130	0,052	0,090	0,095	0,014	0,269	-0,160	-0,031	-0,136	0,025	0,171	-0,172	0,048	1			
NLCAP	-0,179	0,262	0,282	0,325	-0,357	0,254	0,153	0,161	-0,267	0,006	-0,058	-0,016	0,075	-0,020	1		
LPR	-0,529	0,576	0,492	0,378	-0,385	0,289	0,762	0,787	-0,142	-0,224	-0,478	-0,387	-0,153	-0,114	0,247	1	
LLR	-0,184	0,868	0,597	0,465	-0,051	0,722	0,541	0,485	0,088	0,495	0,058	0,248	0,526	-0,094	0,194	0,396	1

In fact, the morphological study of [27] revealed that the analysis of variance showed variations among the 33 accessions of *Ricinus communis* L. from Andaman and Nicobar Islands for all the 18 traits studied. This work reveals also that plant height exhibited high significant positive correlations with the number of nodes on the main stem. In addition, The cluster analysis based on morphological traits grouped the 33 accessions of *Ricinus communis* L. into two major clusters [27].

Furthermore, the study of [28] was found a significant amount of genetic variability for all the twenty morphological parameters studied among safflower germplasm. In addition, this work reveals that seed yield plant had high significant and positive correlation with branches plant, capitulum plant, seeds capitulum and 100 seed weight. Furthermore, the hierarchical cluster analysis based on agro-morphological parameters divided 121 accessions of safflower into 5 main clusters [28].

The morphological study of [29] in rice varieties showed high phenotypic variability ($P < 0.0001$) for the characters : leaf length and leaf width, primary branching, maturity and grain thickness. In addition, this work revealed a positive and strong correlation (0.77) between the height at maturity and leaf length. The cluster analysis of this morphological study based on Euclidian distances between the 98 genotypes of Rice has allowed to identify three major clusters.

4. Conclusion

The comparative study of phenology and morphology diversity carried out on 3 species of Asteraceae (*Anacyclus clavatus*, *Chamaemelum fuscatum* and *Leucanthemum parthenium*) leads us to the following conclusions: The results of the phenological cycles follow-up show that the 3 species studied have distinct phenologies. The longest phenological cycle is observed for *Leucanthemum parthenium.*

The results of the variance analysis showed significant differences to highly significant for the majority of the traits studied. The comparison of means and the principal component analysis reveals that *Anacyclus clavatus* and *Chamaemelum fuscatum* form a single group for most of the traits measured while *Leucanthemum parthenium* is clearly distinct from these two species.

References

1. Mulik NG, Bhosale LJ. Flowering phenology of the mangroves from the West Coast of Maharashtra. J Bom Nat Hist Soc. 1989 ; 86 (3): 355 - 359.

2. Suresh HS, Sukumar R. Vegetative phenology of tropical montane forests in the Nilgiris, South India. Jour Natio Sci Founda Srilan. 2011 ; Vol 39, no 4 : 333 - 343.

3. Rout GR, Sahoo DP, Aparajita S. Studies on inter and intrapopulation variability of *Pongamia pinnata*: a bioenergy legume tree. Crop Breed Appl Biotechnol. 2009 ; 9: 268 - 273.

4. Zhang G, Song Q, Yang D. Phenology of *Ficus racemosa* in Xishungbanna, South West China. Biotropica. 2006 ; 38: 334 - 341.

5. Martins SR, Vences FJ, Miera LE, Barrosa MR, Carnide V . RAPD analysis of genetic diversity among and within Portuguese landraces of common white bean (*Phaseolus vulgaris* L.). Sci Horti. 2006 ; 108 : 133-142.

6. Balkaya A, Ergun A . Diversity and use of pinto bean (*Phaseolus vulgaris*) populations from Samsun, Turkey. N. Z. J. Crop Horti Sci. 2008 ; 36 : 189-197.

7. Shinwari S, Akbar F, Rabbani MA, Mumtaz AS, Shinwari ZK . Evaluation of genetic diversity in different genotypes of *Eruca sativa* from Pakistan by SDS-PAGE Analysis. Pak Jour Botan. 2013 ; 45 : 1235 - 1240.

8. Ghafoor A. Genetic diversity and gene-action in *Vigna mungo* based on morphological and biochemical markers. Ph D thesis : Quaid-i-Azam University. Islamabad, pp. 192 ; 1999

9. Demol J, Baudoin JP, Louant BP, Maréchal R, Mergeai G, Otoul E . Plant breeding: application to the main species grown in tropical regions. Ed presses of Gembloux, pp. 583; 2001.

10. Abdi A, Bekele E, Asfaw Z, Teshome A. Patterns of morphological variation of Sorghum (*Sorghum bicolor* (L.) Moench) landraces in qualitative characters in North Showa and South Welo, Ethiopia. Hereditas. 2002; 137: 161-172.

11. Fufa H, Baenziger PS, Beecher BS, Dweikat I, Graybosch RA, Eskridge KM . Comparison of phenotypic and molecular marker-based classifications of hard red winter wheat cultivars. Euphytica. 2005 ; 145: 133-146.

12. Mal TK, Doust JL. Phenotypic plasticity in vegetative and reproductive traits in an invasive weed, *Lythrum salicaria* (Lythraceae), in response to soil moisture. Amer Jour Botan. 2005 ; 92 (5) : 819 - 825.

13. Ducombs G. Lactones sesquiterpéniques et plantes. Rev Fr Allergol. 1999 ; 39 (4) : 295- 298.

14. Bonnier G. La grande flore, Tome 3. Ed. Berlin, pp. 676; 1990.

15. Pottier AG. Flore de la Tunisie: Angiospermes-Dicotylédones, Gamopétales. Programme Flore et Végétation Tunisienne, pp. 651-1189 ; 1981.

16. Schweizer D, Ehrendorfer F. Giemsa banded Karyotypes, Systematics, and Evolution in *Anacyclus* (Asteraceae-Anthemideae). Plant Syst Evol. 1976 ; 126: 107- 148.

17. Le Floc'h E. Contribution à une étude ethnobotanique de la flore Tunisienne. Programme Flore et Végétation tunisienne : Ministère de l'Enseignement Supérieur et de la recherche Scientifique, pp. 387 ; 1983.

18. Vogt R, Oberprieler C. Chromosome numbers of North African phanerogamsVIII. More counts in Compositae. Willdenowia. 2008; 38: 497- 519.

19. Ameddah S, Menad A, Dendougu H, Meraihi Z, Benayache S, Benayache F . Studies in Hepatocyte activity and the structure relationships and Luteolin Derivatives from *Chrysanthemum fuscatum*. Ecam. 2007; 4(S1): 55- 58.

20. Judd WS, Cambell CS, Kellogg EA, Stevens P . Botanique systématique. 1ère Edition, pp. 612 ; 1999.

21. Besharati-Seidani A, Jabbari A, Yamini Y, Saharkiz MJ. Rapid Extraction and analysis of volatile organic coumpounds of Iranian feverfew (*Tanacetum parthenium*) using headspace solvent

microextraction (HSME), and gas chromatography / mass spectrometry. Flav Fragr Jour. 2006; 21: 502 - 509.

22. Castleman M. Les plantes qui guérissent: Le guide le plus complet sur le pouvoir curatif des remèdes offerts par la nature. Ed. Rodale, pp. 518 ; 1991.

23. Afonso A, Castro S, Loureiro J, Mota L, De Oliveira JC, Torices R . The effects of achene type and germination time on plant performance in the heterocarpic *Anacyclus clavatus* (Asteraceae). Amer Jour Botan. 2014 ; 101 (5) : 1- 7.

24. Kaur G, Pal Singh B, Kaur Nagpal A. Phenology of Some Phanerogams (Trees and Shrubs) of Northwestern Punjab, India. Jour Botan. 2013; Article ID 712405: 1- 10.

25. Clifford HT, Stephenson W. An introduction to numerical classification . Academic Press. New York, USA; 1975.

26. Hair JF, Anderson RE, Tatham RL, Black WC. Multivariate data analysis. Macmillan. New York, USA; 1992.

27. Kanti M, Anjani K, Usha Kiran B, Vivekananda K. Agro-morphological and Molecular Diversity in Castor (*Ricinus communis* L.) Germplasm collected from Andaman and Nicobar Islands, India. Czech J Genet Plant Breed. 2015; 51 (3): 96 - 109.

28. Arif M, Hussain I, Rabbani MA, Ali S, Ali N, Khan SM, Zaheer Tanoli MT, Raza H. Assessment of genetic diversity among safflower germplasm through agro-morphological traits. Intern Jour Biosci. 2016; Vol 9 No 4: 1 - 11.

29. Moukoumbi YD, Sié M, Vodouhe R, Ndri B, Toulou B, Ogunbayo SA, Ahanchede A. Assessing phenotypic diversity of interspecific rice varieties using agro-morphological characterization. Jour Plant Breed Crop Sci. 2011 ; Vol 3(5) : 74 - 86.