

Original Research Article

Study of antioxidant, phytochemicals and heavy metals in crude leaf extract in different solvents of Castor (*Ricinuscommunis*)

Abstract:

The castor oil plant *Ricinuscommunis* is a member of the family Euphorbiaceae. This plant is a member of the genus *Ricinus* which is traditionally called as castor bean. Castor seed is the source of castor oil, which has a number of uses. The sample leaves of *Ricinuscommunis* (castor) obtained from Institute of Applied Science and Technology (IAST), University of Guyana, Turkeyen Campus, Georgetown, Guyana. Leaves were dried in oven at 50 – 60 °C for 72 h. The moisture content was calculated. The dried leaves were grounded and extracted in each chloroform, ethanol, ethyl acetate and methanol solvents. Each solvent extract collected separately and evaporated on rotavapour. The semi solid liquid extract was made to desired volume by the addition of respective solvent. Antioxidant, phytochemical screening of leaves extract for alkaloids, carbohydrates, saponins, proteins & amino acids, tannins, flavonoids, glycosides and terpenoids were done by standard test procedure reported in literature. Atomic absorption spectrophotometric study revealed the presence of the heavy metals Cu, Ca, Cr, Zn, Mn, Fe and Mg in the decrease order of their concentrations. Carbohydrate was tested positive in four solvent extracts. Alkaloid and saponins were tested negative in each four solvent extracts. All four leaves extract were found to have antioxidant potential.

Keywords: *Ricinuscommunis* (castor), antioxidant, phytochemicals, chloroform, ethanol, ethyl acetate, methanol, heavy metals.

INTRODUCTION:

Ricinuscommunis seed is the castor bean, which despites its name, is not a true bean. Castor is indigenous to the Southeastern Mediterranean Basin, Eastern Africa and India, but is widespread throughout tropical regions [1]. *R.communis* can vary greatly in its growth habit and appearance. It is a fast growing suckering perennial shrub that can reach the size of a small tree (around 12 meters or 39 feet). The glossy leaves are 15-45 cm long, long stalked, alternate and palmate with 5-12 deep lobes with coarsely toothed segments.

Three terpenoids and a tocopherol related compound have been found in the aerial parts of *R.communis*. Compounds named (3E, 7Z, 11E) – 19 hydroxycasba – 3, 7, 11 –trien-5-one, 6 α –hydroxyl – 10 β – methoxy - 7 α , 8 α -epoxy-5-oxocasbane-20, 10-olide, 15 α -hydroxylup-20(29)-en-3-one, and (2R, 4aR, 8aR – tetrahydro – 4a – hydroxyl-2, 6, 7 8a – tetramethyl-2 (4, 8, 12 – trimethyltridecyl)- 2H – chromene – 5, 8-dione were isolated from the methanol extracts of *R.communis* by chromatographic methods [2]. Partitioned n-hexane fraction of *R. communis*root method extract resulted in enrichment of two triterpenes lupeol and urs-6-ene-3, 16-dione (erandone). Crude methanolic extract, enriched in hexane fraction and is isolates at doses 100 mg/kg P.O. exhibited significant (P< 0.001) anti-inflammatory activity in carrageenan-induced hind paw edema model [3].

Ebers papyrus is an ancient Egyptian physician describes castor oil as a laxative [4]. Castor oil is well known as a source of ricinoleic acid a monounsaturated, 18-carbon fatty-acid. Among fatty acids, ricinoleic acid is unusual in that it has a hydroxyl functional group on the 12th carbon.

46 This functional group causes ricinoleic acid (and castor oil) to be more polar than most fats. The
47 chemical reactivity of the alcohol group also allows chemical derivatization that is not possible
48 with most other seed oils. Because of its ricinoleic acid content, castor oil is a valuable chemical
49 in feed stocks, commanding a higher price than other seed oils.

50
51 Singh and Geetanjali [5] have described pharmacological (e.g. anti-inflammatory, anti-diabetic,
52 anti-tumor, anti-asthmatic potential and other medicinal properties of extracts of different plant
53 parts of *R. communis*. They have also investigated the presence of important phytochemical
54 constituents such as flavonoids, glycosides, alkaloids, steroid, terpenoids, etc. and their possible
55 structure in the same extract. Anti-dandruff activity of *R. communis* L. methanol, aqueous,
56 chloroform and petroleum ether leaf extracts, against malasseziasspp, causative agent of dandruff
57 in people who have over active sebaceous gland was presented by Sibi et al. [6].

58
59 Gupta et al. [7] demonstrated strong antioxidant potential of the methanolic extract of *R.*
60 *communis* leaves. Antioxidant, anti-microbial and free radical scavenging potential of various
61 extracts of aerial parts of *R. communis* was examined by Iqbal et al. [8]. The present study
62 evidence that *R. communis* proved to be potent natural antioxidants that could replace synthetic
63 antioxidants. Vandita et al. [9] has investigated the effects of tannins, alkaloids, cardiac
64 glycosides, terpenoids, flavonoids and steroids of *R. communis* on antibacterial, fungal and
65 cytotoxic activities. The cytotoxic effect of selected plants were tested against HEK 293T
66 (Human embryonic kidney cell line) and C2C12 (Mouse, muscle cell line) by MTT assay.

67
68 Kadri et al. [10] has investigated in vitro antioxidant properties of essential oil of *R. communis*L.
69 The essential oil from the aerial parts of *R. communis*, was obtained by hydro-distillation and
70 analyzed by GM-MS. Antioxidant activity of the investigated essential oil was evaluated by
71 different test systems: 1, 1-diphenyl – 2 picrylhydrazyl (DPPH) assay, β -carotene bleaching test
72 and reducing power assay. The essential oil exhibited a potential antioxidant activity

73

74 **MATERIALS AND METHODS:**

75 **Collection of Plant materials**

76 The plant material, leaves of *R. communis* was collected from Institute of Applied Science and
77 Technology (IAST), University of Guyana, Turkeyen Campus, Georgetown, Guyana.

78 **Preparation of Plant Materials**

79 The collected leaves material of *R. communis* was weighted on Citizen CTG 3000E electronic
80 balance. The leaves were dried in oven (Gallenhamp Incubator Model IH – 150) at 50-60 °C.
81 The dried leaves were cooled at room temperature and weighted again on same citizen electronic
82 balance.

83 **Extraction and Preparation of Test Solutions**

84 The ground leaves of *R. communis* was extracted in each ethyl acetate, ethanol, ethanol and
85 chloroform solvents. Each time 20 grams of pulverized leaves were soaked with 200 ml of
86 solvent for 48 hours. The solvent is decanted each time and residue again soaked with same
87 solvent for 24 hours. The all extract is combined and filtered. The evaporation of solvent was
88 done on rotavapour (Buchi). The respective solvent was added to viscous semisolid liquid extract
89 to make up the derived volume of extract solution.

90 **Reducing antioxidant (Protective) power**

91 Potassium ferricyanide, trichloroacetic acid, butylated hydroxyl anisol, sodium
 92 dihydrophosphate, ferric chloride, ammonium thiocyanate, ferric chloride, linoleic acid (99.5 %),
 93 thiobarbituric acid, sodium monohydrophosphate, and potassium dihydrophosphate were
 94 obtained from Aldrich, USA. All chemicals were used without further purification. All aqueous
 95 solutions were prepared in double distilled water.

96 The reducing antioxidant or protective power of the plant ethanolic, ethyl acetate, chloroform
 97 and methanolic extract were determined by the method reported in literature [68, 69]. The
 98 different concentration of leaf extracts (100-1000 μL) in 1 m L of distilled water were mixed
 99 with phosphate buffer (2.5 m L, 0.2 pH 6.6) and potassium ferricyanide $\text{K}_3\text{Fe}(\text{CN})_6$ (2.5 m L 1
 100 %). The mixture was incubated at 50°C for 20 mins. Then 2.5 m L of trichloroacetic acid (10
 101 %) was added to mixture, which was then centrifuge for 10 mins at 3,000 rpm. The upper layer
 102 of solution (2.5 m L) was mixed with distilled water 92.5 m L and FeCl_3 0.5 m L 1 %. The
 103 absorbance was measured at 700 nm against a blank using UV-Vis spectrophotometer (Phillips
 104 X 500). Increased absorbance of the reaction mixture indicates increase in the reducing power.

105 **Phytochemical and heavy metals analysis of the plant extracts**

106 Phytochemical analysis of ethanolic, methanolic, ethyl acetate and chloroform leaves extract
 107 were carried out by suitable methodologies in search of active ingredients responsible for
 108 antimicrobial toxicity. The phytochemicals investigated were saponins, terpenoids, alkaloids,
 109 glycoside, carbohydrates, protein and amino acids, tannins and flavonoids. The phytochemical
 110 analysis was carried out according to the method reported in literature by Edeoga et al. [70].

111 The leaves (2g) was treated 10 10 cm^3 aqua regia (75 vol % hydrochloric acid and 25vol% nitric
 112 acid) and heated to dryness. Distilled water (20 cm^3) was added and the mixture stirred and
 113 filtered. The filtrate was subjected to analysis using Xplor AA – GOC Scientific Equipment
 114 Atomic Absorption Spectrophotometer.

115

116 **RESULTS AND DISCUSSION:**

117

118 **Antioxidant potential of *R. communis* leaves extract**

119 Antioxidant potential is the measure of reducing ability of the antioxidant. Antioxidant potential
 120 is evaluated by measuring the transformation of iron (III) to iron (II) in the presence of sample
 121 extract [11]. The ability to reduce iron (III) may be results from hydrogen donation from
 122 phenolic compounds [12], which is also related to the presence of some reducing agent [13]. In
 123 addition, the number and position of hydrogen group of phenolic compounds also affect their
 124 antioxidant potential [14]. The increase in concentrations of leaves extract may also cause
 125 deviation from increase in its reducing power which may be due to decrease in hydrogen donor
 126 ability of phenolic compounds. The reducing power of chloroform, ethanolic, ethyl acetate and
 127 methanolic *R. communis* leaf extracts are given in Tables 1, 2, 3 and 4, respectively.

128

129 **Table 1: Reducing antioxidant power of chloroform leaf extract of *R. communis* (Castor)**

S. No.	Volume of Leaf Extract (μL)	Absorbance of Leaf Extract (nm)
1	Chloroform (control)	0.000
2	1.00	0.001
3	2.00	0.004
4	3.00	0.004
5	4.00	0.005

6	5.00	0.006
7	6.00	0.006
8	7.00	0.007
9	8.00	0.006
10	9.00	0.005
11	10.00	0.009

130 **Table 2: Reducing antioxidant power of ethanol leaf extract of *R. communis***

S. No.	Volume of Leaf Extract (µL)	Absorbance of Leaf Extract (nm)
1	Ethanol (control)	0.000
2	1.00	0.001
3	2.00	0.003
4	3.00	0.004
5	4.00	0.002
6	5.00	0.007
7	6.00	0.007
8	7.00	0.008
9	8.00	0.009
10	9.00	0.006
11	10.00	0.007

131 **Table 3: Reducing antioxidant power of ethyl acetate leaf extract *R. communis*(Castor)**

S. No.	Volume of Leaf Extract (µL)	Absorbance of Leaf Extract (nm)
1	Ethyl acetate (control)	0.000
2	1.00	0.005
3	2.00	0.006
4	3.00	0.007
5	4.00	0.008
6	5.00	0.009
7	6.00	0.009
8	7.00	0.008
9	8.00	0.009
10	9.00	0.009
11	10.00	0.007

132
133 **Table 4: Reducing antioxidant power of methanol leaf extract of *R. communis*(Castor)**

S. No.	Volume of Leaf Extract (µL)	Absorbance of Leaf Extract (nm)
1	Methanol (control)	0.000
2	1.00	0.005
3	2.00	0.006
4	3.00	0.007
5	4.00	0.008
6	5.00	0.009
7	6.00	0.009

8	7.00	0.008
9	8.00	0.009
10	9.00	0.009
11	10.00	0.007

134
 135 From Tables 1 to 4 regarding antioxidant power of chloroform, ethanolic, ethyl acetate and
 136 methanolic leaf extracts of *R. communis* results reveal that the antioxidant power of chloroform,
 137 ethanolic, ethyl acetate and methanolic leaf extract of *R. communis* were found to be nearly equal.
 138 In chloroform extract (Table 1) highest absorbance was observed at 10.0 μ L concentration, while
 139 lowest at 1.0 μ L concentration of leaf extract was found to have maximum antioxidant power. In
 140 ethanolic extract (Table 2) highest absorbance was recorded at 8.0 μ L concentration, while
 141 lowest at 1.0 μ L concentration. Antioxidant power decreases after 8.0 μ L concentration. This
 142 may be due to decrease in hydrogen donor ability of phenolic compounds. In ethyl acetate extract
 143 (Table 3) minimum difference in the absorbance of leaves extract and control was observed at
 144 1.0 μ L concentration, while highest and same (0.009 nm) difference in absorbance was noted at
 145 5.0 μ L, 6.0 μ L, 8.0 μ L and 9.0 μ L concentrations. Methanolic leaves extract (Table 4) was found
 146 to have same antioxidant power or difference in absorbance from control as ethyl acetate. The
 147 maximum difference in absorbance (0.009 nm) was found for each solvent extracts. There is no
 148 definite order of increase or decrease in anti-oxidant power (from 1.0 μ L to 10.0 μ L
 149 concentration of extract) was observed in all solvent systems.

150
 151 **Table 5: Phytochemical analysis of Castor (*R. communis*) leaves extract**
 152

S.No	Phyto constituents	Ethanol	Methanol	Ethyl acetate	Chloroform
1	Alkaloids	-	-	-	-
2	Carbohydrate	+	+	+	+
3	Saponins	-	-	-	-
4	Protein and amino acids	-	+	-	-
5	Tannins	+	+	-	+
6	Flavonoids	+	-	+	-
7	Glycosides	-			
8	Terpenoids	+	-	-	+

153
 154 **Note:** - = Absent + = Present
 155

156 From the Table 5 results reveal that Phyto - constituent, alkaloids is absent in each four solvents
 157 (chloroform, ethanol, ethyl acetate and ethanol) extracts. Carbohydrate is present in the leaves
 158 extract of each solvent. Saponin is found to be negative in each four leaves extracts. Protein and
 159 amino acids are found to be present in methanolic extract while absent in chloroform, ethyl
 160 acetate and methanolic extract. Tannins were found to be positive in ethanolic, methanolic and
 161 chloroform extracts, while negative in ethyl acetate leaves extract. Flavonoids were found to be
 162 present in ethanolic and ethyl acetate leaves extract, while absent in methanolic and chloroform
 163 leaves extracts. Glycosides were found negative in ethanolic extract and could not be detected in
 164 methanolic, ethyl acetate and chloroform extracts. Terpenoids are found to be present in ethanolic
 165 and chloroform extracts and absent in ethyl acetate and methanolic extract.

166 **Table 6. Heavy metal analysis in Castor(*R. communis*) leaves in mg/ kg**
167

Name of plant	Used part	Metals	Mg/kg
Castor (<i>R. communis</i>)	Leaves	Zn	30.87
		Cu	2.14
		Ni	Nd
		Mn	21.64
		Fe	128.20
		Ca	84.42
		Mg	247.10

168
169 **Nd = Not detected**
170

171 Heavy metal analysis was done in leaves of *R. communis* by using Xplor AA – GOC Scientific
172 Equipment Atomic Absorption Spectrophotometer. It shows the highest amounts was that of Mg
173 (247 .10 mg/kg), followed by Fe (128.20mg/kg), whereas the least amount was that of Cu (2.14
174 mg/kg), but did not detect Ni metal.

175
176 **CONCLUSION:**

177 This study scientifically validates the used of the levers of *Ricinuscommunis* as antioxidant
178 power of chloroform, ethanolic , ethyl acetate and methanolic leaf extracts of *R. communis* results
179 reveal that the antioxidant power of chloroform, ethanolic, ethyl acetate and methanolic leaf
180 extract of *R. communis* were found to be nearly equal Phyto - constituent, alkaloids is absent in
181 each four solvents but carbohadrate present extract each solvent. In this study, we detected seven
182 components of heavy metals not previously reported, and confirmed the high Mg and Fe
183 presence in *R. communis* leaves. This information give light to the present intention to find
184 chemical proof that supports the pharmacological activities of *R. communis* leaves.

185
186 **REFERENCES:**

- 187
188 1. Roger, P., and R. Martyn, (1999). *Annuals and Biennials*, London, Macmillan, P. 106.
- 189 2. Tan, Q.G., X. –H. Cai, Z. –Z. Dua, and X. –D. Luo, (2009). Three terpenoids and a toxic
190 phenol
191 – related compound from *Ricinuscommunis*, *Helvetica ChimicaActa*92 (12) pp 2762 –
192 2768.
- 193
194 3. Srivastava. P., Jyotshna, N. Gupta, M. Kumar, K. Shankar, (2013). New anti-inflammatory
195 triterpenefrom the root of *Ricinucommunis*, *Natural Product Research*, (doi 10.1080 /
196 14786419.2013 861834).
- 197
198 4. Tunaru. S., T. F. Althoff, R. M. Nursing and M. Diener,(2012). Castor oil induces laxation and
199 uterus contraction via Ricinoleic acid activating prostaglandin EP3 receptors, *Proceedings of*
200 *the National Academy of Sciences of USA*; 109(23) pp 9179 – 9184.
- 201
202 5. Boel. M.E., S. J. Lee, M. J. Rijken, M. K. Paw and R. McGready, (2009). Castor oil for

- 203 induction of
204 labor, not harmful, not helpful, *Australian and New Zealand Journal of Obstetrics and*
205 *Gynecology*49 (5) pp 499 – 503.
- 206 6. Singh. R and Geetanjali, (2015) Phytochemical and pharmacological investigations of *Ricinus*
207 *communis*, *Algerian Journal of Natural Products*, 3: pp120 – 129.
- 208
209 7. Kumar. K., P. K. Sharma, R. Singh and S. H. Ansari,(2007) Antioxidant activity of the
210 methanolic extract of *R. communis* leaves, *Asian J. Chem.* 19 (5)pp 3387 – 3392.
- 211
212 8. Iqbal. J., S. Zaib, U. Farooq, A. Khan, I. Bibi and S. Suleman, (2012), Antioxidant,
213 antimicrobial
214 and free radical scavenging potential of *Perilocaaphylla* and *R. communis*, *ISRN –*
215 *pharmacology*, Volume, Article ID 563267 pp. 1-6
- 216
217 9. Vandita.V., A. Nirali, P. Khyati and K. Monisha, (2013).Effect of phytochemical constituents
218 of *R.communis*, *P. Santa linus*, *T. Belerica* on antibacterial, antifungal and cytotoxic activity,
219 *Int. J. Toxicol, Pharmaco. Res.*5(2)pp 47 -54.
- 220 10. Kadri, N. Gharsallah, M. Damak and R. Gidoura, (2013). Chemical composition and in vitro
221 antioxidant properties of essential oil of *Ricinuscommunis* L. *Journal of Medicinal Plant*
222 *Research* 5(8) (2011) pp 1466 – 1470.
- 223
224 11. Gulcin, I., M. Oktay, E. Kirecci and O. I. Kufreviogh,(2003). Screening of antioxidant and
225 antimicrobial activities of anise *Pimpinellaanisum* (L.) seed extracts, *Food Chem.* 83 (3)
226 pp 371 – 382.
- 227
228 12. Shimada.K., K. Fujikawa, K. Yahara and T. Nakamura, (1992), Antioxidant properties of
229 Xanthunon antioxidation of Soyabean oil in cyclodextrin emulsion. *J. Agri. Food Chem.* 40
230 pp 945– 948.
- 231
232 13. Duh. P. D., Antioxidant activity of burdock *Arctiumlappa* (L.) (1998). its scavenging effect
233 on free radical and active oxygen, *J. Am. Oil Chem. Soc.* 75pp 455 – 461.
- 234
235 14. Sawaddiwang.R., A. Jongjareorak and S. Bazakul, (2008). Phenolic content and antioxidant
236 activity of germinated brown rice as affected by germination temperature and extraction
237 solvent. *KMITL Sci. J.* 8 (2)pp 54 – 56.