

Growth responses of Clariid Catfish Hybrid (*Clarias gariepinus* ♀ X *Heterobranchus bidosarlis* ♂) fingerlings fed dietary Lablab Bean Meal (*Lablab Purpureus*)**Abstract**

This study examined the replacement of soyabean meal (SBM) with Lablab bean meal (LBM) in the practical diets of Clariid catfish (*Heteroclaris*). Five Iso-nitrogenous diets containing varying levels of LBM were incorporated as a non-conventional feedstuff at D₁(10% LBM); D₂(20%LBM); D₃(30%LBM); D₄(40%LBM) and D₅(50% LBM) as a replacement for soyabean meal. Seventy five (75) *Heteroclaris* fingerlings with an initial mean weight of 1.46±0.01g were stocked randomly to five treatments in triplicate groups and were fed to satiation twice daily for a period of 70 days. At the end of the 70days experimental period all growth parameters decreased across all diets from D₁ to D₅. D₁(10% LBM) had the best growth rate as it recorded the highest value in terms of weight gain (1.25); feed intake (2.34); feed conversion ratio (1.86); relative weight gain (4.86) and specific Growth rate of (1.33) while D₅ recorded the least values across all parameters. D₁ was not significantly different (P>0.05) from D₂ in all growth parameters listed above but D₁ and D₂ were significantly different (P<0.05) from D₃, D₄ and D₅.

Keywords: Lablab meal, soyabean replacer, *Heteroclaris* and non-conventional feedstuff

Introduction

Nutrition is one of the characteristics of all living things, fish inclusive. Organisms are fed in order to metabolize the energy stored in food into chemical energy used in maintaining their body. Nutrition is the synopsis of all the process whereby an organism provide with those material necessary for energy release, growth and repair, for its various secretion, for storage and for maintenance of internal osmotic and pH of the environment (Falayi, 2009). In fish farming, nutrition is critical because feed represents 60-70% of the production cost (Houlihan *et al.*, 2001). Low quality fish feed and its attendant high cost is the major factor limiting the development of aquaculture in Africa (Jamu and Ayinla, 2003).

In recent years, the use of grain legumes in the diet formulation has received considerable attention due largely to its ready availability, low cost and high amino acid composition (Adeparusi, 2001) compared to conventional fish meal which is scarce and expensive. Lablab bean (*Lablab purpureus*), a legume high in crude lignin and protein, has nutrient density compared to common beans (Aletor, *et al.*, 1989), but is grossly under-utilized in Nigeria. Lablab bean originated in India and has been widely distributed to many tropical countries where it is grown as an annual or a short-lived perennial. The seeds and immediate pods are used as human food, while the herbage is used as green manure, for erosion control and as feed supplement for cattle grazing. The use of indigenous legumes in diet formulation is generally limited by the presence of anti-nutritional factors –tannin, phytates, saponin, and trypsin inhibitor (Borget, 1992). Consumption of feeds containing these factors reduces nutrient utilization, feed efficiency and animal productivity. At high levels of intake, toxicity ensues and animal sometimes die (Makker, 1994). The activity of these compounds can be reduced by dehulling, soaking, cooking, toasting and fermenting (Deka *et al.*, 1990). *Heteroclaris* which is used in this investigation is increasingly cultured in Nigeria because

48 of its remarkable fast growth (Aluko, 1998), the resistance to diseases and poor environment.
 49 The rapid increase in its market demand because of its fleshy and tasty body has added
 50 stimulus to the aquaculture sector to supplement the deficit in the needed sustainable
 51 production and supply.

52 **Materials and Methods**

53 The study was conducted in the wet laboratory of the Department of Aquaculture and
 54 Fisheries Management, Faculty of Agriculture, University of Benin, Benin-city, Edo state for
 55 Seventy days.

56 **Experimental Diets:** The lablab beans (LB) contains anti-nutritional factors such as tannins,
 57 phytate and trypsin inhibitors. The beans were toasted for about 25 minutes to destroy the
 58 presence of the anti-nutritional factors, which are readily destroyed by heat. The toasted
 59 beans were then ground finely to yield the lablab bean meal. Fishmeal, soybeans cake, corn
 60 meal, palm oil, Vitamin E-gel and bone meal were purchased from a retail outlet at Murtala
 61 Mohammed Way in Benin City. The composition of the experimental diets is shown in Table
 62 1.

63 **Table 1: Composition of the Experimental Diets**

| INGREDIENTS | D ₁ | D ₂ | D ₃ | D ₄ | D ₅ |
|--------------------------------|----------------|----------------|----------------|----------------|----------------|
| % replacement of Lablab | 10% | 20% | 30% | 40% | 50% |
| LBM | 10.00 | 20.00 | 30.00 | 40.00 | 50.00 |
| Fish crumbs (50% CP) | 25.40 | 25.40 | 25.40 | 25.40 | 25.40 |
| SBC (48.0% CP) | 42.00 | 32.00 | 22.00 | 12.00 | 2.00 |
| Yellow maize (9.5% CP) | 10.00 | 10.00 | 10.00 | 10.00 | 10.00 |
| Palm oil | 8.00 | 8.00 | 8.00 | 8.00 | 8.00 |
| Bone meal | 4.00 | 4.00 | 4.00 | 4.00 | 4.00 |
| Vitamin premix | 0.60 | 0.60 | 0.60 | 0.60 | 0.60 |

64 The various ingredients were measured accurately to their required quantity, after which they
 65 were homogenously mixed, finely pelleted and dried at the departmental fish farm.
 66 *Heteroclarias* fingerlings with mean weight of 1.46±0.01g were obtained from the nursery
 67 pond of the department.

69 **Feeding Trial:** The study was conducted in the wet laboratory of Department of Aquaculture
 70 and Fisheries Management, University of Benin, Benin City. Fifteen (15) rectangular plastic
 71 tanks, five (5) treatments in three (3) replicates measuring (30cm×36cm×52cm) were used.
 72 Each tank was filled up to 2/3 of its volume with bore-hole water attached to the laboratory.
 73 The fishes were weighed in batches of five into each of the experimental units replicated
 74 thrice for each treatment. They were fed twice daily to satiation to ensure maximum growth
 75 between 08:00hrs and 16:00hrs. Feeding was monitored for each unit to ensure that fishes
 76 were not underfed or overfed. The experimental units were cleaned by total changing of the
 77 water daily. All fishes tanks were weighed and counted weekly to determine growth and
 78 survival, also the weekly weighing of feed was also carried out. The data obtained from the
 79 feeding trials were tested for significant differences using one way Analysis of Variance
 80 (ANOVA) test and the means were separated using Duncan's Multiple Range Test, all at 5%
 81 level of significance.

82 **Parameters Monitored:** Data on feed consumed and weight gain were collected weekly for
 83 each unit from which the following performance parameters were evaluated.

84 1. Weight gain (WG) = $W_2 - W_1$ (g) Where; W_1 = initial weight

- 85 $W_2 = \text{final weight}$
- 86 2. Feed intake = Initial weight of feed – Final weight of feed
- 87 3. Specific growth rate per day (SGR) % = $\frac{\text{Loge } W_2 - \text{loge } W_1}{T_2 - T_1} \times 100$
- 88 Where: T_1 and T_2 are time of experiment in days.
- 89 $W_2 = \text{final weight at } T_2$
- 90 $W_1 = \text{initial weight at } T_1$
- 91 Loge = natural logarithm.
- 92 4. Relative weight gain (PWG) % = $\frac{\text{Weight Gain}}{\text{Initial Weight}} \times 100$
- 93 5. Food conversion ratio (FCR) = $\frac{\text{Feed Intake(g)}}{\text{Wet Weight Gain(g)}} \times 100$
- 94 6. Protein efficiency ratio (PER) = $\frac{\text{Weight Gain (g)}}{\text{Protein Intake}} \times 100$
- 95 7. Survival rate % = $\frac{\text{Initial stocked} - \text{mortality}}{\text{Initial stocked}} \times 100$
- 96

97 **RESULTS**

98 The water temperature of the experimental tanks containing the fish within the experimental
99 period was within the range of 26 °C to 28 °C and water PH at 6.9 to 7.8

100
101 **Table 2: Proximate Composition (%) Of Lablab Bean Meal (Lbm) and Experimental**
102 **Diets.**

| DIETS | Moisture | Ash | Fat | Fibre | Crude Protein | NFE |
|----------------|----------|-------|-------|-------|---------------|-------|
| LBM | 6.90 | 8.25 | 9.54 | 3.27 | 24.49 | 47.83 |
| D ₁ | 5.91 | 10.24 | 15.37 | 3.72 | 46.08 | 18.78 |
| D ₂ | 7.06 | 10.32 | 16.30 | 4.18 | 40.25 | 21.90 |
| D ₃ | 6.14 | 9.60 | 15.60 | 4.31 | 34.42 | 30.12 |
| D ₄ | 5.62 | 9.93 | 16.30 | 3.89 | 31.50 | 33.40 |
| D ₅ | 6.19 | 10.07 | 15.32 | 3.90 | 28.35 | 36.20 |

103
104 NFE= nitrogen-free extract. It was determined by subtracting the summation of the values of crude
105 protein, fat, fibre, ash and moisture from 100%

106
107 From the result above, the crude protein level of lablab meal is 24.49% with fat content of
108 9.54% and a crude fibre content of 3.27. Among the various diets incorporated with lablab
109 meal, D₁ with 10% incorporation level had the highest crude protein value (46.08) while D₅
110 had the least value (28.35) as the crude protein level reduced with increase in corporation
111 percentage of lablab meal.

112 **4.1 Growth and Feed Utilization Parameters**

113 **Table 3. Growth performance and feed utilization of Clariid catfish, (*Heteroclaris*) to**
114 **lablab bean meal (LBM) based diet.**

| PARAMETERS | Treatments | | | | | SEM |
|------------|----------------|----------------|----------------|----------------|----------------|-----|
| | D ₁ | D ₂ | D ₃ | D ₄ | D ₅ | |
| | 10% | 20% | 30% | 40% | 50% | |
| | | | | | | |

| | | | | | | |
|-----------------------------|--------------------|--------------------|--------------------|--------------------|--------------------|------|
| Weight gain(g) | 1.25 ^a | 1.17 ^a | 1.08 ^a | 0.77 ^b | 0.37 ^c | 0.30 |
| Specific Growth Rate(%/day) | 1.33 ^a | 1.16 ^a | 1.08 ^b | 0.46 ^c | 0.33 ^c | 0.70 |
| Relative Weight Gain (%) | 4.86 ^a | 3.64 ^a | 1.67 ^b | 0.87 ^c | 0.63 ^c | 4.58 |
| Protein Efficiency ratio | 59.12 ^a | 29.00 ^b | 26.76 ^b | 14.18 ^c | 14.03 ^c | 50.0 |
| Feed Intake(g) | 2.34 ^a | 2.17 ^a | 2.08 ^a | 1.51 ^b | 1.63 ^b | 1.76 |
| Feed Conversion Ratio | 1.87 ^a | 1.86 ^a | 1.93 ^a | 1.96 ^a | 4.41 ^b | 8.75 |

115 **N/B: Mean Values with the same superscript on the same row are not significantly**
 116 **different, (P> 0.05)**

117 Result showed that Weight gained by *Heteroclaris* fingerlings after ten weeks was not
 118 significantly different (P> 0.05) in D₁(1.25), D₂(1.17) and D₃(0.77) while D₅(0.37) was
 119 significantly depressed (P< 0.05) with least weight gain value (0.37).

120 The Specific growth rate was significantly higher (P<0.05) in D₁(1.33) and D₂(1.16) than D₅
 121 (0.33) with the least Specific growth rate.

122 The feed conversion ratio (FCR) recorded was an indication that food was converted to flesh at different rate.
 123 The best FCR value was reported in D₁ (1.87) while the D₅ had the worst value (4.41).

124 Fish fed with 10% LBM was superior in terms of relative weight gain to other diets.
 125 However, the Relative Weight gain was not significantly different (P>0.05) between D₁(4.86)
 126 and D₂(3.64) while D₅(0.63) had the least relative weight gain.

127 The Protein Efficiency Ratio for D₁(59.12) was significantly different (P<0.05) from all other
 128 treatments, while D₅(14.03) had the least protein efficiency ratio.

129 Feed Intake by fish amongst D₁(2.34), D₂(2.17) and D₃(2.08) were not significantly different
 130 (P>0.05), while D₅(1.63) was significantly depressed. However, Fish fed with 10% LBM
 131 recorded the highest amount of feed intake

132 **Table 4: Carcass composition of the experimental fish (%)**

| DIETS | Crude protein | Fat | Ash | MC | NFE |
|------------------------|---------------|-------|-------|------|-------|
| Fish (initial) carcass | 50.17 | 13.49 | 10.21 | 5.15 | 21.00 |
| TF ₁ | 53.08 | 12.32 | 10.22 | 5.26 | 19.12 |
| TF ₂ | 66.50 | 11.22 | 10.22 | 5.15 | 6.91 |
| TF ₃ | 44.92 | 12.12 | 9.80 | 5.10 | 28.22 |
| TF ₄ | 62.42 | 11.59 | 9.74 | 5.21 | 11.04 |
| TF ₅ | 63.00 | 12.87 | 10.07 | 5.11 | 8.95 |

133 **MC= moisture content, NFE= nitrogen-free extract, TF= Test Fish**

134 Table 4 show the carcass composition of experimental fish after being fed with experimental
 135 diets for ten weeks. From the result, the crude protein level of the initial carcass was 50.17%
 136 and this increased with treatment level but however decreased at TF₃. Fishes fed with
 137 20%LBM incorporation had the highest level of crude protein (66.50%) in their carcass while
 138 TF₃ with 30% inclusion level had the least crude protein value of 44.92%.

139

140 **DISCUSSION**

141 The analyzed crude protein of the lablab bean meal in this study was 24.49%. This falls
 142 within the range of 20.46-25.47% reported by Kamatchi *et al.* (2010) and also between 20-
 143 28% reported by Cook *et al.*(2005). The fat content value of 9.54% reported in this study was
 144 higher than the 2.69-4.17% reported by Kamatchiet *al.* (2010), it was also higher than the
 145 report of Aliu *et al.*(2014) which reported a low fat content of 5.45%. The high fat content
 146 reported on the Lablab meal may have led to the higher concentration of fat in the fish
 147 carcass as the fat content exceeds the maximum inclusion level of 8% in a normal catfish
 148 diet. The ash content was also higher than the ash content of 3.97-4.48% reported by
 149 Kamatchiet *al.* (2010).

150 The growth rate varied with different inclusion level of lablab bean meal. This variation in
 151 growth rate that was highest in *lablab* may be related to anti nutritional factor(s) present in
 152 the seeds such as trypsin inhibitors, proteins inhibitors and phytic acid (Robinson, 2001). It is
 153 a common knowledge that heat treatment is known to detoxify anti-nutrients but affects
 154 growth response, the retarded growth and nutrients utilization recorded in this study was in
 155 line with the findings of Sotolu and Faturoti (2008), who reported that heat treated *leucaena*
 156 seeds gave lower performance than *leucaena* soaked in water and sundried. Toasting of the
 157 seeds could have also resulted in the destruction of the amino acid bonds thereby reducing the
 158 protein quality of the feed ingredients. This was supported by Osuigwe *et al.*(2005) who
 159 reported that heating destroys and reduces nitrogenous compounds in legume seeds.

160 Protein efficiency ratio (PER) was highest in fish fed with 10% LLBM meal. This is in
 161 conformity with what was stated by Sotolu and Faturoti (2009), who reported that similarity
 162 in the PER of *Clarias gariepinus* has a direct link with feed intake. All diets produced
 163 different values of fish carcass protein and lipid than initial values with marginal difference
 164 among them indicating different retention and utilization levels of the diets. This is in line
 165 with Alegbeleye *et al.*(2001) who reported that effective utilization of bambara groundnut at
 166 different inclusion levels was responsible for variations in *Heteroclaris* carcass protein and
 167 lipid.

168 The lower the FCR of a feed, the higher the efficiency of the feed. D₁ had the best feed
 169 conversion ratio while D₅ had the poorest FCR. There was an increase in the FCR as the
 170 LBM inclusion level increased and this could be attributed to low feed utilization, low
 171 digestibility and the presence of anti-nutrients which is in line with Scott *et al.* (1976) who
 172 stated that fish decreased digestibility is caused mainly by increased cumulative residual
 173 effect of anti-nutritional factors.

174 There is reduction in the feed intake as the percentage of LBM increased. This is in line with
 175 Ragab *et al.* (2013) who reported similar reduction in feed intake with increased level of
 176 legume concentrate. This reduction was attributed to unpalatable residual effect of the anti-
 177 nutrients which increased with the dietary level of test feedstuff (hyacinth bean).

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