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***Original Research Article***

# Improving the Slow Release System Using Chitosan-Alginate Nanoparticles with Various Methods for Curcumin


## ABSTRACT

Biodegradable polymeric materials have been used as a drug delivery system due to its encapsulation, release system and less toxic properties. Chitosan-alginate nanoparticles were prepared by ionotropic pre-gelation of alginate core, followed by polyelectrolyte complexation of chitosan and curcumin was chosen as a model drug.

**Aims:** The aim of this study was to evaluate the possibility of chitosan-alginate nanoparticles as carriers for curcumin using various methods.
**Study design:** Lab. Experience
**Place and Duration of Study:** Department of Chemistry (LPPT and Analytical Laboratory), Universitas Gadjah Mada, Special Region of Yogyakarta, between September 2013 and May 2014.
**Methodology:** This preparation was divided into 4 methods to produce Alginate-Calcium-Chitosan (ACaK), Alginate-Chitosan-Calcium (AKCa), Chitosan-Alginate-Calcium (KACa) and Alginate-Chitosan (AK) nanoparticles. The amount of drug entrapped was determined by shaking the solution and then supernatant was analyzed for drug loading by measuring the absorbance, morphology, particle sizes and kinetics.
**Results:** The encapsulation efficiency was up to 85% in higher drug-polymer ratio and the effect of egg-box model made diameter of nanoparticles between 51 and 90 nm. The kinetic model of desorption for Alginate-Calcium-Chitosan, Alginate-Chitosan-Calcium and Chitosan-Alginate-Calcium nanoparticles followed Korsmeyer-Peppas.
**Conclusion:** This study suggest that these materials serve as suitable for uptake in capillary due to their nano-sized range and could stay long time into bloodstream.


*Keywords: curcumin, release system, drug delivery, nanoparticles, kinetic*

## 1. INTRODUCTION

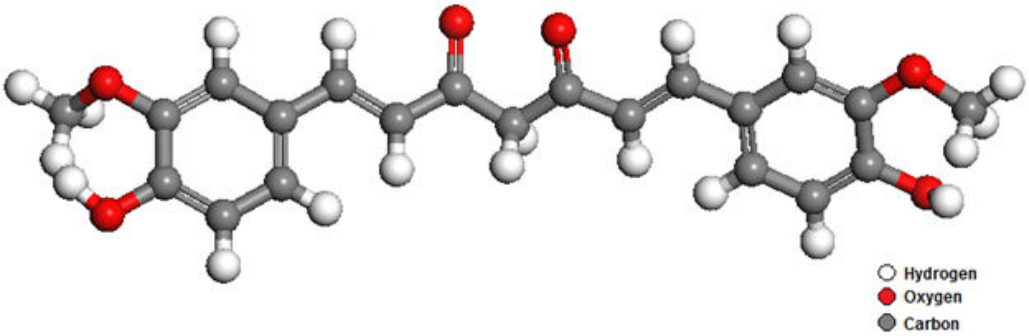



**Fig. 1. Chemical structure of curcumin.**

Curcumin or (Diferuloylmethane) is a polyphenol derived from the herb Curcuma longa and commonly known as turmeric [1](Fig. 1). Curcumin has anti-carcinogenic [2-5], anti-HIV [6], anti-inflammatory [7-

UNDER PEER REVIEW

12], antioxidant [13-15] and antitumor properties [16]. Despite extensive research and development, the properties of curcumin in aqueous solution were low bioavailability and poor solubility [17]. Therefore, curcumin is often limited by its potential to reach the site of therapeutic action. Curcumin has been reported that it is complex with cyclodextrin [18] and phospholipids [19] or encapsulated in solid lipid microparticles such as chitosan [20] and albumin [21]. A drug delivery system has opened up new possibilities in targeted and sustained release of drugs by nanotechnology [22]. A drug delivery system was designed by tiny nanoparticles and it can reach less accessible sites in the body. Controlled release of the drug from the nanoformulations could maintain steadier levels of drug in bloodstream for longer durations. Sustained release of drug can be achieved by encapsulation the active ingredient in a polymer matrix.

Chitosan (K) and sodium alginate (A) are extensively used in encapsulation of drug for the purpose of the sustained release [23]. Sodium alginate is an anionic polysaccharide obtained from marine algae and it is a water-soluble natural copolymer composed of guluronic and mannuronic acid units. In addition, chitosan is deacetylate derivatives of chitin, composed by β-(1, 4)-2-amino-2-deoxy-D-glucopyranose units and small amount of N-acetyl-D-glucosamine residues [24]. These are polysaccharide polymers, formed from repeating units (either mono- or di- saccharides) joined together by glycosidic bond [25]. They have many properties of an ideal carrier for drug delivery such as biocompatibility, biodegradability, low cost and non-toxicity [26]. Encapsulation of hydrophobic drugs into an aqueous nanoparticulate system has been attempted so as to deliver such drugs with their full potential [27]. Rajaonarivony et al. reported that poly-L-lysine/alginate nanoparticles were produced using a pre-gel method [28].

The objective of this study was to evaluate the possibility of chitosan-alginate nanoparticles as carriers for curcumin. The challenge was to entrap hydrophobic molecules such as curcumin into hydrophilic nanoparticles formed by ionotropic pre-gelation of the alginate core followed by chitosan polyelectrolyte complexation. The final samples were characterized using Fourier Transform Infrared spectrometer (FTIR), Scanning Electron Microscope (SEM) and Transmission Electron Microscope (TEM), respectively. In addition, curcumin release was analyzed using Ultraviolet-Visible (UV-Vis). The release properties of nanoparticles in ratio of phosphate buffered saline and ethanol (80:20) at pH 7.4 were also studied.

## 2. MATERIAL AND METHODS / EXPERIMENTAL DETAILS / METHODOLOGY

### 2.1 Preparation of Curcumin Loaded Chitosan-Alginate Nanoparticles

Sodium alginate and calcium chloride solutions were prepared by dissolving the chemicals in distilled water. However, chitosan was dissolved in 1 % acetic acid solution and curcumin was dissolved in ethanol. The pH of the sodium alginate and chitosan solution was adjusted to 5 using hydrochloric acid and sodium hydroxide, respectively.

This preparation was divided into 4 methods to produce Alginate-Calcium-Chitosan (ACaK), Alginate-Chitosan-Calcium (AKCa), Chitosan-Alginate-Calcium (KACa) and Alginate-Chitosan (AK) nanoparticles. Calcium chloride solution 18 mM (2.5 mL) was added dropwise to 25 mL aqueous sodium alginate 0.12% (w/v) while stirring for 30 minutes. Two mL of curcumin solution 1% (w/v) and 10 mL chitosan solution 0.05% (w/v) were added into the resultant calcium alginate pre-gel and stirred for an hour called as ACaK. AKCa were obtained as ACaK, but calcium chloride solution was added after mixing alginate and chitosan solution. KACa were obtained as AKCa, but 10 mL alginate solution was added into 25 mL of chitosan solution. Chitosan solution (10mL) and 23 mL distilled water were added into 25 mL alginate solution and stirred for an hour called as AK.

The resultant suspensions were equilibrated overnight at room temperature to allow nanoparticles and to form uniform particle sizes and then separated by centrifugation at 4℃. Particles were dried using a freeze dryer overnight. Blank nanoparticles were obtained as a previous procedure of curcumin loaded nanoparticles, but they were not added by curcumin solution. They were called as AK-0, ACaK-0, AKCa-0 and KACa-0.

The amount of drug entrapped was determined by shaking the solution and then supernatant was analyzed for drug loading by measuring the absorbance at 426 nm in UV spectrophotometer. The

UNDER PEER REVIEW

confirmation of drug loading was done by comparing optical absorbance of nanoformulations with and without drug loading.

% encapsulation=(drug input(mg)-drug in supernatant(mg))/(drug input (mg)) x 100%

The morphology, measurements of particle sizes and functional groups of the nanoparticles were performed using Scanning Electron Microscope (SEM) (JEOL, JSM-6510), Transmission Electron Microscope (TEM) (JEOL, JEM-1400) and Fourier Transform Infrared spectra (FTIR), respectively. Sample preparation using FTIR was taken with KBr pellets on a FTIR spectrometer (Shimadzu-Prestige 21).

### 2.2 In Vitro Release of Curcumin

To establish the curcumin release profiles from nanoparticles, the curcumin loaded nanoparticles were separated from matrix by shaking at 37℃ in various times. At appropriate intervals, five mL solution was taken and analyzed using UV spectrophotometer. The mathematical models such as zero-order, first-order, Higuchi, and Korsmeyer-Peppas equations were fitted to individual dissolution data.

Zero-order model is used to describe the drug solution of modified release drug slowly. It can be represented by the question: Q0-Qt = K0·t, where K0 is the zero order release constant expressed in units of concentration/time, Q0 is the initial amount of drug in the solution and Qt is the amount of drug dissolved in time t. Data obtained from in vitro drug release studies were plotted as cumulative amount of drug release vs time [29].

First order model has also been used to describe absorption and/or elimination of some typical drugs. This release model can be expressed by the equation: dC/dt = -Kc, where K is the first order rate constant expressed in units of time-1, t is the time and C is concentration of drug. The data obtained were plotted as log cumulative percentage of drug remaining vs time which would yield a straight line with a slope of (- K)/2.303 [30].

Higuchi model is to describe drug release from a matrix system and based on hypotheses that (i) initial drug concentration in the matrix is much higher than drug solubility; (ii) drug diffusion takes place only in one dimension (edge effect must be negligible); (iii) drug particles are much smaller than system thickness; (iv) matrix swelling and dissolution are negligible; (v) drug diffusivity is constant; and (vi) perfect sink conditions are always attained in the release environment. To express model is given by the equation: ft = Q = A √(D(2C-Cs) Cs·t), where C is the drug initial concentration, Cs is the drug solubility in the matrix media, D is the diffusion coefficient in the matrix substance, ft or Q is the amount of drug released in time (t) per unit area (A) and t is time. In addition, the model is possible to simplify the Higuchi model as ft = Q = KH·t1/2 [31], where KH is the Higuchi dissolution constant [32]. The data obtained were plotted as cumulative percentage drug release vs square root of time [33].

Korsmeyer-Peppas model has been used to describe drug release from a polymeric system equation. This equation can be expressed as Mt / M∞ = K·tn, where k is the release rate constant, Mt / M∞ is a fraction of drug released at time (t) and n is the release exponent. The n value is used to characterize different release for cylindrical shaped matrices [34]. In this case of cylindrical tablet, 0.45 ≤ n corresponds to a Fickian diffusion mechanism, 0.45 < n < 0.89 to non-Fickian transport, n = 0.89 to case II transport (relaxation), and n > 0.89 to super case II transport [35]. To study the release kinetics, data obtained from in vitro drug release studies were plotted as log cumulative percentage drug release vs log time. To find out the mechanism of drug release, the first 60% drug release data should be used.

## 3. RESULTS AND DISCUSSION

### 3.1 Preparation of Chitosan-Alginate Nanoparticles

Chitosan-alginate nanoparticles were obtained by spontaneously under very mild conditions. Preparation of chitosan-alginate nanoparticles appeared some typical peaks. IR spectra of chitosan (Fig. 2(a)) showed the broad band at 3448 $cm^{-1}$ corresponded to the amine and hydroxyl groups. The absorption bands of carbonyl stretching of the secondary amide (amide I band) and the bending vibrations of the N-H (amide II band) were at 1651 $cm^{-1}$ and 1597 $cm^{-1}$, respectively. The peak of secondary hydroxyl group observed was at 1087 $cm^{-1}$. The band in the IR spectra of sodium alginate

UNDER PEER REVIEW

(Fig. 2(b)) at 1419 $cm^{-1}$ and 1612 $cm^{-1}$ was assigned to symmetric and asymmetric stretching peaks of carboxylate salt groups. In addition, the absorption band of alginate at 1612 $cm^{-1}$ shifted to 1627 $cm^{-1}$ after reacting with chitosan (Fig. 2(c)) as polyelectrolyte complexation. The AK-0 was used to control between other samples added by calcium ions (Ca). FTIR spectra of sodium alginate and calcium ions appeared a specific peak at 1080 $cm^{-1}$ indicating hydroxyl groups interacted with calcium ions. The peaks of ACaK-0 and KACa-0 were observed at 1620 $cm^{-1}$ and 1604 $cm^{-1}$, respectively (Fig. 2(d and e)). They indicate that calcium ions interacted with carboxylate ions of sodium alginate to form an egg-box model. The IR spectra of AKCa-0 (not shown) had identical peaks with ACaK-0.

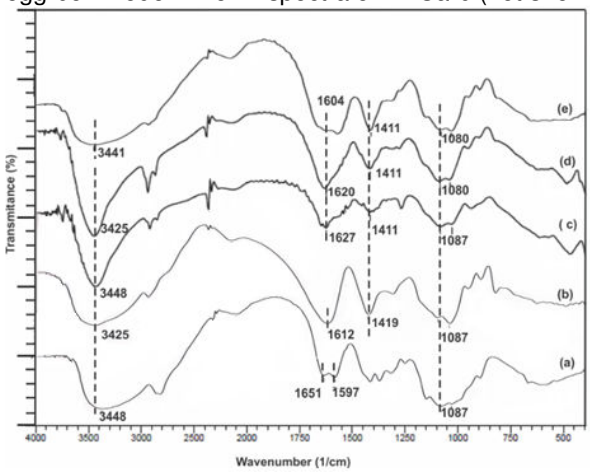



**Fig. 2. FTIR spectra of chitosan (a), sodium alginate (b), AK-0 (c), ACaK-0 (d) and KACa-0 (e).**

The FTIR was adopted to characterize the potential interaction between 2 polymers and calcium ions into nanoparticles. As shown in Fig. 3 (a), IR spectra of curcumin had specific peaks at 1427 $cm^{-1}$ and 1512 $cm^{-1}$ as aromatic rings, Carbon double bond was at 1627 $cm^{-1}$, methoxy stretching peak appeared at 2854 $cm^{-1}$ and a broad band at 3510 $cm^{-1}$ was attributed to the stretching vibration of the OH groups. IR spectra of AK that observed the asymmetrical stretching of carboxyl groups at 1627 $cm^{-1}$ shifted to 1635 $cm^{-1}$ (Fig. 3 (b and c)), which probably indicates that curcumin molecule was filled in the polymeric network. In addition, these results indicate that carboxylate groups of alginate associate with ammonium groups of chitosan through electrostatic interactions to form the polyelectrolyte complexation. The IR spectra of ACaK-0 and ACaK at 1620 $cm^{-1}$ shifted to 1627 $cm^{-1}$ and 3425 $cm^{-1}$ to 3433 $cm^{-1}$ (Fig. 3 (d and e)). In addition, IR spectra of KACa-0 and KACa at 1604

UNDER PEER REVIEW

$cm^{-1}$ shifted to 1627 $cm^{-1}$ and 3441 $cm^{-1}$ shifted to 3433 $cm^{-1}$ (Fig.3 (f and g)).

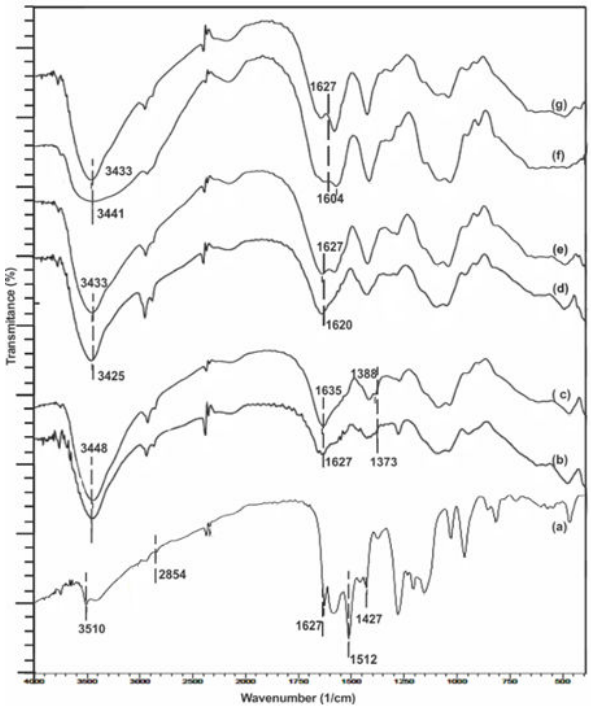



**Fig. 3. FTIR spectra of curcumin (a), AK-0 (b), AK (c), ACaK-0 (d), ACaK (e), KACa-0 (f) and KACa (g).**

An analysis of electron microscopy was confirmed the presence of nanoparticles and provided morphological information of the typical curcumin loaded chitosan-alginate nanoparticles using the transmission electron microscope (TEM) and scanning electron microscope (SEM), respectively. Fig. 4 showed the comparison of 2 methods between ACaK and AKCa because calcium ion had unique characteristics to design small particles. Particle sizes were about between 51 and 90 nm depending on methods and particles were seen to be spherical solid. The interaction between alginate and calcium ions produced egg-box model and the diameters were smaller than without producing egg-box model because calcium-alginate complex in pre-gel state was enveloped by chitosan as cationic polymer. Fig. 5 showed appearances in the form of dry solids between with and without calcium ions. If they were dispersed, they would be able to decompose easily into nanometer sizes. Encapsulation efficiencies of ACaK, AKCa and KACa were 85.38, 65.00 and 73.22%, respectively (Fig. 6). The result shows calcium ions interacted between alginate and chitosan because the matrix produced egg-box model and nano-sized particles. Therefore, these nanoparticles are effective to entrap curcumin as a drug model.

UNDER PEER REVIEW

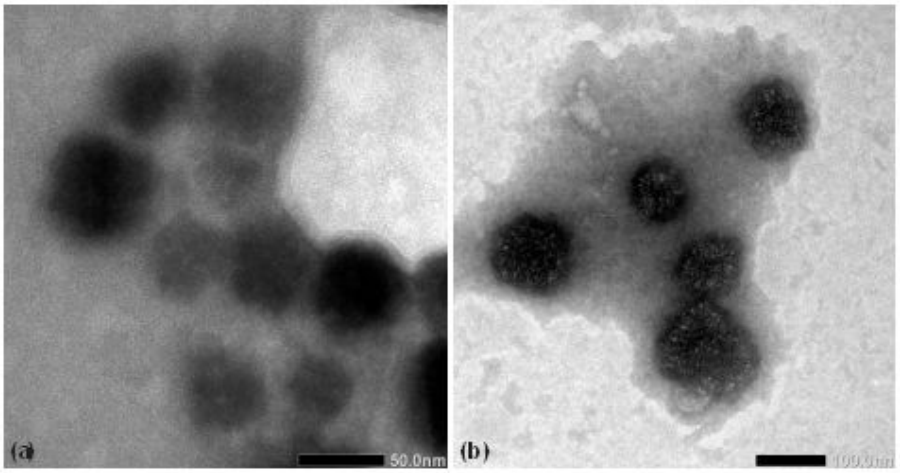



**Fig. 4. TEM Images of curcumin loaded nanoparticles, ACaK (a), AKCa (b).**

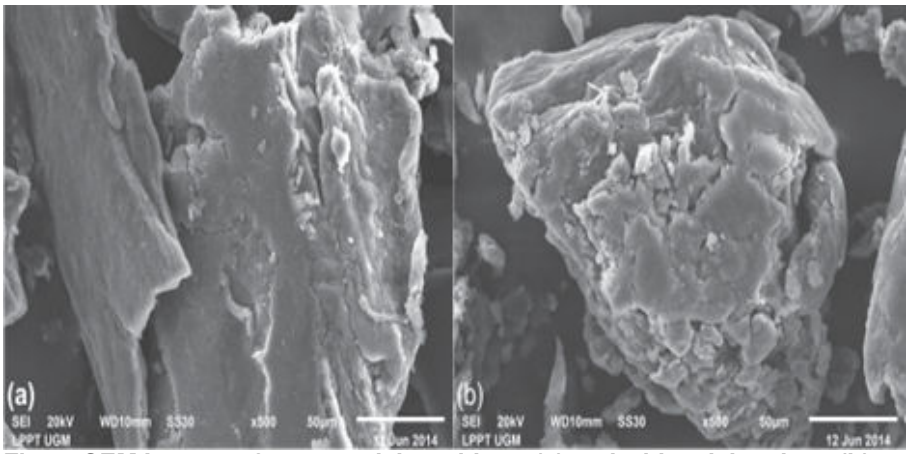
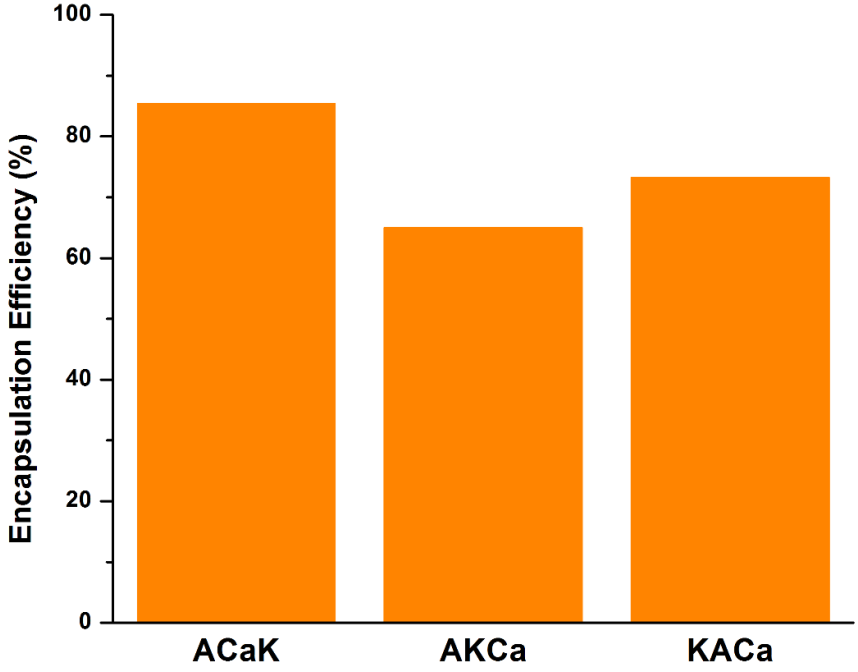



**Fig. 5. SEM Images of nanoparticles without (a) and with calcium ions (b).**



UNDER PEER REVIEW

**Fig. 6. Encapsulation efficiency of ACaK, AKCa and KACa nanoparticles.**

### 3.2 In Vitro Release of Curcumin

The release of ACaK has been reached the slowest because ACaK produced egg-box models that have an ability to control curcumin release from matrix slowly, but AKCa has the fast curcumin release because they could not produce egg-box models well and the effect is that only little curcumin entrapped into matrix between alginate and chitosan. In many experimental situations, including the case of drug release from swellable polymeric systems, the mechanism of drug release diffusion deviates from the Fickian equation and follows a non-Fickian behavior (anomalous). In the cases, the following general equation or its logarithmic form can be used as cylindrical tablets, values of n between 0.45 and 0.89 indicate both diffusion and swelling controlled drug release (anomalous transport). Values above 0.89 indicate case-II transport which related to polymer relaxation during hydrogel swelling and values below 0.45 indicate that drug release from polymer is due to Fickian diffusion.

The n and k values were obtained from releasing curve (Fig. 7) and presented in Table 1. The n value is the diffusional exponent that can be related to the drug transport mechanism. The ACaK, AKCa and KACa fitted to Korsmeyer-Peppas' model. The values of n for ACaK, AKCa and KACa were 0.591, 0.333 and 0.283, respectively. The ACaK mechanism followed diffusion and erosion, but both AKCa and KACa mechanisms only followed diffusion. Therefore, ACaK was very effective to be a drug release system because a drug model could release slower than others and then the matrix will be broken by erosion after releasing curcumin slowly. In addition, the ACaK produced egg-box models easily because calcium ions were interacted into carboxylic groups of alginate well, but the first interactions between alginate and chitosan of AKCa and KACa did not produce egg-box models because carboxylate of alginate have interacted with amine of chitosan to form polyelectrolyte complex. Therefore, calcium ions did not have ability to interact into functional group of alginate and ethanol was added into phosphate buffered saline to increase the solubility of curcumin, then the kinetic model could be reached gap clearly than others.

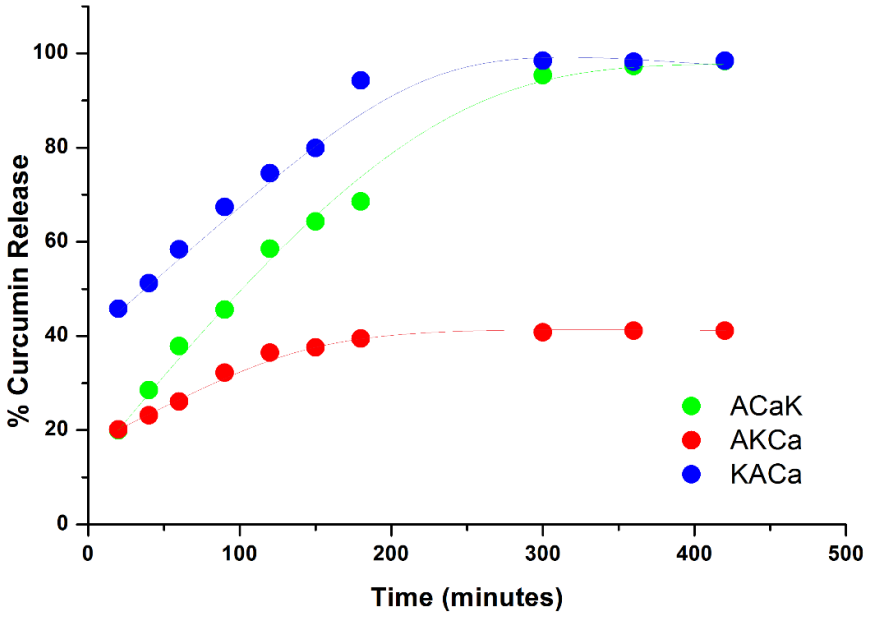



**Fig. 7. In vitro curcumin release (%) from ACaK, AKCa and KACa in mixing PBS and ethanol solution at 37℃**

UNDER PEER REVIEW

**Table 1. Estimated parameters and drug release mechanisms of curcumin loaded chitosan-alginate nanoparticles**

| Nanoparticle | Kinetic Model | Kinetic constant (k) | $r^2$ | Diffusional exponent (n) |
|---|---|---|---|---|
| ACaK | Zero Order | 0.198 | 0.928 | |
| | First Order | 0.010 | 0.976 | |
| | Higuchi | 5.266 | 0.984 | |
| | Korsmeyer-Peppas | 3.327 | 0.995 | 0.591 |
| AKCa | Zero Order | 0.048 | 0.698 | |
| | First Order | 0.001 | 0.722 | |
| | Higuchi | 1.341 | 0.838 | |
| | Korsmeyer-Peppas | 7.091 | 0.968 | 0.333 |
| KACa | Zero Order | 0.133 | 0.818 | |
| | First Order | 0.013 | 0.937 | |
| | Higuchi | 3.606 | 0.918 | |
| | Korsmeyer-Peppas | 19.191 | 0.977 | 0.283 |

## 4. CONCLUSION

In this study, the possibility to entrap hydrophobic curcumin was successfully compounded within chitosan-alginate nanoparticles using a very simple ionotropic pre-gelation and polyelectrolyte complex techniques. The diameters of nanoparticles were obtained between 51 and 90 nm. The encapsulation efficiency was up to 85% in higher drug-polymer ratio. The kinetic model of desorption for ACaK, AKCa and KACa nanoparticles followed Korsmeyer-Peppas. These materials are suitable for uptake in capillary due to their nano-sized range and could stay long time into bloodstream.

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UNDER PEER REVIEW

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## APPENDIX