

Original Research Article

Exploration of the antimicrobial properties of *Ficus exasperata* leaves from Akure

Metropolis.

Abstract

Aim: The study was conducted to explore the antibacterial and antifungal properties of leaf extracts of *Ficus exasperata* *in vitro*.

Study design: Extracts from *Ficus exasperata* leaves collected from Akure was qualitatively screened for the phytochemical constituents, and its' *in vitro* antimicrobial potency was evaluated against fourteen (14) fungal and bacterial isolates.

Results: The tested extracts contained tannins, flavonoids, terpenoids, alkaloids and cardiac glycosides whereas, saponin, steroids, phlobatannin and anthraquinone were absent. The acetone extract of the leaf demonstrated better antimicrobial activity against 10 of the test organisms. However, the highest antimicrobial activity (31.27mm) was exhibited by the methanol extract against referenced culture of *Staphylococcus aureus*. In addition, the extracts also displayed better antibacterial than antifungal activity. The minimum inhibitory concentration (MIC) of the extracts ranged between 0.391-1.563mg/mL, with the acetone extract displaying lower MIC values.

Conclusion: The occurrence of the observed phytochemicals in the extracts of *Ficus exasperata* could be involved in the antimicrobial efficacy of the plant. The result from this study thus supports the folkloric use of the plants. Additionally, the plant could also be exploited for the production of drugs especially for staphylococci infections.

Keywords: Medicinal plants; Antimicrobial; Phytochemicals; Extracts; *Ficus exasperata*

Introduction

For ages, mankind has faced a constant battle with infectious diseases. This has led to increased morbidity and mortality especially among population from developing countries. Many populations have adopted the traditional healthcare system as a way of preventing and treating diseases of microbial origin [1]. Traditional medicine remains the most sort after, as it is considered safer, affordable, and readily available [1].

Due to the upsurge in resistance to conventional drugs by microbial agents, novel antimicrobial agents from different biological sources have been sort after and reported to be effective in combating pathogenic organisms. The use of herbal remedies containing plants or part of plants has in recent years gained ground in developed countries [2]. Pharmaceutical companies have thus developed new antimicrobial drugs and also improved on the existing ones through the modification of their structures with a view to increasing their efficacy [3].

Ficus exasperata otherwise known as the sandpaper tree is native to tropical Africa [4]. The leaves of *Ficus exasperata* have been employed in folkloric medicine for the treatment of various diseases such as ophthalmic and oral infections, venereal diseases, parasitic infection (cutaneous, subcutaneous), leprosy, and malaria [5,6]. The study therefore investigates the claim of the antimicrobial potential of *Ficus exasperata*, in a bid to develop novel antimicrobials.

Materials and Methods

Collection and preparation of extracts from leaves Samples

The leaves of *Ficus exasperata* were collected from its tree at a building near Life Spring College, Apatapiti layout, Federal University of Technology, Akure, Ondo State (Latitude:7.289N, Latitude:5.150E) Nigeria in the month of April, 2015. Samples of the leaves were taken to the Department of Crop, Soil and Pest, FUTA for authentication. Afterwards the leaves were cleansed with water, shade dried, grinded and stored in airtight container. Thereafter, the powdered leaves (100g) of *Ficus exasperata* was weighed separately into different plastic containers and 1000mL of 100% acetone and methanol added to the containers for extraction. Aluminium foil was placed on each container before covering. Each solution was allowed to stand for 3 days with continuous stirring. The extracts were thereafter obtained by filtering the solutions through a funnel fitted with a filter paper. The filtrates were thereafter evaporated to dryness at 50 °C in a rotary evaporator (RE-52A; Union Laboratory, England) with 90 rpm under reduced pressure. The obtained concentrated extracts were stored in dark at 4 °C until further analysis.

Phytochemical screening of leaf extract of *Ficus exasperata*

The plant extracts were subjected to qualitative phytochemical screening using standard protocols described by Odebiyi and Sofowora [7], Trease and Evans [8], and Harborne [9].

62 Measurement of antibacterial and antifungal activities of leaf of *Ficus exasperata*

63 Varying concentrations of the leaf extracts of *Ficus exasperata* (3.125-100mg/mL) were
 64 prepared by dissolving different amount of the extracts in 5mL of 30% tween 20. For
 65 example, concentrations of 100, 50, and 25mg/mL were prepared by dissolving 500, 250 and
 66 125mg of the extracts into 5ml of 30% tween 20 respectively. Afterwards the prepared
 67 extracts were sterilized by passing them through a 0.22µm millipore membrane filter. The
 68 agar well diffusion method as described by Schinor *et al.* [10] was employed in assessing the
 69 antimicrobial activity of *Ficus exasperata* leaf extracts. A total of 14 clinical and referenced
 70 microbial strains were used for the experiment. The test organisms were obtained from the
 71 Pathology and Clinical Laboratory (PATHCARE), Lagos State University Teaching Hospital,
 72 Lagos State, Nigeria and the Department of Microbiology, FUTA. Active broth cultures of
 73 the test organisms were prepared from stock cultures. An aliquot of 100µL of bacterial and
 74 fungal solution was evenly spread on already solidified Mueller Hinton agar plates.
 75 Afterwards, wells of 7mm diameter were bored in the solidified Mueller Hinton agar plates
 76 using a sterile cork-borer. Thereafter, an aliquot of 100uL of the sterilized extract was added
 77 into the bored agar wells. The plates were thereafter incubated at 37 °C for 24 hour for
 78 bacteria and at 26 ± 1°C for 48 to 72 hours for fungi. The plates were observed for clear
 79 zones of inhibition and the measurements tsaken using a ruler calibrated in millimetres.
 80 Commercial antifungal drugs (clotrimazole, nystatin and gluseofluvin) and commercial
 81 antibacterial drug (ciprofloxacin (10µg), rocephin (25µg)) were used as the positive control,
 82 while 30% tween 20 was used as the negative control.

83 Statistical analysis

84 Experiments were carried out in triplicates were applicable. The results were expressed as
 85 mean ± standard error of three values. Data analysis was carried out using the One Way
 86 Analysis of Variance (ANOVA) and treatment means were compared using New Duncan's
 87 Multiple Range Test (SPSS version 16). Differences were considered significant at P<0.05.

88 Results

89 Table 1 shows the presence of tannin, flavonoid, terpenoids, alkaloids and cardiac glycosides
 90 in *Ficus exasperata* leaf extracts, and the absence of saponin, steroids, phlobatannin and
 91 anthraquinone.

93

94 **Table 1:** Qualitative phytochemical screening of *Ficus exasperata* leaf extracts

Phytochemical	Extracts	
	FEM	FEA
Saponin	-	-
Tannin	+	+
Flavonoid	+	+
Steroids	-	-
Terpenoids	+	+
Alkaloids	+	+
Phlobatannin	-	-
Anthraquinone	-	-
Cardiac Glycosides		
Legal test	+	+
Keller kiliani	+	+
Salkowski	+	+
Lieberman test	+	+

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96 **KEYS:** FEA: Acetone leaf extract of *Ficus exasperata*; FEM: Methanol leaf extract of *Ficus*
97 *exasperata*.

98 The antimicrobial activity of the leaf extracts of *Ficus exasperata* shows that the acetone leaf
99 extract exhibited better activity against most of the test organisms used for the study (Table
100 2). However, the highest antimicrobial activity (31.27mm) was exhibited by the methanol
101 extract of *Ficus exasperata* against referenced *Staphylococcus* and this was found to be
102 slightly higher that observed in the acetone extract (29.40mm) against the same organism. In
103 like manner, the leaves extracts displayed better antibacterial than antifungal activity. The
104 antifungal activity of the acetone extract of the plant was however a better than that of the
105 methanol extract.

106 Upon comparison of the activities of the leaf extracts against organism with Gram reaction
107 positive and Gram negative bacterial isolaes, the Gram positive organism were more
108 susceptible than the Gram negative organism in most cases. The extracts antibacterial activity
109 was comparatively better than that of the commercial antibacterial drugs in most of the tested
110 organisms. Reverse was the case for the commercial antifungal drugs as they exhibited better
111 activity than the extracts. The acetone extract of *Ficus exasperata* was found to exhibit lower
112 minimum inhibitory concentration values than the methanol extracts. The results are
113 displayed in Table 3.

114 **Table 2:** Antimicrobial activity of leaves extracts of *Ficus exasperata* and commercial drugs

Test Organism	Zone of inhibition (mm)						
	FEA	FEM	CPX	R	CLOT	GRIS	NYST
<i>Salmonella</i> Typhi (ATCC 33489)	15.20 ± 0.12 ^c	12.43 ± 0.18 ^a	12.27± 0.15 ^a	14.30±0.12 ^b	NT	NT	NT
<i>Salmonella</i> Typhi	18.43 ± 0.18 ^c	12.47 ± 0.15 ^a	12.40±0.23 ^a	14.37±0.20 ^b	NT	NT	NT
<i>Staphylococcus aureus</i> (ATCC 43300)	29.40 ± 0.17 ^c	31.27 ± 0.15 ^d	14.40±0.12 ^a	15.50±0.17 ^b	NT	NT	NT
<i>Staphylococcus aureus</i>	27.33 ± 0.24 ^b	28.40 ± 0.12 ^c	15.43±0.20 ^d	12.50±0.12 ^a	NT	NT	NT
<i>Escherichia coli</i> (ATCC 35218)	22.20 ± 0.12 ^b	15.53 ± 0.15 ^a	11.20±0.12 ^b	14.27±0.15 ^c	NT	NT	NT
<i>Escherichia coli</i>	17.50 ± 0.26 ^{ab}	15.60 ± 0.17 ^{ab}	12.50±0.23 ^{ab}	49.97±36.67 ^a	NT	NT	NT
<i>Pseudomonas aeruginosa</i> (ATCC 27853)	12.27 ± 0.15 ^c	15.40 ± 0.12 ^b	15.43±0.15 ^a	16.33±0.18 ^b	NT	NT	NT
<i>Shigella dysenteriae</i>	17.40 ± 0.21 ^a	16.30 ± 0.12 ^b	14.40±0.12 ^b	14.27±0.15 ^c	NT	NT	NT
<i>Bacillus cereus</i>	15.60 ± 0.17 ^c	20.40 ± 0.12 ^d	12.33±0.18 ^a	14.53±0.20 ^b	NT	NT	NT
<i>Bacillus subtilis</i>	21.30 ± 0.12 ^d	12.43 ± 0.15 ^a	14.33±0.15 ^b	15.50±0.12 ^c	NT	NT	NT
<i>Candida albicans</i>	18.37 ± 0.23 ^d	10.27 ± 0.15 ^b	NT	NT	16.65 ± 0.68 ^c	20.50 ± 0.29 ^e	6.40 ± 0.21 ^a
<i>Aspergillus niger</i>	15.60 ± 0.17 ^b	3.47 ± 0.20 ^a	NT	NT	22.33 ± 0.33 ^d	21.67 ± 0.33 ^d	17.47 ± 0.32 ^c
<i>Aspergillus flavus</i>	12.40 ± 0.17 ^c	3.30 ± 0.12 ^a	NT	NT	25.00 ± 0.15 ^e	9.77 ± 0.15 ^b	18.73 ± 0.22 ^d
<i>Aspergillus fumigatus</i>	13.30 ± 0.17 ^c	5.30 ± 0.15 ^a	NT	NT	35.67 ± 0.44 ^e	9.33 ± 0.44 ^b	20.57 ± 0.30 ^d

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116 Each value is expressed as mean ± standard error (n = 3). Values with different superscript within a row are significantly different at ($P=0.05$).

117 **Keys:** **FEA:** Acetone leave extract of *Ficus exasperata*; **FEM:** Methanol leave extract of *Ficus exasperata*; **CPX:** Ciprofloxacin (10ug); **R:**

118 Rocephin (25ug); **CLOT:** Clotrimazole; **GRIS:** Griseofluvin; **NYST:** Nystatin; **NT;** Not tested.

119 **Table 3:** Minimum inhibitory concentration (mg/ml) of leaf extracts of *Ficus exasperata*

Test organisms	MIC (mg/mL)	
	FEA	FEM
<i>Salmonella typhi</i> (ATC 33489)	0.781	0.781
<i>Salmonella typhi</i>	0.391	1.563
<i>Staphylococcus aureus</i> (ATC 43300)	0.391	0.781
<i>Staphylococcus aureus</i>	0.391	1.563
<i>Escherichia coli</i> (ATC 35218)	0.391	0.781
<i>Escherichia coli</i>	0.391	0.391
<i>Pseudomonas aeruginosa</i> (ATC 27853)	0.781	0.781
<i>Shigella dysenteriae</i>	0.781	1.562
<i>Bacillus cereus</i>	0.391	0.391
<i>Bacillus subtilis</i>	0.391	1.562
<i>Candida albicans</i>	0.391	0.391
<i>Aspergillus niger</i>	0.391	0.391
<i>Aspergillus flavus</i>	0.391	1.563
<i>Aspergillus fumigatus</i>	0.391	1.563

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121 **Keys:** FEA: Acetone leave extract of *Ficus exasperata*; FEM: Methanol leave extract of
122 *Ficus exasperata*

123 Discussion

124 Plants remain an inexhaustible source of novel antimicrobials. Africa with its tropical and
125 subtropical climate is richly blessed with an array of plants that have naturally acquired
126 secondary metabolites in order to survive the harsh environment [1, 11]. Compounds with
127 antimicrobial properties that also offer protection against drug resistant microorganisms have
128 been isolated from medicinal plants [12, 13]. The present study investigated the secondary
129 metabolite profile and antimicrobial efficacy of leaves of *Ficus exasperata* collected from
130 Akure.

131 The presence of the observed secondary metabolites in the leaf extracts validates the
132 medicinal potentials of this plants as these compounds have been reported to play a protective
133 role against pathogenic organisms [13]. The absence of saponin, steroids, phlobatannin and
134 anthraquinone in the extracts might be attributed to solubility of the compounds in the
135 extraction solvent used.

136 The antimicrobial activity of the extracts could be attributed to the observed phytochemicals
137 in the extracts. In addition, the variation observed in the antimicrobial activity of the extracts

might be linked to differences in the type and amount of phytochemicals present in the extracts. The structural differences in the cell wall of Gram positive and Gram negative bacteria may account for the higher susceptibility of Gram positive bacteria to the plant extracts. The complexity in the cell wall Gram negative bacteria gives them better buffering capacity thus making their cell wall less impermeable, whereas Gram positive bacteria have only an outer peptidoglycan cell wall which makes them more susceptible [14].

The higher antibacterial activity demonstrated by the extracts than antifungal activity is in consonance with findings of several authors [15, 16] that have reported higher sensitivity of bacteria to antimicrobials. The chitinous cell wall of fungi promotes lesser susceptibility to antimicrobials than bacteria [17]. Antibiotics have been mostly reported to produce better performance against microorganisms than plants as a result their higher purity and smaller molecular sizes which aid their penetration into the cell wall of the organisms [18]. The better activity produced by the extract suggests that they can be explored for potent antimicrobial compounds.

Conclusion

The results obtained from the study support the possible use of leaves of *Ficus exasperata* in folkloric medicine. The plant extracts produced an effective performance against the growth of the tested organisms and could also be exploited for the production of drugs especially for staphylococci infections.

References

1. Mahomoodally MF. Traditional Medicines in Africa: An Appraisal of Ten Potent African Medicinal Plants. *Evidence-Based Complementary and Alternative Medicine*, 2013; Article ID 617459, 14 pages,
2. Chintamunnee V, Mahomoodally MF. Herbal medicine commonly used against infectious diseases in the tropical island of Mauritius. *Journal of Herbal Medicine*, 2012; 2:113–125.
3. Silva NCC, Fernandes JA. Biological properties of medicinal plants: a review of their antimicrobial activity. *The Journal of Venomous Animals and Toxins including Tropical Diseases*, 2010; 16(3): 402-413
4. Arbonnier M. **Trees, shrubs and lianas of West African dry zones**. Margraf Publishers, Paris, France; 2004.

- 169 5. Titanji VP, Zofou D, Ngemenya MN. The antimalarial potential of medicinal plants
170 used for the treatment of malaria in Cameroonian folk medicine. *Afr J Tradit*
171 *Complement Altern Med.*, 2008; 5:302-21.
- 172 6. Joseph B, Raj SJ. Phytopharmacological and phytochemical properties of three *Ficus*
173 species - An overview. *Int J Pharma Bio Sci*, 2010;1:246-53.
- 174 7. Odebiyi OO, Sofowora EA. Phytochemical screening of Nigerian medicinal plants.
175 *Lloyd*. 1978;41(3):234–246.
- 176 8. Trease GE, Evans MC. **Pharmacognosy** (14th ed.). New Delhi, India: Elsevier; 2005.
- 177 9. Harborne JB. **Phytochemical methods. A guide to modern techniques of plant**
178 **analysis** (3rd ed.). New Delhi: Springer Pvt Ltd; 2005.
- 179 10. Schinor EC, Salvador MJ, Ito IZ, Dias DA. Evaluation of the antimicrobial activity of
180 crude extracts and isolated constituents from *Chresta scapigera*. *Braz J Microbiol*,
181 2007; **38**: 145-149.
- 182 11. Manach C, Scalbert A, Morand C, Rémésy C, Jiménez L. Polyphenols: food sources
183 and bioavailability. *American Journal of Clinical Nutrition*, 2004; 79(5):727–747.
- 184 12. Miyasaki Y, Rabenstein JD, Rhea J, Crouch ML, Mocek UM, Kittell PE, et al.
185 Isolation and Characterization of Antimicrobial Compounds in Plant Extracts against
186 Multidrug-Resistant *Acinetobacter baumannii*. *PLoS ONE*, 2013;8(4): e61594.
- 187 13. Manganyi MC, Regnier T, Olivier EI. Antimicrobial activities of selected essential
188 oils against *Fusarium oxysporum* isolates and their biofilms. *South African Journal of*
189 *Botany*, 2015; 99:115–121.
- 190 14. Tadeh H, Mohammed E, Asres K, Gebre-Mariam T. Antimicrobial activities of
191 some selected traditional Ethiopian medicinal plants used in the treatment of skin
192 disorders. *J Ethnopharmacol*, 2005;100:168-175.
- 193 15. Modarresi-Chahardehi A, Ibrahim D, Fariza-Sulaiman S, Mousavi L. Screening
194 antimicrobial activity of various extracts of *Urtica dioica*. *Revista de Biología*
195 *Tropical*, 2012; 60(4):1567-1576.
- 196 16. Momoh AO, Oladunmoye MK, Adebolu TT. Evaluation of the Antimicrobial and
197 Phytochemical Properties of Oil from Castor Seeds (*Ricinus communis* Linn). *Bulletin*
198 *of Environment, Pharmacology and Life Sciences*, 2012: 1(10);21-27
- 199 17. Madigan MT, Martinko JM. **Brock Biology of Microorganisms**. New Jersey, USA,
200 Pearson-Prentice Hall; 2006
- 201 18. Raza MW, Shad A, Pedler SJ, Karamat KA. Penetration and activity of antibiotics in
202 brain abscess. *JCPSP*, 2005;15(3):165-167.