SYNTHESIS, ANTIBACTERIAL AND ANTIOXIDANT ACTIVITIES OF SOME ONO SUBSTITUTED SALICYLALDIMINES

ABSTRACT

Five substituted ONO salicylaldimines, (E)-2-((2-hydroxybenzylidene)amino)phenol, (E)-2-(((2-hydroxyphenyl)imino)methyl)-4-nitrophenol, (E)-4-chloro-2-(((2-hydroxyphenyl)imino)methyl)phenol, (E)-2-(((2-hydroxyphenyl)imino)methyl)-4-methoxyphenol, (E)-4-bromo-2-(((2-hydroxyphenyl)imino)methyl)-6-methoxyphenol were synthesized and characterized by elemental analysis, IR, UV and NMR (^{1}H and ^{13}C). They were screened against some multi-drug resistance Gram-positive (Streptococcus agalactiae and Staphylococcus aureus), and Gram-negative (Escherichia coli, Klebsiella pneumonia, Proteus mirabilis, Pseudomonas aeruginosa and Salmonella typhimurium) organisms by the agar-well diffusion method. The total antioxidant capacities of the salicylaldimines were determined by phosphomolybdenum assay. Their antibacterial and antioxidant activities were screened to understand the substituents effects.

The result showed that the methoxy-substituted compound exhibited the highest antibacterial and antioxidant activities while the nitro-substituted compound exhibited the least activities. This implies that the electron donating group, methoxy on the compound increases its antibacterial and antioxidant activities.

Keywords*: Schiff base, substituents, antioxidants, characterized, antibacterial.*

1 Introduction

Salicylaldimines are 2-hydroxyl Schiff bases formed from the reaction between salicylaldehyde and a primary amine [1-3] while Schiff bases are condensation products of carbonyl compounds and primary amines. They are aldehyde or ketone like compounds in which the carbonyl (C=O) group is replaced by an imine or azomethine (−HC=N−) group. Schiff bases having the bioactive sites of N, O and/or S atoms have attracted much attention and have a wide range of applications in medicinal, pharmaceutical, [4-8]. For instance, Schiff bases have been used as anti-inflammatory [9], analgesic [10], antimicrobial [1, 6, 11-17], anticonvulsant [18], antitubercular [19-22], anticancer [23-25], antioxidant [6, 15-17, 26], anthelmintic and antimalarial [27, 28] which make them gain importance in medicinal and pharmaceutical fields.

The steric and inductive effects introduced by substituents present on the aromatic portion of the Schiff base can influence its properties significantly [29]. As an additional contribution to understanding the substituent effects on the antibacterial and antioxidant activities of Schiff bases, we herein report the synthesis, spectral, antibacterial and antioxidant activities of some ONO salicylaldimines.

2 Materials and method

2.1 Reagents

Salicylaldehyde, 5-methoxysalicylaldehyde, 5-bromo-3-methoxysalicylaldehyde, 5-nitrosalicylaldehyde, 5-chlorosalicylaldehyde, and *o*-aminophenol were purchased from Merck (Germany) and used as supplied. The solvent DMSO (dimethyl sulfoxide) and

absolute ethanol were of analytical grade and were used without further purification. Elemental analysis was carried out on Finnigan Flash EA 1112 series. The electronic spectra were recorded on Shimadzu UV-2600 series (Japan), in DMSO. The infrared spectra were recorded on a Perkin-Elmer 400 FT-IR/FT-FIR while the NMR spectra were recorded on Bruker Avance III 600 in DMSO-d_6 solution with tetramethylsilane (TMS) as internal reference.

2.2 Synthesis of Schiff bases

A 0.015 mole of *o*-aminophenol in 15 ml of absolute ethanol was added to a stirring solution containing 0.015 mole of the appropriate salicylaldehyde in 10 ml absolute ethanol. The resulting mixture was stirred for 2 hrs. The precipitates were filtered and washed with cold ethanol, recrystallized from ethanol and dried in a desiccator over silica gel for two days.

2.3 Antibacterial Activity

The antibacterial potentials of the samples were evaluated by agar-well diffusion method as described by Ghosh, Mitra [17] against multi-drug resistance Gram-positive (*Streptococcus agalactiae and Staphylococcus aureus),* and Gram-negative *(Escherichia coli, Klebsiella pneumonia, Proteus mirabilis, Pseudomonas aeruginosa and Salmonella typhimurium*) organisms. The bacteria isolates were sub-cultured in Nutrient agar and incubated at 37 ^{o}C for 24 hours. All the bacteria cultures were adjusted to 0.5 McFarland standards, 20 ml of sterilized Nutrient agar medium was dispensed into each petri dish aseptically and allowed to gel. The plates were swabbed with inocula of the test organisms and kept for 15 minutes for adsorption unto the gel. Using sterile cork borer of 6 mm diameter, wells were bored into the seeded agar plates, and these were loaded with different concentrations of the samples. The plates were allowed to stand in the refrigerator for 1 hour to allow proper diffusion of the sample into the medium and incubated at 37 ^{o}C for 24 hours before visual assessment of the inhibition zones. Antimicrobial activities were expressed as inhibition diameter zones in millimeter (mm). Gentamicin was used as a control.

2.4 Phosphomolybdate Total Antioxidant Capacity (PTAC) Assay

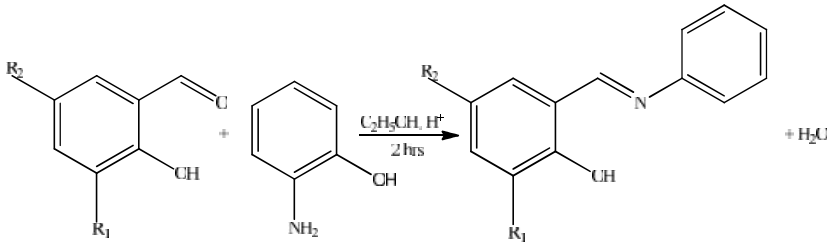
The total antioxidant capacities (TAC) of the samples were determined by phosphomolybdenum assay using ascorbic acid as the standard. An aliquot of 1.0 ml of extract (1000 µg) solution was combined with 1.0 ml of reagent (0.6 M sulphuric acid, 28 µM sodium phosphate and 4 µM ammonium molybdate). The tubes were capped and incubated in a boiling water bath at 95 ^{0}C for 90 min and cooled to room temperature. The absorbance of the aqueous solution of each mixture was measured at 695 nm in UV spectrophotometer. The blank solution having only reagent solutions was treated and analyzed in a similar manner as described above. The total antioxidant capacity was expressed as equivalents of ascorbic acid. ~~(AA).~~

3 Results and Discussion

3.1 Synthesis

The condensation (Scheme 1), of *o*-aminophenol and corresponding substituted salicylaldehyde gave the following Schiff bases: **I** (*E*)-2-((2-hydroxybenzylidene)amino)phenol. **II** (*E*)-2-(((2-hydroxyphenyl)imino)methyl)-4-nitrophenol. **III** (*E*)-4-chloro-2-(((2-hydroxyphenyl)imino)methyl)phenol. **IV** (*E*)-2-(((2-

hydroxyphenyl)imino)methyl)-4-methoxyphenol. **V** (*E*)-4-bromo-2-(((2-hydroxyphenyl)imino)methyl)-6-methoxyphenol.



(**I**) R_1= R_2= H

(**II**) R_1= H, R_2= OH

(**III**) R_1= H, R_2= Cl

(**IV**) R_1= H, R_2= OCH_3

(**V**) R_1= OCH_3, R_2= Br

Scheme 1: Synthesis of Schiff Bases (I-V).

3.2 Characterization of the Schiff Bases.

Table 1: Analytical Data of the Schiff Bases.

Compounds	Empirical formula	Molecular weight (g/mol)	Yield (%)	Elemental analysis % C	H	N
I	$C_{13}H_{11}NO_2$	213.23	86%	73.22 (73.3)	5.21 (5.77)	6.57 (6.16)
II	$C_{13}H_{10}N_2O_4$	258.23	80%	60.50 (60.47)	3.89 (3.90)	10.82 (10.85)
III	$C_{13}H_{10}ClNO_2$	247.68	82%	63.03 (63.04)	4.07 (4.07)	5. 68 (5.66)
IV	$C_{14}H_{13}NO_3$	243.26	82%	69.10 (69.12)	5.40 (5.39)	5.80 (5.76)
V	$C_{14}H_{13}NO_3$	322.15	84%	52.18 (52.20)	3.80 (3.75)	4.46 (4.35)

Key: Calculated values are in parenthesis

Table 2: Important IR, ^{1}H NMR and UV of the Schiff Bases.

Compounds	IR (cm^{-1})			NMR (^{1}H and ^{13}C)		UV-Vis (nm)	
	O−H	C=N	C−O	δ(ppm)	Assignments	n-π*	π-π*
I	3746	1627	1274	13.78 9.73 8.92 7.56-6.84 162.21	(s, 1H, −OH) (s, 1H, −OH) (s, 1H, −HC=N) (m, 8H, $CH_{Aromatic}$) (s, 1C, −CH=N)	297	353
II	3067	1614	1306	13.50 10.34 9.26 8.56-6.84 160.59	(s, 1H, −OH) (s, 1H, −OH) (s, 1H, −HC=N) (m, 7H, $CH_{Aromatic}$) (s, 1C, −CH=N)	297	353

III	3730	1615	1272	13.77 9.76 8.92 7.68-6.83 160.51	(s, 1H, −OH) (s, 1H, −OH) (s, 1H, −HC=N) (m, 7H, $CH_{Aromatic}$) (s, 1C, −CH=N)	282	360
IV	3747	1626	1247	13.07 9.68 8.89 7.29-6.82 3.71 161.52	(s, 1H, −OH) (s, 1H, −OH) (s, 1H, −HC=N) (m, 7H, $CH_{Aromatic}$) (s, 3H, $CH_{Methoxy}$) (s, 1C, −CH=N)	270	370
V	3740	1615	1253	13.37 9.86 8.91 7.36-6.83 3.79 160.12	(s, 1H, −OH) (s, 1H, −OH) (s, 1H, −HC=N) (m, 7H, $CH_{Aromatic}$) (s, 3H, $CH_{Methoxy}$) (s, 1C, −CH=N)	290	350

***Key**: s = singlet, m = multiplet.

The compounds were obtained as solids in good yields, their colors range from orange-wine-yellow. They are air stable. Their analytical data are summarized in Table 1.

The Important IR, ^{1}H NMR and UV of the Schiff Bases are presented in Table 2. The IR spectral data of each of the Schiff Bases confirms the formation of the azomethine bond ν(−HC=N). Their IR spectral data showed the azomethine ν(HC=N) bands in the range 1627-1614 cm^{-1}. All the compounds displayed a band at 1306-1247 cm^{-1} which was assigned to the phenolic stretching ν(C−O) vibration while the hydroxyl (O−H) band appeared in the range 3747-3067 cm^{-1} [13, 21, 30-35].

The ^{1}H NMR spectra of the Schiff bases (Fig. 1-5) showed two singlet signals at δ 13.78-13.07 ppm and δ 9.86-9.68 ppm which were assigned to two phenolic –OH protons [8, 13, 21, 34, 36]. All the Schiff base ligands showed a singlet signal at δ 9.26-8.91 ppm attributed to the azomethine (−HC=N) protons [8, 15, 20, 30]. The aromatic protons appeared as multiplets at δ 7.68-6.82 ppm [8, 33, 35, 37, 38]. One sharp singlet signal assigned to the protons of methoxy (–OCH_3) groups appeared at δ 3.71 and 3.80 ppm in the spectra of compounds **IV** and **V** respectively [8, 21]. The carbon-13 NMR spectra of the compounds showed singlet signals assigned to the azomethine carbon in the range 162.21-160.12 ppm. This further confirms the formation of the Schiff bases.

The electronic spectra of the Schiff bases showed two absorption peaks at 297-270 nm and 370-350 nm assigned to transitions of $n–\pi^*$ of the azomethine and $\pi–\pi^*$ of the aromatic ring in the Schiff bases respectively.

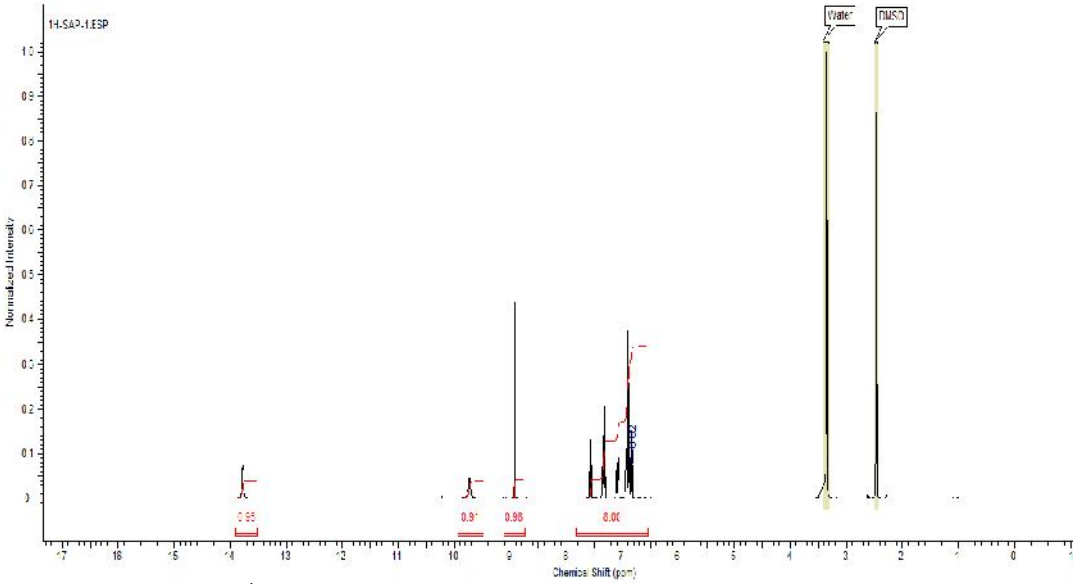


Figure 1 The proton (^{1}H), NMR spectrum of compound (I)

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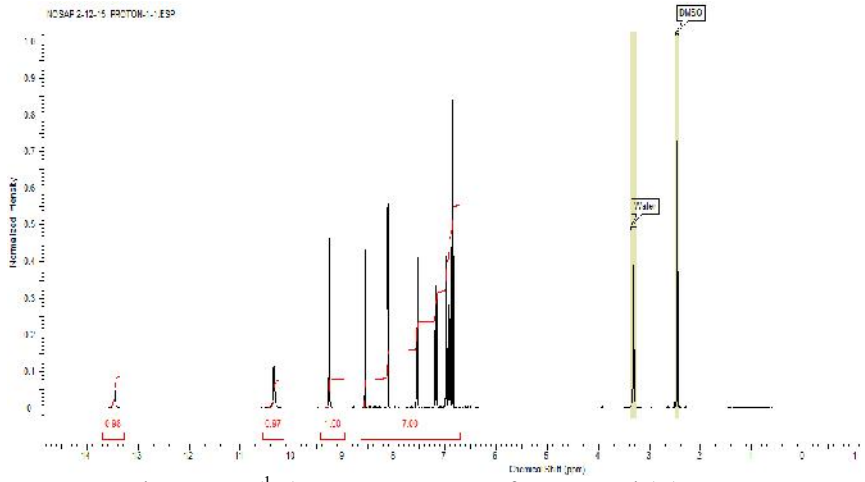

Figure 2: The proton (^{1}H), NMR spectrum of compound (II)

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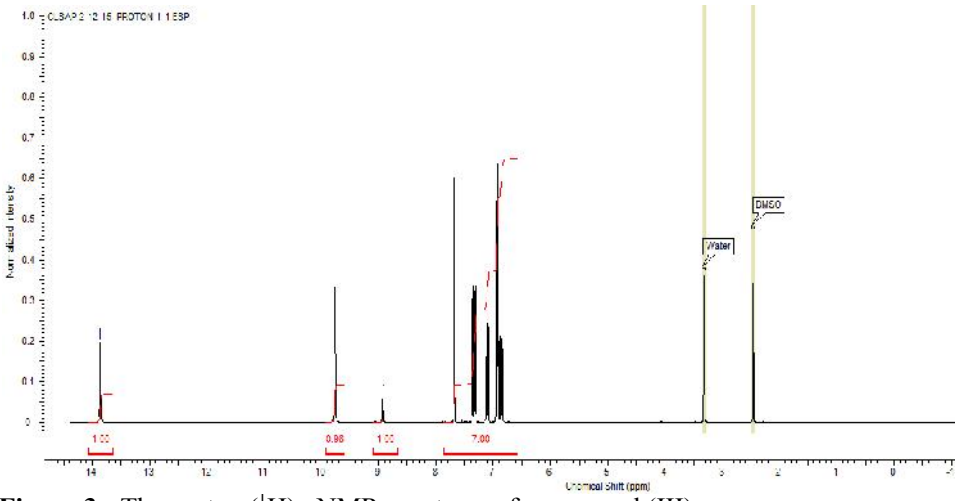

Figure 3: The proton (^{1}H), NMR spectrum of compound (III)

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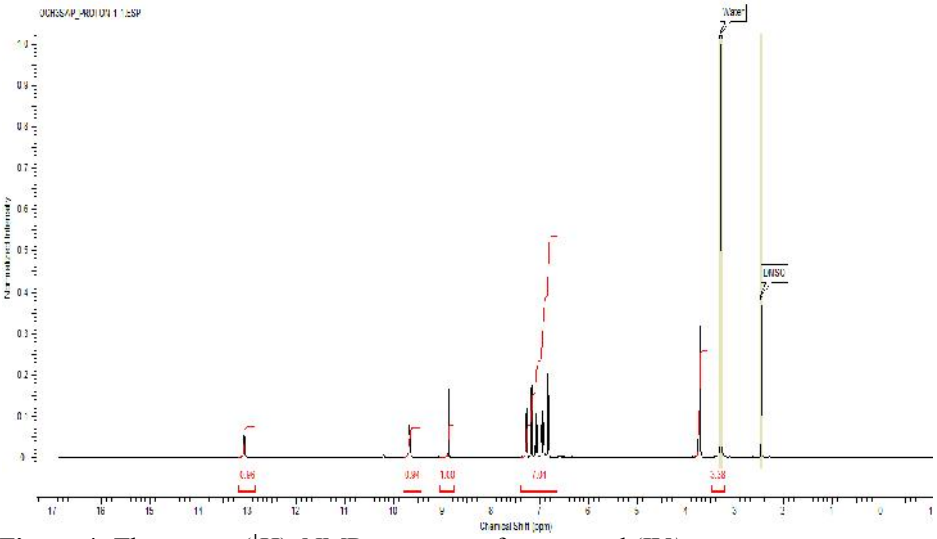

Figure 4: The proton (^{1}H), NMR spectrum of compound (IV*)*

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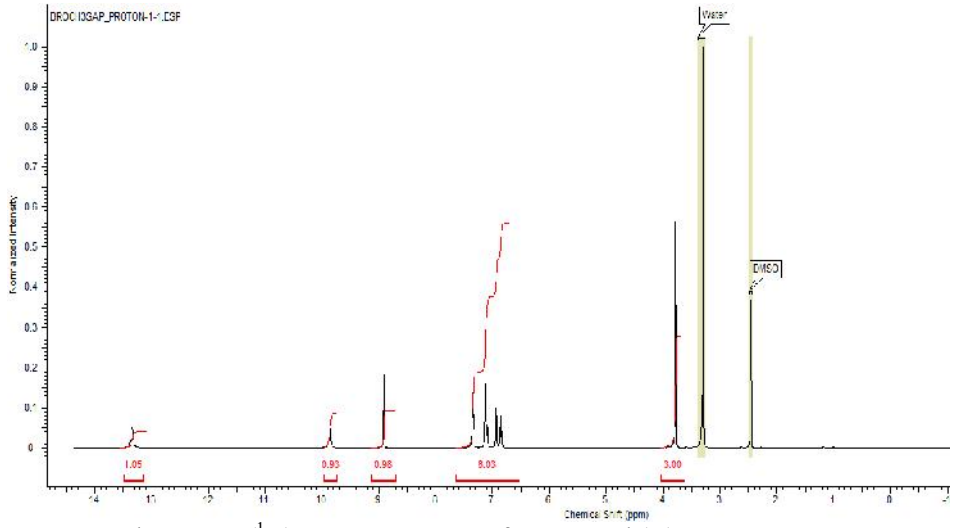

Figure 5: The proton (^{1}H), NMR spectrum of compound (V)

3.3 Antimicrobial Activity

Table 3: Zone of Inhibition Showing the Antimicrobial Potentials of Compounds (1-V).

Compounds	*E.coli*	*K. pneumonia*	*P. aeruginosa*	*S. agalactiae*	*S. aureus*	*S. typhimurium*	*P. mirabilis*
I	16.3	15.7	17.7	14	16.3	15.7	14
II	13.7	15	12	15.3	08	–	11
III	15.3	13.3	16	20	15.3	16	20
IV	22	20	30	22	30	18	25
V	11.3	13	13.7	13.3	15	11	12.3
DMSO	–	–	–	–	–	–	–
Gentamicin	20	18	20	–	20	11	20

Key: Resistant, = (-) ; not sensitive = (<8 mm), sensitive = (9 to14 mm), very sensitive = (15 to19 mm) and ultrasensitive = (>20 mm).

The result of the antimicrobial activity of the compounds (Table 3) revealed that all the synthesized compounds are sensitive to all the bacteria strains except compound **II** to which *S. typhimurium* is resistant. All the compounds are sensitive to *S. agalactiae* which is resistant to Gentamycin. Compound **IV** is ultrasensitive to all the bacteria strains but for *S. typhimurium* to which it is very sensitive. It showed the highest sensitivity to all the bacteria strains which could be accounted for by the methoxy-substituent ($–OCH_3$) on the compound. Despite the $–OCH_3$ substituent on compound **V**, it is not as sensitive to the bacteria strains as compound **IV** owing to the presence of bromo (–Br) substituent on the compound. This implies that the bromo substituent reduces the sensitivity of the compound. Compound **III** showed higher sensitivity to some bacteria strains particularly *S. typhimurium* and *S. agalactiae* than compound **I**, this could be accounted for by the presence of the chloro-substituent (–Cl) on the compound. The nitro-substituted ($−NO_2$) compound (**II**) showed the least sensitivity to the bacteria strains.

The resistance of some of the pathogens towards the tested compounds can be attributed to the existence of cell wall in the bacteria which reduces the permeability of the tested compounds while the activity against them can be attributed to the greater lipophilicity of the compounds.

Total Antioxidant Capacity

Table 4 Total Antioxidant Capacity (TAC)

Samples	I	II	III	IV	V
TAC µg per mg AA	0.68	0.52	0.73	0.78	0.56

The difference in the total antioxidant capacities of the Schiff bases presented in Table 4, could be explained by the presence of the different substituents on the compounds. The effect of the substituents on the total antioxidant capacities of the Schiff bases is same as their effect on the antimicrobial activities. Compound **III** showed the highest total antioxidant capacities while compound **II** showed the least capacities. Hence, it is a better free radicals scavenger.

4 Conclusion

The methoxy-substituted Schiff base exhibited the highest antibacterial and antioxidant activities compared to the nitro-substituted compounds. The antimicrobial and total antioxidant activities results revealed the order of activity of the compounds as **IV** > **III** > **I** > **V** > **II**. Thus, it can be assumed that the antibacterial and antioxidant activities of the Schiff bases under consideration in this study depend on the substituents on the salicylaldehyde.

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