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3 **Impact of papaya seed soaking in different BA,**
4 **colchicine and EMS solutions on germination,**
5 **growth and chromosomal behaviour**

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9 **ABSTRACT (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)**

10 The present investigation was carried out during two consecutive seasons 2015 and 2016 in fruit nursery of faculty of Agriculture at Moshtohor, Benha University, in order to throw some spotlight on the impact of some chemical substances (Ethylmethanesulphonate – EMS in 10, 20 and 30 ppm); (colchicine at concentrations of 1%, 2% and 3%) and (benzyl adenine – BA, at concentrations of 1,2% and 3%) on seed germination %, seed germination rate, some seedling growth measurements and cytological examination of root tip of *Carica papaya* cv. Solo. The treatments were arranged in complete randomized block design with nine replicates (polyethylene bags), however, each replicate was represented by two papaya seedlings. The seedlings were divided into three categories according to their growth vigor, each category represented by three replicates for each treatment and subsequently each category sampled by 60 seedlings for all studied treatments. Seedling growth and chromosomal behavior as imported by the three studied chemical substances were evaluated on the 1st week of December. Data obtained revealed that both BA at 2% and BA 3% increased significantly germination %, germination rate and growth measurements. On the contrary, the least significant increase was always in concomitant to EMS at 3% and colchicine at 3 % during both experimental seasons. Moreover, EMS was more inhibitor of cell division followed by BA than Colchicine. This may be due to more damage resulted by BA and EMS affected on DNA replication during mitosis.

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12 *Keywords: [Carica papaya, germination %, seed germination rate, growth measurements, cytological*
13 *examination, BA, Colchicine and EMS.]*

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16 **1. INTRODUCTION (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)**

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18 The papaya (*Carica papaya* L.) is cultivated for its ripe fruits, favored for people in the tropical region as breakfast fruit, and as ingredient in juice, jellies and preserves or cooked with young leaves and shoots as a vegetable plant. The fruit contains high level of papain; the proteolytic enzyme used for medical purposes and as a tenderizer for meat. The fruit, also, contains considerable quantities of vitamin A, B and C and about 10% sugar. Fruits and seeds extract have pronounced bactericidal activity against *Staphylococcus aureus*, *Bacillus cereus* and *Escherichia coli* and the latex is used to remove freckles. Other parts such as bark, are used for making rope while leaves are also used as a soap substitute supposed to remove stains.

25 Cytokinins can alter flower sex ratio in species with imperfect flowers. Cytokinins generally, increase the ratio of female flowers to male flowers which has implications for fruit production [1]. BA has also been used in the vegetable crop industry to alter flower sex ratios of monoecious and dioecious plants to increase the number of female flowers available to produce fruit [2]. Exogenous cytokinins can promote an accumulation of chlorophyll and promote the conversion of etioplasts into chloroplasts [3] even in dark grown seedlings. This may appear as a greening effect on ornamental crops which may be perceived as an increase in quality in green leaved crops and a decrease in quality in crops with other leaf colors. There is also some evidence that cytokinins can help increase the flower size of some plants. Cytokinins increased the size of petunia flowers [4]. In ferns however, cytokinins appear to induce maleness in the gametophytes [5].The

33 reduction in percentage of seed germination and survival was due to the disturbances caused at the physiological level
34 coupled with chromosomal damage. Disturbance in the formation of enzymes involved in the germination process may be
35 one of the physiological effects caused by mutagenic treatments particularly chemical mutagens [6].

36 Colchicine ($C_{22}H_{25}NO_6$), originally extracted from *Colchicum autumnale*, may induce some morphological, cytological and
37 histological changes, and even changes in the gene expression level [7]. Chemical mutagens such as ethyl methane
38 sulfonate (EMS), a compound of the alkaline sulfonate series, is most frequently used for chemical mutagenesis in higher
39 plants due to its potency and the ease with which it can be used [8]. It usually causes high frequency of gene mutations
40 and low frequency of chromosome aberrations [9].

41 The present investigation was planned and carried out to study the influence of some chemical substances i.e., (BA,
42 colchicine, ethyl methane sulphonate) at different concentrations on some seed germination parameters, some vegetative
43 growth measurements, as well as root till chromosomal behavior of papaya cultivar "Solo" through. the cytological
44 examination of papaya seedling.

45 **2. MATERIAL AND METHODS / EXPERIMENTAL DETAILS / METHODOLOGY (ARIAL, BOLD, 11 FONT,** 46 **LEFT ALIGNED, CAPS)**

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49 The present investigation was carried out during two consecutive seasons 2015 and 2016 in fruit nursery of
50 faculty of Agriculture at Moshtohor, Benha University, in order to throw some spotlight on the impact of some chemical
51 substances (Ethyl Methane Sulphonate – EMS; colchicine and benzyl adenine "(BA) on seed germination %, seed
52 germination rate, some seedling growth measurements and cytological examination of root tip of *Carica papaya* cv. Solo.

53 In this regard, mature papaya fruits were collected from the trees which grown at fruit farm of Faculty of
54 Agriculture, Moshtohor, Benha Univ., seed were extracted when the fruits have been ripened, and washed three times
55 with tap water to get rid of fruit pulp residual. Finally, seeds were kept in shading place to be dried and stored in small
56 coped glass contain calcium chloride to be ready for carrying out the investigation.

57 On the first week of March of both seasons, dried stored papaya seeds were soaked in tap water for 24h then
58 taken out and placed in shade for 10 minutes to dry. Those seeds were divided into ten groups. Each group represented
59 by two hundred seeds and subjected to one of the following treatments:

- 60 1- Soaking in tap water for 12 hours (control).
- 61 2- Soaking for 12 hours in benzyl adenine (BA) at 1 %.
- 62 3- Soaking for 12 hours in benzyl adenine (BA) at 2 %.
- 63 4- Soaking for 12 hours in benzyl adenine (BA) at 3 %.
- 64 5- Soaking for 12 hours in colchicine at 1 %.
- 65 6- Soaking for 12 hours in colchicine at 2 %.
- 66 7- Soaking for 12 hours in colchicine at 3 %.
- 67 8- Soaking for 12 hours in ethyl methane sulphonate (EMS) at 10 ppm.
- 68 9- Soaking for 12 hours in ethyl methane sulphonate (EMS) at 20 ppm.
- 69 10- Soaking for 12 hours in ethyl methane sulphonate (EMS) at 30 ppm.

70 The dried seeds were soaked in aqueous solutions of the three investigation chemical substances as well as
71 control seed were soaked in tap water for 12 hours. Those seeds were re-dried for 10 minutes in shade after soaking in
72 the investigation chemical substances and immediately sown on March 9th and 21st during 2015 and 2016 seasons,
73 respectively, in black polyethylene bags (30 cm in diameter) filled with a mixture of sandy and clay soil (1:1 v/v) and kept

under greenhouse conditions. The seeds were watered every other day in the morning till the appearance of plumule. Furthermore, fungicide was applied at the time of seed sowing as a tool protection against the fungal attack of *Rhizoctonia solani* and *Fusarium species*, as well as weeds were completely removed along with their roots as soon as they appear. The first appearance of plumule was recorded in the 1st week of April during both seasons of study.

The abovementioned ten investigated treatments were arranged in complete randomized design, where each treatment was replicated ten times (10 polyethylene bags) and each replicate represented by an individual polyethylene bag which contains twenty papaya seeds. Furthermore, the number of emerged seedlings was counted as soon as the appearance of first true leaves on the 4th week at April of three days' intervals until seed germination was completely ceased, then the following seed germination parameters were calculated:

$$1- \text{Germination percentage} = \frac{\text{Total number of emerged seedling}}{\text{Total number of planted seeds}} \times 100$$

2- Germination rate according to equation [10] :

$$\text{Germination rate} = \frac{A_1 T_1 + A_2 R_2 + A_3 T_3 + \dots A_n T_n}{A_1 + A_2 + A_3 \dots A_n}$$

T1 = Number of days passed from soaking till first count 1.

T2 = Number of days passed from soaking till second count to Tn.

A1 = Number of germinated seeds at first count.

A2 = Number of germinated seeds at second count to An.

3- Number of days required for germination completion.

In order to study the impact of the three investigated chemical substances on some seedling growth measurements and chromosomal behavior of sprouted papaya seedlings, thin out of un-desirable seedlings (the weakest and the strongest ones) was done on the first week of July, while the nearly uniform seedlings in their growth vigor were remained in the polyethylene bags.

The treatments were arranged in complete randomized blocks design with nine replicates (polyethylene bags), however, each replicate was represented by two papaya seedlings. The seedlings were divided into three categories according to their growth vigor, each category represented by three replicates for each treatment and subsequently each category sampled by 30 seedlings for all studied treatments.

Seedling growth and chromosomal behavior as impacted by the three studied chemical substances were evaluated on the 1st week of December through studding the following parameters:

A- Growth parameters: -

- 110 1- Seedling height.
111 2- Stem diameter (cm).
112 3- root length
113 4- Number of leaves/seedling.

114 **B -cytological studies: -**

115 Papaya (*Carica papaya* L.) seedling roots were used for bioassay. Papaya seeds were kindly supplemented from
116 the research farm of Faculty of Agriculture, Moshtohor, Benha University to be used in this study. Seeds were soaked in
117 three different concentrations of Benzyl adenine, EMS and Colchicine. Root meristem raised in water were fixed in a
118 fixative solution (3:1) and kept in alcohol 70 % in refrigerator until used for cytological examination.

119 About 100 cleaned papaya seeds were set up in petri dishes and soaked for 24 hours here in tap water here in 10
120 seeds were re-soaked in tap water and used as a control while the other 90 seeds were picked out and divided into 3
121 groups, each one contain thirty seeds and subjected to 1%, 2% and 3% of Benzyl adenine (BA), Ethyl methane sulphonate
122 (EMS) and Colchicine for 12 hours.

123 **Mutagenic agents:**

124 Ethyl methane sulfonate (EMS): The linear formula of EMS is $\text{CH}_3\text{SO}_3\text{C}_2\text{H}_5$. This formula was referred to the free
125 chemical database: (ChemSpider ID: 5887). Seeds before germination were subjected to the following concentrations;
126 1%, 2% and 3% for twelve hours. Benzyl adenine (BA) or 6-Benzylaminopurine (BA) is $\text{C}_{12}\text{H}_{11}\text{N}_5$. Cyclophosphamide
127 (Colchicine) at the concentrations of 1%, 2% and 3%. The linear formula of colchicine is $\text{C}_7\text{H}_{15}\text{C}_{12}\text{N}_2\text{O}_2\text{P}$.

128 **Fixation and storage solutions:**

129 Root tips of the germinated Papaya seeds in the different investigated substances and tap water as control were
130 excised and fixed in 1: 3 acidic alcohol consisted of a mixture of glacial acetic acid and ethanol respectively and later
131 preserved in 70 % ethyl alcohol.

132 Staining agent (acetocarmine).

133 A carmine stain was prepared at the concentration at 1% by dissolving it in 45% acetic acid. Before adding the
134 stain, root tips were put in a boiling acetocarmine for one minute for losing the tissue.

135 **Root collection and slide preparation**

136 Papaya seeds were germinated at lab temperature using petri dishes filled with enough tap water to top four to
137 five weeks for root tips to grow. Seeds subjected to treatments were transferred to each concentration of BA, EMS and
138 Colchicine after the length of the roots reached to 1-1.5 cm maximum. Roots were harvested at the morning. Root tips
139 excised from treated and controlled materials were fixed in 1: 3 acidic alcohols and preserved in 70% ethyl alcohol. Root
140 tips squashed were conducted using 1% Acetocarmine stain.

141 **Mitotic index (MI) determination:**

142 The slides were viewed under the light microscope, by using 40 objective lens. On one slide for each treatment
143 dividing cells (prophase, metaphase, anaphase and telophase) were counted to determine MI. MI was expressed as the
144 number of dividing cells per 1000 cells scored.

145 Chromosomal aberrations were characterized and classified in the following types: large chromosomal deletion or losing a
146 hole chromosome, sticky chromosomes, anaphase bridge chromosomes, lagging chromosomes, disrupted chromosome
147 segregation, star cluster chromosomes, clumped chromosomes in metaphase. These aberrations were saved in
148 photographic pictures.

149 **Statistical analysis:**

150 All the obtained data during each season of this study were subjected to statistical analysis of variance according
151 to the method described by [11]. However, the differences means were differentiated by using Duncan's multiple range
152 test [12].

154 **3. RESULTS AND DISCUSSION**

157 **Effect of seeds pre-sowing soaking in different BA, colchicine and EMS solutions on some germination** 158 **measurements.**

159 In this regard some germination measurements germination percentage and germination rate of papaya Solo cv.
160 in response to pre-sowing soak in some BA, colchicine and EMS solutions were investigated during 2015 and 2016
161 experimental seasons are presented in **Table (1)**.

162 **- Seeds germination percentage:**

163 Data presented in **Table (1)**, indicate that the seeds germination percentage of papaya "Solo" cv. after 4 weeks
164 from planting as influenced by their soaking for 12 hours in different BA, colchicine and EMS solutions significantly
165 increased during both experimental seasons. However, pre-sowing soak in the highest BA concentration surpassed
166 significantly than investigated treatments. On the other side, the least concentration of colchicine and EMS solutions at (1
167 %) showed significantly the highest increase over control during two experimental seasons. In addition, other pre-sowing
168 soak solutions (1% & 2%) of BA ranked statistically the second one. Moreover, BA as a growth promoter explain the
169 function for activating growth and germination particularly cell division.

170 **- Seeds germination rate:**

171 **Table (1)** reveals obviously that germination rate followed typically the same trend previously discussed with
172 germination percentage. Herein, all BA, colchicine and EMS solutions resulted in a significant increase over the tap water
173 soaked seeds (control) during both experimental seasons. The highest BA solution were statistically the superior, while
174 their lowest concentration (at 1% & 2%) ranked statistically second. In addition, tap water soaked seeds (control) was the
175 inferior such trend was true during 2015 and 2016 experimental seasons.

176 These results are in accordance with the findings of [13] reported that freshly extracted seeds of acid lime (*Citrus*
177 *aurantifolia* swingle) were shade dried and were soaked in 15, 30, 45 or 60 mM EMS solution for 12 hours caused
178 decrease of percentage seed germination (36%) with increasing of EMS concentrations to 60 mM. Despite, seeds of *L.*
179 *esculentum* cv. Roma, were treated with 0.1, 0.5 and 1% ethyl methane sulphonate (EMS) and exposed for 3 and 6
180 hours, decrease in seed germination was observed with increasing EMS% [14]. Papaya seeds treated with colchicine at

0.5% or 1.0% and EMS at 200 ppm and 100 ppm improved germination parameters compared with untreated seeds (control) [15]. A clear effect of different EMS-treated on seeds germination percentage of *L. esculentum* (cv. Pusa – Early–Dwarf) showed that germination percentage increased with increasing EMS concentrations from 0.0150% to 0.1205%. Thereafter, decrease in germination percentage was observed at the highest concentration (0.2410%) [16]. Addition colchicine to cultured medium of *Solidago altissima* at 125 mg/l had an inhibition, while the other treatments (low concentration of colchicines) possessed the most promotion influences on survival capacity of explants (75-100%) [17].

Seeds of water melon without coat during the seeding, nicking at radicle end with colchicine-treated showed high germination rates 84.3% and 77.1%, respectively [18]. The effect EMS and colchicine-treated seeds of Papaya at 0.1% and 0.5%, they found the stimulatory effects of low-dose colchicine treatment on seedling emergence and seed germination decreased with the increasing doses of colchicine [19]. Reduced seed germination due to the effect of increasing doses of chemical mutagens on the meristematic tissues of the seeds may be causing damage of cell constituents at a molecular level or to disturbance in the formation of enzymes involved in the germination process caused by EMS and colchicine. Impact of mutagenic treatments i.e., EMS-treated seeds at 0.25-0.30% of rice causing the reduction in percentage of seed germination and survival was due to the chromosomal damage and disturbance in the formation of enzymes involved in the germination process [20] and [6].

Table (1): Impact of papaya seed soaking in different BA, colchicine and EMS solutions on seed germination percentage and germination rate during 2015 & 2016 experimental seasons.

Parameters Treatments	Germination percentage %		Germination rate	
	First season	Second season	First season	Second season
1. control	55.67 g	54.33 h	3.68 i	3.42 i
2. BA at 1 %.	77.00 b	79.33 b	5.10 c	5.04 c
3. BA at 2 %.	80.67 a	81.00 b	5.32 b	5.24 b
4. BA at 3 %.	81.67 a	83.33 a	5.43 a	5.33 a
5. colchi at 1 %.	68.67 d	68.00 e	4.09 f	3.96 f
6. colchi at 2 %.	73.33 c	71.67 d	4.24 d	4.11 e
7. colchi at 3 %.	75.00 c	74.33 c	4.28 d	4.13 d
8. EMS at 10 ppm	61.00 f	63.67 g	3.75 h	3.57 h
9. EMS at 20 ppm	65.33 e	65.67 f	3.89 g	3.78 g
10. EMS at 30 ppm	65.67 e	67.33 ef	4.15 e	4.10 e

Means followed by the same letter/s within each column during every season are not significantly at 5 % level.

202 - **Impact of papaya seed soaking in different BA, colchicine and EMS solutions on some growth measurements during 2015**
203 **& 2016 experimental seasons.**

204 In this concern average seedling height, stem diameter, root growth and average number of leaves/seedling in
205 response to various treatments were investigated during two 2015 and 2016 experimental seasons are presented in
206 **Tables (2).**

207 **- Average seedling height (sm):**

208 Concerning the response of average seedling height to the differential treatments, it is quite clear as shown in
209 **Table (2)**, that all investigated treatments with various solutions from BA, colchicine and EMS resulted in an increase in
210 average seedling height of papaya "Solo" cv. translocated seedlings during both experimental seasons. Anyhow, the
211 increase was more pronounced with (BA at 3%) treated seeds, **descendingly** followed by BA at 2%, BA at 1%, colchicine
212 at 2% and colchicine at 3%. However, such increase was too few to reach level of significance either the investigated
213 treatments were compared each other's or to tap water soaked seeds (control) only with few exceptions particularly with
214 colchicine at 3% in the second season. Such trend of response was true during both 2015 and 2016 experimental
215 seasons.

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217 **-Seedling diameter (cm):**

218 Regarding the effect of different investigated treatments on stem diameter of papaya "Solo" cv. translocated
219 seedlings **Table (2)** displays obviously that both (T3 & T4) treatments of BA solutions at 2% and BA 3% induced
220 significantly the thickest stem. Such trend was true during two seasons of study. Moreover, (T10 and T2) treatments of
221 (EMS at 3 % and BA at 1 %), respectively, ranked statistically second as their effect on stem diameter was concerned for
222 papaya Solo cv. translocated seedlings during two experimental seasons. On the other side other investigated treatments
223 increased significantly the average stem thickness during both seasons of study but T8 (EMS 1 %) showed statistically
224 the least significant increase in stem diameter during 2015 and 2016 experimental seasons. In addition, other investigated
225 treatments were statistically in between the aforesaid two extremes during two experimental seasons.

226 Moreover, BA as a growth promoter explain the function for activating growth specially stem diameter by increase
227 cell division which gave more thickness for the stem.

228 **- Root length (cm):**

229 This is the response of root length to various investigated treatments during both 2015 and 2016 experimental
230 seasons, and data obtained during both seasons for papaya Solo cv. translocated seedlings are presented in **Table (2)**. It
231 is quite evident as shown from tabulated data that a noticeable grade of variance in trend of response could be observed
232 between investigated treatments in this concern. Anyhow, the greatest length of root was significantly in closed
233 relationship to BA at 3% during two seasons of study. Moreover, BA at 2% came statistically second. On the contrary, the
234 least significant increase in root length was always in concomitant to EMS at 3% and colchicine at 3% during 2015 and
235 2016 experimental seasons of study. In addition, other treatments were statistically in between the aforesaid two
236 extremes. Such trend was true during both seasons.

237 Moreover, the trend of response of root length of **seedling** may be attributed to the variance in biological and
238 physiological roles could be played by BA pertaining shoot growth and root length and development.

239 - Number of leaves/seedling:

240 With regard to the response of leaves number per seedling an individual seedling to the differential investigated
241 treatments, obtained data are presented in **Table (2)**. It is quite evident that the greatest leaves number of per seedling
242 was significant in closed relationship to such seedling was subjected to BA at 3% during 2015 and 2016 experimental
243 seasons. Moreover, BA at 2 % ranked statistically second. Anyhow, pre-sowing soaked in BA at 1 % solution ranked
244 statistically 3rd, descendingly followed by soaking in EMS 1%, EMS 2% and EMS 3% during both 2015 and 2016
245 experimental seasons. On the contrary, the least significant leaves number per seedling that exhibited by three
246 investigated treatments (colchicine at 3 %, control and colchicine at 2 %), respectively. Such trend was true during 2015
247 and 2016 experimental seasons. The seeds of two pea cultivars were treated with EMS at concentrations of 0.5%, 0.75%
248 and 1.0%. In M1-generation, number of branches decreased with EMS at 0.75% and 1.0% [21].

249 The cytokines promote shoot development through increased cell division, regulation of the cell cycle and the number of
250 cycles that **cells in** the meristems [22]. After addition of 20 mg/l colchicine into the medium for one week, induced
251 tetraploidy plants were subsequently added. Morphological observations showed that the stems and the leaves of
252 tetraploid plants were thicker and larger than in diploid ones [23] (1999). Also, BA treatment at 10 ppm increased growth
253 characters i.e., plant height, total root length fresh and dry weights of shoots and roots of maize plants [24]. Foliar spray of
254 soybean plants with benzyl adenine at 75 ppm significantly increased plant height, leaves number and branches per plant
255 and dry matter of plant [25]. The effect beneficial of foliar application of soybean plants with benzyl adenine at 50 ppm
256 significantly increased stem length, diameter, leaf area surface, branches number, leaves number per plant and fresh and
257 dry weights of plant [26]. Similarity, the foliar application of pelargonium (Geranium) plants with BA at 20 and 40 mg/L
258 significantly increased plant height and number of branches/plant finding by [27]. Egyptian lupine plants exposed to salt
259 stress, observed that foliar application of benzyl adenine (BA) (1 & 100 ppm) has stimulating effect on all growth
260 characters, i.e., plant height and number of branches/plant grown under normal and saline conditions [28]. **In Nigella**
261 *sativa* plants which benzyl adenine (5 & 25 ppm) treatments as seed soaking increased root length and diameter, plant
262 height stem diameter, number of leaves, total leaf area/plant and net assimilation rate [29]. Foliar spray of snap bean
263 plants with benzyl adenine (BA) at 20 & 40 ppm and putrescine (Put) at 200 ppm significantly increased plant height,
264 leaves number/plant and branches and fresh and dry weights of shoots [30]. The increased values of vegetative
265 parameters due to the lower dose of colchicine might be due to enhance the action of auxin (indole-3-acetic acid) and the
266 cells divided more actively in *Helianthus tuberosus* [31]. Higher doses of colchicine led to increased leaf size and number
267 of leaves per plant in colchicine-treated plants over control in *Gossypium arboreum* L [32]. EMS-treated plants were also
268 reported in papaya increased cell division, as well as activation of growth hormones such as auxin [33]. The effect of
269 colchicine-treated seeds of *Phlox drummondii* has been found to increase the seed germination and morphological
270 characteristics at low concentrations [34]. The effect of EMS-treatments on induced micro mutations and obtained on
271 dwarf plant types. The minimum plant height in dwarf mutant was below 90 cm. The maximum frequency of dwarf mutants
272 was observed in 30kr + 0.1% EMS followed by 40kr + 0.25% EMS treatment. The tallest mutant (155cm) was observed in
273 0.25% EMS treatment followed by a mutant with 131 cm in 30kr+0.25% EMS while the parent of rice *Akshaya* cv. possess
274 100-110cm height [35].

279 Table (2): Impact of papaya seed soaking in different BA, colchicine and EMS solutions on some growth measurements
 280 during 2015 & 2016 experimental seasons.

Parameters Treatments	No. leaves /seedling		Seedling height (cm)		Seedling diameter (cm)		Root length (cm)	
	First season	Second season	First season	Second season	First season	Second season	First season	Second season
1. control	9.33 f	7.67 f	52.33e	58.67f	2.53e	2.45de	14.73d	14.85d
2. BA at 1 %.	14.00 c	11.67 cd	99.00a	101.00cd	2.77d	2.83c	18.53c	18.63c
3. BA at 2 %.	15.33 b	13.00 b	97.00ab	103.00bc	3.13b	3.20b	21.38b	21.40b
4. BA at 3 %.	17.67 a	16.33 a	96.83ab	100.00d	3.37a	3.40a	23.80a	23.87a
5. colchi at 1 %.	10.33 e	13.00 b	75.00c	101.33cd	2.93c	3.13b	13.63ef	13.50f
6. colchi at 2 %.	10.67 e	8.00 ef	95.07b	105.00b	2.65de	2.62d	13.32fg	13.30f
7. colchi at 3 %.	7.67 g	8.67 e	97.00ab	113.33a	2.37f	2.27e	13.02g	13.07f
8. EMS at 10 ppm	12.67 d	11.00 d	70.00d	78.67e	2.50ef	2.45de	13.45fg	13.50f
9. EMS at 20 ppm	12.67 d	12.00 c	76.33c	80.00e	2.65de	2.57d	13.93e	13.92e
10. EMS at 30 ppm	12.67 d	13.33 b	69.00d	80.67e	2.97c	2.87c	14.10e	14.23e

281 Means followed by the same letters within each column during every season are not significantly at 5 % level.

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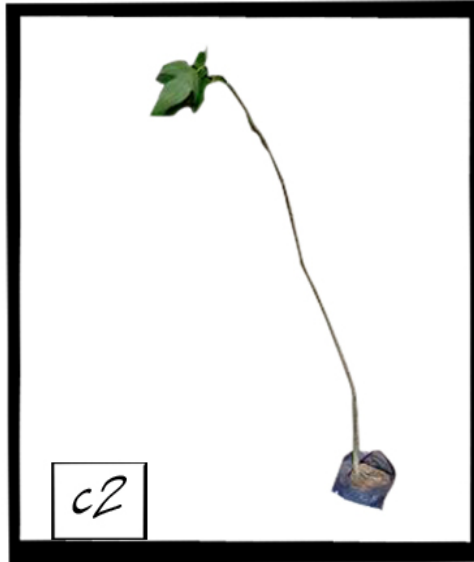
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Photo (1) Impact of papaya seed soaking in different BA, solutions on vegetative growth during 2015 & 2016 experimental seasons.

Con= control, BA1= BA solution at 1% , BA2=BA solution at 2% and BA3=BA solution at 3%.



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Photo (2) Impact of papaya seed soaking in different colchicine solutions on vegetative growth during 2015 & 2016 experimental seasons.

C1= control , C2= Colchicine solution at 1% , C3= Colchicine solution at 2% and C4= Colchicine solution at 3%.

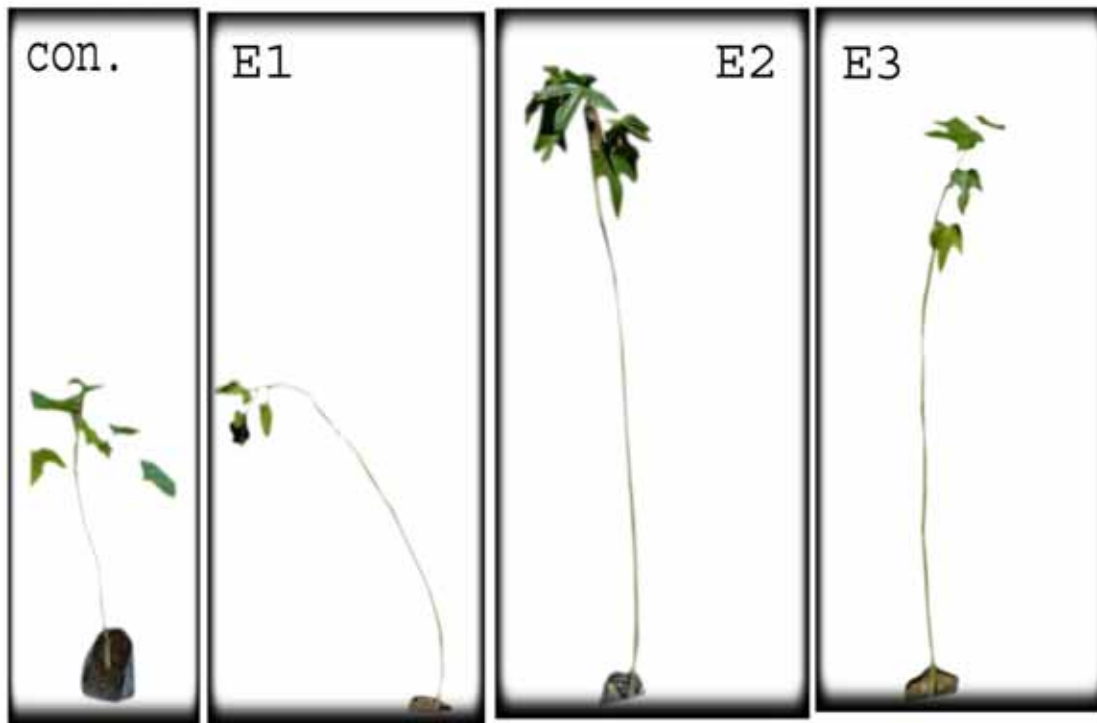


Photo (3) Impact of papaya seed soaking in different EMS solutions on vegetative growth during 2015 & 2016 experime seasons.

Con= control, E1= EMS solution at 10ppm, E2= EMS solution at 20ppm and E3= EMS solution at 30ppm.

Mitotic Index:

Means of mitotic index (MI %) resulted by BA, EMS and Colchicine are shown in Table 3. The means of mitotic index at three levels of Colchicine were close to each other and the same trend was also obtained by EMS. These results appeared that the differences between different levels of each agent were insignificant.

The means of dividing cells treated with Colchicine were significantly higher than of BA and EMS. This indicated that Colchicine did not interfere with mitosis and did not prevent cell division if compared with of BA and EMS which decreased the mitotic index and interfered with mitosis to greater extent.

Therefore, it can be concluded that EMS was more inhibitor of cell division followed by BA than Colchicine. This may be due to more damage resulted by BA and EMS affected on DNA replication during mitosis.

The figure shows the different chromosomal aberration as follows:

Sticky chromosomes at metaphase, but also laggards and lagging chromosomes, as well as polyploidy, were the main chromosomal aberrations or abnormalities during the cell division of papaya after treatment with the three mutagens with different ratio and different appearance.

Colchicine and EMS showed disrupted type of chromosomal aberrations which appeared during metaphase stage. It appeared that disrupted metaphase varied from Colchicine to EMS. In addition, EMS caused disrupted chromosomes in metaphase followed by anaphase which did not occur with Benzyl adenine.

Both Colchicine and EMS caused abnormal mitosis which appeared as sticky chromosomes. Colchicine caused sticky chromosomes in during metaphase and telophase. Similarly, EMS showed sticky with polyploidy chromosomes during metaphase, anaphase and telophase. These results indicated that colchicine had strongest effect on chromosomal

323 behavior during mitosis and exerted more chromosomal damage. Indeed, sticky chromosomes would cause the death of
324 those cells. Similar results were obtained by authors among them.

325 A chromatid bridge would occur as a result of the weakness of the spindle fiber. Bridge structure as an aberration
326 occurs due to treatment by both EMS and Colchicine.

327 During abnormal chromosomal behavior of mitosis, spindle fiber can not to attract one chromosome, this
328 chromosome remains near the middle of the cells. This phenomenon called lagging chromosome and resulted genome
329 aneuploidy $2n-1$. This kind of aberration did not occur among the chromosomal aberrations caused by Colchicine or EMS.
330 Formation of chromosomes from "star type" was observed. Both Colchicine and EMS caused this aberration type.

331 In conclusion, the treatments by colchicine and EMS caused different types of chromosomal aberrations with
332 variable percentages than the normal cells in control experiment the same time there were differences of the percentage
333 ratio of each. This indicated that both chemical agents are dangerous. EMS was more dangerous than Colchicine
334 because of cytotoxicity delaying mitosis and inducing mass chromosomal aberrations.

335 Sex determination in papaya (*C. papaya* L.) is due to a single gene with three allelic forms: *m*, *M1* and *M2*. The
336 *mm*, *M1m*, and *M2m* genotypes represent gynocious, androecious and hermaphrodite individuals, respectively. The
337 *M1M1*, *M2M2* and *M1M2* genotypes are not found due to the zygotic lethality. The *m* homologous region is normal and
338 the viable genotypes are *M1m* (male plant), *M2m* (hermaphrodite plant) and *mm* (female plant). A large concentration of
339 genes for femaleness is in the sex chromosomes but genes for maleness are in the autosomes. Therefore, the *mm*
340 genotype is distilled and its homozygote condition confers phenotypic stability [36] and [37]. Small doses of colchicine
341 enhanced the action of auxin (indole-3-acetic acid) because the cells divided more actively; instead, at higher doses,
342 colchicine led to C-mitoses and inhibited cell multiplication in *Helianthus tuberosus* [38].

344 The karyotype of *Carica papaya* L. consisted of eight medians (metacentric) four submedian, four sub terminal
345 and two terminal-centromeric chromosomes, formed that the arm ratio value of eight median centromeric chromosomes
346 range from 1.0 to 1.3 while the arm ratio value of four submedian centromeric chromosomes were very close to 3.1 the
347 lowest extreme of the arm ratio range of the sub terminal centromeric chromosome [15]. The cells with a larger
348 complement of chromosomes grow larger to maintain a constant ratio of cytoplasmic to nuclear volume, and express
349 more proteins with the presence of more genes. This increase in size may translate to an increase in the plant and its
350 organs [32]. Also, using several BAC clones that were explaining mapped to the papaya X/Y chromosomes, found that the
351 presumed sex chromosomes of *J. spinosa* are homomorphic and pair completely. In other species, chromosomes had
352 been counted with traditional means, and all were reported to have a diploid number of $2n = 18$. The remaining three
353 genera have never been studied, yet are disproportionally important because, respectively, they represent the deepest
354 divergence in the Caricaceae (*Cylicomorpha*) and the sister clade to *Carica* [39]. Gamma radiation, EMS, and their
355 combinations are potent mutagens, well known for their action causing point mutations, enzyme inhibitions and
356 chromosomal aberrations [40]. Sister to all New World Caricaceae is an African genus (*Cylicomorpha*) with two species. A
357 draft of the papaya genome became available in 2008, and since then, considerable effort has gone into understanding
358 the sex chromosomes of *C. papaya* [41]. All Caricaceae species are classified as diploids ($2n=2x=18$ chromosomes) and
359 dioecious, except for *C. papaya*, *V. monoica* e *V. cundinarmacensis*. The plant sexual determination in papaya is due to
360 one gene with three alleles. It was not observed sexual chromosome in their study. Thus, if there are sexual
361 chromosomes in *C. papaya*, they are probably homomorphic [42].

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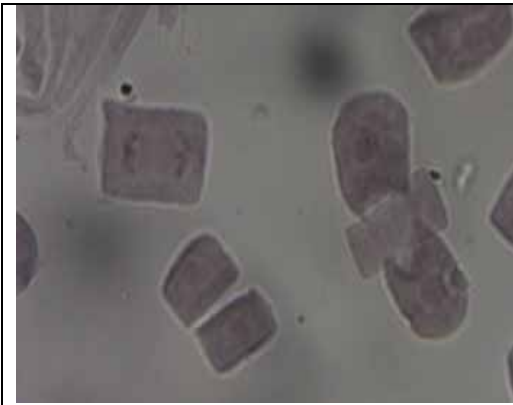


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photo (4): Normal metaphase without any treatment in the mitotic cell of papaya.

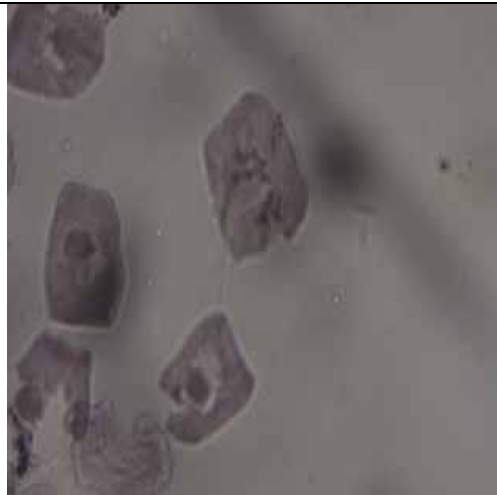
367



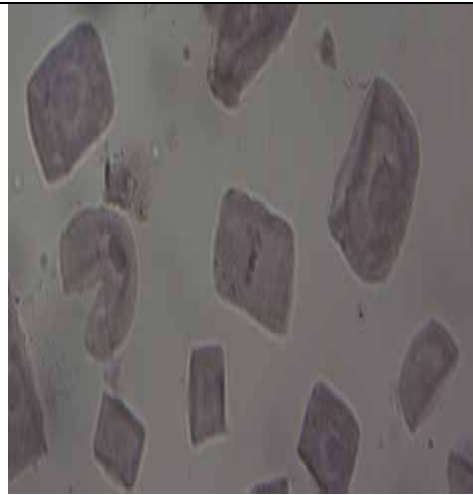
A



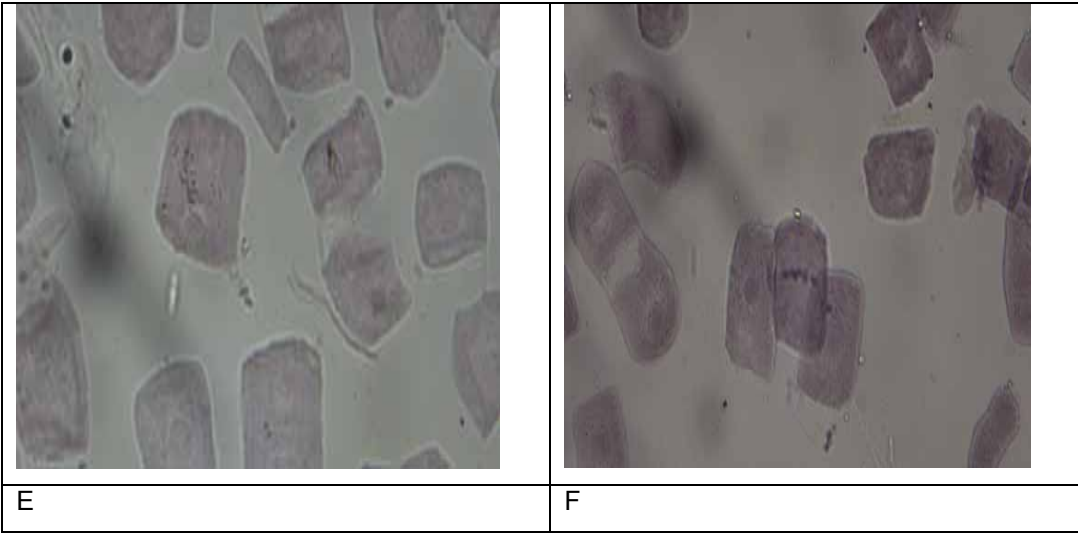
B



C



D



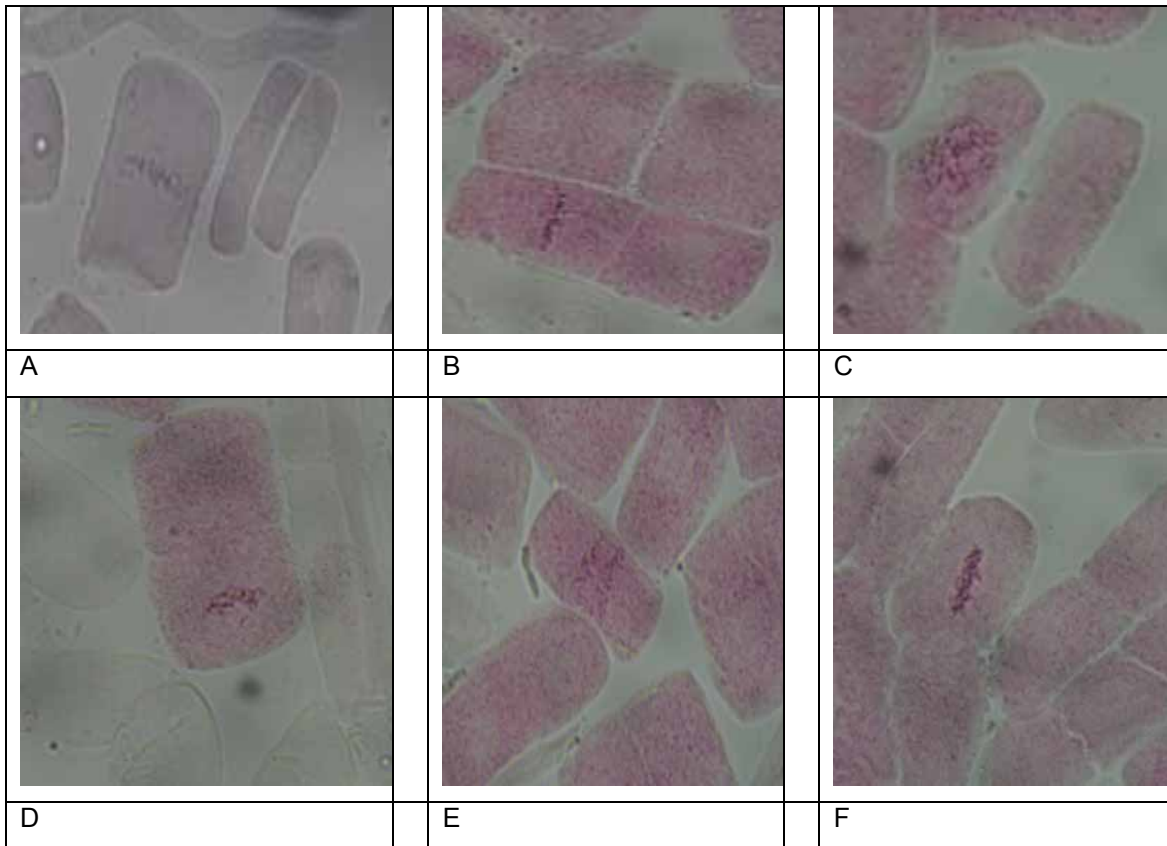
368 **photo (5): The effect of Benzyl adenine with three different concentrations on the mitotic cells of papaya.**

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370 photo (5): The effect of Benzyl adenine with three different concentrations on the mitotic cells of papaya. photo 5-
 371 A and B anaphase with irregular distribution of chromosomes between the two poles. photo 5-C three star groups of
 372 scattering of chromosomes in a dividing cell of a root tip at the beginning of telophase. photo 5-D one fragment at the
 373 equator of the metaphase. photo 5-E irregular distribution of chromosomes at the metaphase. photo 5-F Two laggards at
 374 metaphase.

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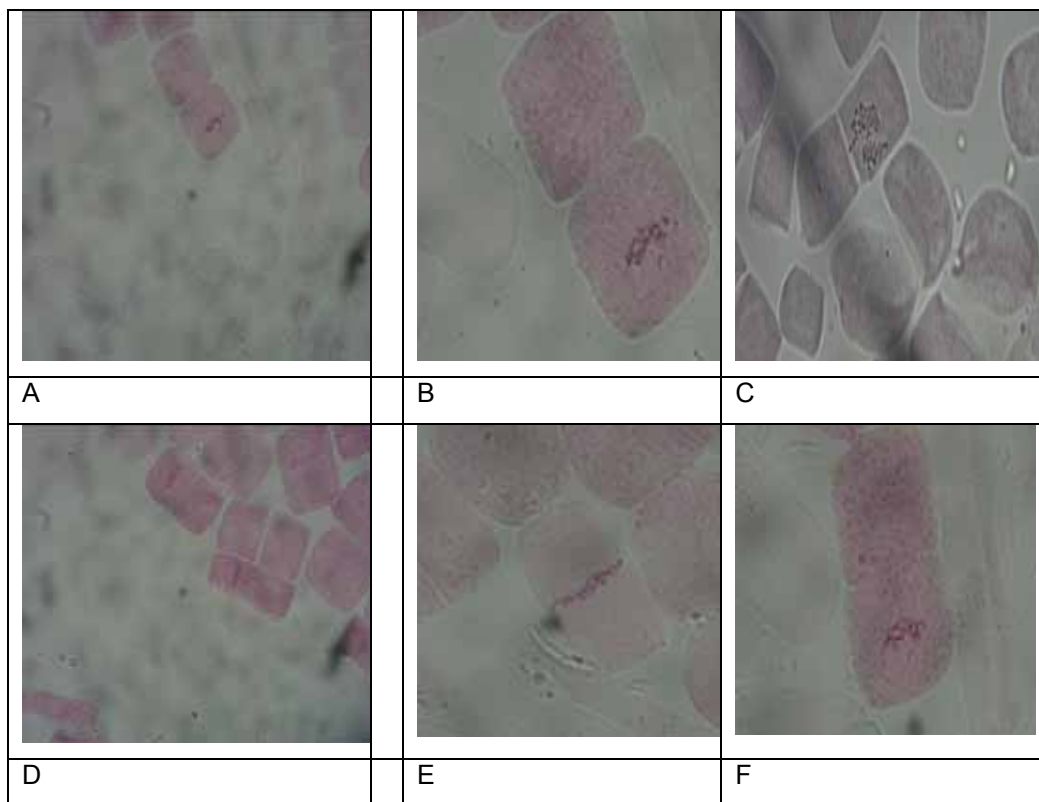
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377 **photo (6): The effect of EMS with three different concentrations on the mitotic cells of papaya.**

378 photo (6): The effect of EMS with three different concentrations on the mitotic cells of papaya. photo 6-A and B
 379 metaphase with one lagging chromosome. photo 6-C Scattering of chromosomes in a dividing cell of a root tip at
 380 metaphase. photo 6-D one lagging chromosome at metaphase. photo 6-E irregular distribution of chromosomes at the
 381 beginning of anaphase. photo 6-F clear polyploidy in metaphase with tetraploid number of chromosomes and C-
 382 metaphase.

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387 **photo (7): The effect of Colchicine at three different concentrations on the mitotic cells of papaya.**

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389 photo (7): The effect of Colchicine with three different concentrations on the mitotic cells of papaya. photo 7-A
 390 metaphase with one lagging chromosome. photo 7-B metaphase with two lagging chromosomes. photo 7-C Unequal
 391 distribution of chromosomes in anaphase with polyploidy. photo 7-D sticky chromosomes at metaphase. photo 7-E
 392 metaphase with tetraploid number of chromosomes. photo 7-F scattering of chromosomes in a dividing cell of a root-tip
 393 exposed to 3% colchicine.

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Table (3): Type and percentage of mitotic abnormalities in the root tips of papaya exposed to the Benzyl adenine, Ethylmethanesulphonate and colchicine with three different concentrations.

Conc. Ppm of mutagen	Total cells scores	No.of Divid . cells	MI %	Number of cells in the different phases of the cell cycle				
				Interpha se	proph ase	Metapha se	Anaph ase	Telopha se.
Control	500	92	18.4%	15.9%	2.20%	0.12	0.5	0.13
BA 1%	500	47	9.40%	8.02%	1.09%	0,10	0.04	0.15
BA 2%	500	32	6.40%	5.10%	0.98%	0.09	0.11	0.12
BA 3%	500	18	3.60%	2.11%	1.20%	0.07	0.02	0.20
Control	500	87	17.4%	14.8%	2.00%	0.22	0.08	0.0.8
EMS 1%	500	40	8.00%	6.01%	1.35%	0.16	0.04	0.28
EMS 2%	500	24	4.80%	3.00%	0.80%	0.25	0.49	0.26
EMS 3%	500	20	4.00%	2.90%	1.10%	0.00	0.00	0.00
Control	500	97	19.4%	17.95%	1.06%	0.20	0.09	0.11
Colchicine 1%	500	81	16.2%	15.05%	0.85%	0.40	0.05	0.30
Colchicine 2%	500	69	13.8%	12.00%	1.12%	0.23	0.30	0.15
Colchicine 3%	500	53	10.6%	9.00%	0.95%	0.16	0.30	0.19

4. CONCLUSION

According the results of the current study, both BA at 2% and 3% increased significantly germination %, germination rate and growth measurements. Moreover, in the presence of BA EMS inhibited the process of cell division, unlike addition of Colchicine. This effect might be explained with affected DNA-replication during mitosis by more damages, caused by BA and EMS.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- [1] Halmann, M. (1990). Synthetic plant growth regulators. *Advances in Agronomy* 43:47-105.
- [2] Khryanin, V.N. 2002. Role of phytohormones in sex differentiation in plants. *Russian Journal of Plant Physiology* 49 (4):545-551.
- [3] Davies, P.J. (2004a). The plant hormones: Their nature, occurrence and function. In *Plant hormones biosynthesis, signal transduction, action!*, ed. P. J. Davies, 750. Dordrecht; Norwell, MA: Kluwer Academic Publishers.
- [4] Nishijima, T., M. Hideari, K. Sasaki, and T. Okazawa. 2006. Cultivar and anatomical analysis of corolla enlargement of petunia (*Petunia hybrida* Vilm.) by cytokinin application. *Scientia Horticulturae* 111:49-55.

- 429 [5] Menendez, V., M.A. Revilla, and H. Fernandez. (2006). Growth and gender in the gametophyte of *Blechnum spicant* L.
430 Plant Cell Tissue and Organ Culture 86 (1):47-53.
- 431 [6] Kulkarni GB. (2011). Effect of mutagen on pollen fertility and other parameters in horsegram (*Macrotyloma uniflorum*
432 (Lam.) Verdc). Bio. Sci. discovery. 2 (1): 146-150.
- 433 [7] Murali K.M., Jeevanandam V., Shuye J. and Srinivasan R. (2013). Impact of colchicine treatment on *Sorghum bicolor*
434 BT× 623, Mol. Plant Breed. 4(15) 128–135.
- 435 [8] Wattoo J.I., Aslam K., Shah S.M., Shabir G., Sabar M., Naveed S.A., Waheed R., Samiullah, Muqaddasi Q.H.,
436 [9] Mohamed Z., Ho W.S., Pang, S.L., Ahmad F.B. (2014). EMS-induced mutagenesis and DNA polymorphism
437 assessment through ISSR markers in *Neolamarckia cadamba* (kelampayan) and *Leucaena leucocephala* (petai belalang),
438 Eur. J. Exp. Biol. 4(4):156–163.
- 439 [10] Chacko, E.K. and R.N. Singh. (1966). The effect of gibberellic acid on the germination of papaya seeds and
440 subsequent seedling growth. Trop. Agr. 43:341–346.
- 441 [11] Snedecor, G.W. and Cochran, W.G. (1989). Statistical methods, pp 177-195. 8th edition. Iowa state university press.
- 442 [12]Duncan, D.B. (1955): Multiple range and multiple F tests. Biometrics, II: 1-42.
- 443 [13] Jawaharlal , M.; Sambandamoorthy, S. and Irulappan, L. (1991). Effect of gamma ray and EMS on seed germination
444 and seedling growth in acid lime (*Citrus aurantifoliaswingle*). South Indian Horticulture .39 (6). 332 – 336.
- 445 [14] Nusrat, S. and Mirza, B. (2002). Ethyl methane sulfonate induced genetic variability in *Lycopersiconesculentum*.
446 International J. of Agric. and biology. 4:1.89-92.
- 447 [15] Bakry, KH. A. and Ismaeil, F. H. (2002): Pre-sowing treatments of papaya seeds as influenced by some chemicals
448 and irradiation on germination, growth, flowering, sex expression and fruit quality. 2ndIntre. Conf. Hort. Sci., 10- 12 sept.,
449 Kafr El-sheikh, Tanta Univ. Egypt.
- 450 [16] Padma, K. and Chauhan, P. (2005). Effect of EMS on germination, plant height and fruiting of
451 *Lycopersiconesculentum*. Flora and fauna (Jhansi), 11. 39-41.
- 452 [17] Sayed, S. Sawsan; Yousef, Hanan, M. A. and Yousef, E.M.A. (2007). Influence of colchicines and sodium azide
453 treatments on micropropagability and biochemical constituents of *Solidago altissima* Gray var "Tara" explants in vitro. J.
454 Biol. Chem. Environ. Sci. Vol. 2 (2): 257-276.
- 455 [18] Jaskani, M.J.; Kwon, S. W.; Kim, E. and Bokrae, K. (2004): Polypoidy affects fruit characteristics, seed morphology
456 and germination in watermelon (*Citrulluslanatus*). J. of the Korean society for Horti. Science. 45 (5) 233 – 237.
- 457 [19] Pranath, B., Rekha, A. and Pandey, A.K. (2015). Effect of pre-sowing treatments with chemical mutagens on seed
458 germination and growth performance of jamun (*Syzygium cumini* L. Skeels) under different potting substrates. Fruits. vol.
459 70(4): 239-248.
- 460 [20] Chakraborty N. R. and Kole P.C. 2009.Gamma ray induced morphological mutations in non-basmati aromatic rice.
461 *Oryza* 46 (3): 181-187.
- 462 [21]El-Kobisy, O. S. A. (1988). Ethylmethane sulphonate morphological .Ms.D. Thesis of Agric.cairo Univ .
- 463 [22] Arigita, L.; Fernandez, B.; Gonzalez, A. and Sanchez Tames, R. (2005): Effect of the application of benzyladenine
464 pulse on organogenesis, acclimatisation and endogenous phytohormone content in kiwi explants cultured under
465 autotrophic conditions. Plant Physiology and Biochemistry, 43:161-167.
- 466 [23]Wang-Honghe; X.U. Gexin; Q. (1999). In vitro induction of applied plants in colchicine-treated *Sinningia speciosa*. J. of
467 Tropical and Subtropical Botany, 7: 237-242.
- 468 [24] Shadi, A. I.; Sarwat, M. I.; El-Din, M. A. T. and Abou Deif, M, H. (2001): Effect of benzyladenine treatment on
469 chemical composition and salt tolerance of some maize in breeds under salt a stress. J. of Agric., Sci., 9(1):95-108.
- 470 [25] atil, R. R.; Deotale, R. D.; Hatmode, C. N. and Band, P. E.; Basole, V. D. and Khobragad, T. R. (2002 b).Effect of 6-
471 benzyladenine on biochemical and yield contributing parameters and yield of soybean. India. J. of Soils and Crops, 12(2):
472 270-273.
- 473 [26] Gad, M. S. H. (2005): Physiological studies on the effect of some growth regulators on soybean plant. M.Sc. Thesis,
474 Fac. of Agric., of Moshtohor; Zagazig Univ.
- 475 [27] Youssef, A. A. (2004). Influence of foliar spray with brassinosteroid and benzyladenine on the growth, yield and
476 chemical composition of {*Pelargonium graueolens* L) plants. Annals Agric. Sci, Ain Shams Univ Cairo.49(1)313-326.
- 477 [28]Medani, R. A. (2006): Effect of salinity, benzyladenine and their interaction on botanical characters and chemical
478 constituents of Egyptian lupine plant (*Lupinus termis*, L.) Annals of Agric. Sci., Moshtohor, 44(4): 1609-1628.
- 479 [29] Abd El-Gawad, H. A. (2006): Growth performance of black cumun (*Nigella sativa* L.) plants using certain growth
480 conditions. Ph.D. Thesis, Fac. of Agric., Moshtohor Benha Univ.
- 481 [30] Abo El-Saoud, M. S. (2005): Physiological studies on the role of some bioregulators in growth, flowering and yield of
482 snap bean. Ph.D. Thesis, Fac. of Agric., Moshtohor, Benha Univ.
- 483 [31] Bennici A., Silvia S. and Bruno M. (2006). Morphogenic effect of colchicine in *Cichorium intybus* L. Root explants
484 cultured in vitro, Caryologia 59(3) 284–290.
- 485 [32]Raufe S., Khan I.A. and Khan F.A., (2006). Colchicine-induced tetraploidy and changes in allele frequencies in
486 colchicine-treated populations of diploids assessed with RAPD markers in *Gossypium arboreum* L, Turk. J. Biol.(30) 93–
487 100.
- 488 [33] Singh S.V., Singh D.B., Yadav M., Roshan R.K. and Pebam N.(2010). Effect of EMS on germination, growth and
489 sensitivity of papaya (*Carica papaya* L.) cv. Farm Selection-1, Acta Hort. 851 113–116.

- 490 [34]Tiwari A.K.and Mishra S.K. (2012). Effect of colchicine on mitotic polyploidization and morphological characteristics
491 of *Phlox drummondii*, Afr. J. Biotechnol. 11(39) (2012) 9336–9342.
- 492 [35] BOLBHAT SADASHIV N. BHOGE VIKRAM D. AND DHUMAL KONDDIRAM N. (2012). Effect of mutagens on seed
493 germination, plant survival and quantitative characters of horsegram (*Macrotyloma uniflorum* (Lam.) Verdc). Research
494 Article. Vol 2(4).Oct-Dec. 129-136.
- 495 [36] Hofmeyr J. D. J. (1938). Genetical studies of *Carica papaya* L. the inheritance and relation of sex and certain plant
496 characteristics. South African Department of Agri. and Science Bulletin, n. 187:123-155.
- 497 [37] Storey W. B., (1953).Genetics of the papaya. Journal of Heredity, 44 (2): 70-78.
- 498 [38] Martin G., (1945). Action de la colchicine sur les tissus de topinambour cultivé in vitro. Rev. Cytol. Cytophysiol. Veg.
499 8. 1–34. Mooney, P.A. and Van Staden, J. 1986. J. Plant physiology. 123, 1-21.
- 500 [39] Alexander Rockinger 2 , Aretuza Sousa , Fernanda A. Carvalho , and Susanne S. Renner 2016. Chromosome
501 number reduction in the sister clade of *Carica papaya* with concomitant genome size doubling. AMERICAN J. OF
502 BOTANY. 103 (6):1082 – 1088.
- 503 [40] Auti SG. 2005. Mutational Studies in mung (*Vigna radiata* (L.) Wilczek). Ph.D. Thesis.University of Pune, Pune (MS),
504 India.
- 505 [41] Ming , R. , S. Hou , Y. Feng , Q. Yu , A. Dionne-Laporte , et al. 2008 . The draft genome of the transgenic tropical fruit
506 tree papaya (*Carica papaya* Linnaeus). Nature 452 : 991 – 996 .
- 507 [42] Corrêa, D.J.P., Fabiane, R.C., Telma, N.S., Pereira, M.F.N. and Messias, G.P.(2009). Karyotype determination in
508 three caricaceae species emphasizing the cultivated form (*C. papaya* L.). CARYOLOGIA Vol. 62, no. 1: 10-15.

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