Gliocladium virens,	Original Research Artic COMPATIBILITY OF Streptomyces griseorub and Trichoderma harzianum AGAINTS Fusar OF TOMATO WILT DESEASES.
ABSTRACT	
Streptomyces griseorum Fusarium oxysporum f.s. 2012-2013 at East Ja completely randomized the compatibility of the intagonistic test of bi marzianum capable inta aboratory and at the sinarzianum were compa- circeen house. Clear zo shown that the antibiosis the antibiosis of other protected plant tomato for protection by BCAs mixed	ded to discover the compatibility of biological control agents (BC bens, Gliocladium virens, and Trichoderma harzianum aga sp. capsici in vitro and in vivo. Study was done in rainy sea ava-Indonesia. Study was a true experiment designed, usin design (CRD), consist of three stages research. These stages we hree biological agents in Potato Dextrose Agar (PDA) med iological agents combinations of <u>S. griseorubens, G. virens, and</u> hibited microbial pathogens <i>F. oxysporum</i> f.sp. <i>lycopersici</i> in screen house. Mixed of BCAs <i>S. griseorubens, G. virens, and</i> table and effectively against <i>F. oxysporum</i> in Petri dishes an ona avarage of antibiosis of BCAs filtrat in Potato Glucose ex s from mixed of <i>S. griseorubens</i> and <i>T. harzianum</i> was higher BCAs treatment. Plant infested with mixed of BCAs significat from <i>F. oxysporum</i> compared to the untreated control plants. F ed of <i>T. harzianum</i> with <i>S. griseorubens</i> was more pronounced of <i>S. griseorubens</i> with <i>G. virens</i> and single BCAs.

Fusarium oxysporum f.sp. *lycopersici* (FOL) is a highly destructive pathogen of both greenhouse and field grown tomatoes in warm vegetable production areas. The disease caused by this fungus is characterized by wilted plants, yellowed leaves and minimal or absent crop yield (10,14). Based on our survey in East Java (Malang, Pare, Kediri) there was a 10 to 12 % yield loss.

22 F. oxysporum is soil-borne plant pathogens and the most difficult to control. Chemical control effect negative to environment, there is renewed interest in biological control 23 24 based on application of populations of antagonistic micro-organisms. (2,15). Soil microorganism challenge to be BCAs, the advance of technology these days has come to 25 the application on field, by hoping to be able to make efficient of natural resources, 26 27 conservation and the everlasting environment, also to produce cheaper and healthier 28 agriculture products. Some saprophyte soil microorganism have been used as comercial 29 biological agents, It can be single antagonistic or multi antagonistic (7). The research and 30 the usage of S. griseorubens as biological agent is rarely conducted in agriculture field 31 especially in Indonesia, meanwhile the bacteria, fungus and virus had been through many researches. *T. harzianum* is a soil saprophytic fungus able to become hyperparasitic to several species of fungal pathogen. The growth of *T. harzianum* is very rapid and none 32 33 pathogenic. The hyphae threads of pathogenic fungus will be cut to pieces because it 34 winded by Trichoderma hyphae (as antagonist fungus). Trichoderma eventually release 35

36 antibiotic to phatogenic fungus which is glicotoxin (9,13,23,25). G. virens control plant pathogen by several mechanisms such as parasitism, antibiosis, competition and cell 37 38 destruction. Gliocladium will grow around the pathogen and release enzyme that can destroy 39 chitin of pathogen. G. virens also producing glicotoxin antibiotics (11) S. griseorubens, G. 40 virens and T harzianum as the single antagonist is able to control fusarium wilt disease on 41 tomato and melon in the greenhouse scale (10,22). This research was intended to discover 42 the biological agents S. griseorubens, G. virens, and T. harzianum capable inhibited 43 microbial pathogens F. oxysporum f.sp. lycopersici in the laboratory and in the screen 44 house..

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46 2. Method and Material

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48 **2.1 Isolation of Biological agents**

Isolation of biological agents used soil platting method by Dhingra and Sinclair (21):
1 gram of Pare-Kediri chilli and tomato soil was made suspension by dilution 10⁴.
Subsequently 1 mL of soil suspension was spreaded on Glucose Nutrienth Agar (GNA) to
get *S. griseorubens* and 1 mL of *T. harzianum* and *G. virens* suspension (BPTPH Pandaan)
also spreaded on Potato Dextrose Agar (PDA). Biological agents obtained was purified and
propagated on PDA in Petri dishes.

56 2.2 Isolation of *F. oxysporum* f.sp. lycopersici

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58 Cut into the base of the stem of a diseased plant lengthwise to reveal the xylem just below the epidermis. Turn off all the leaves and secondary roots, leaving only the main stem 59 60 and the hypocotyls and main root. Stem sterilization was done by soaking in 10% bleach solution for 5 minutes. Dry the stem on paper towels. Using sterile technique, cut thin : 2-4 61 mm wedges out of one side of the stem near the root/stem junction making sure to include 62 63 xylem tissue with each wedge. Placed 5-6 wedges on PDA plates. Incubate the plates under fluorescent lights. Once the fungus has grown sufficiently from the pieces, transfer 64 65 isolates onto fresh PDA plates. Incubate the plates for 10-14 days. Colonies of F. 66 oxysporum are pigmented with a reddish purple color and surrounding by a pinkish white 67 aerial mycelium (18).

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2.3 Compatibility test

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Compatibility test was done in Microbiology laboratorium of Agricultural Faculty by watching the type of biological agents growth in discriptive and colony diameter, Every treatment was repeated five times. Colony diameter average were analyze by t-test (12)

Preparing 0.5 cm colony diameter of 10th day biological agents, *S. griseorubens T. harzianum, G. virens*. Compatibility tests on the PDA media plating in 20 diameter cm Petri dishes were three types of biological agents colony *S. griseorubens., G. virens.,* and *T. harzianum* (SGT) were placed on PDA medium, each with the same distance of 5 cm, and were incubated for 14 days. Along with the treatment of compatibility, also it was prepared control treatment as a comparison.

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2.3 Antagonistic in vitro test

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83 Antagonistic *in vitro* test was done by completely randomized design (CRD) with 84 seven treatment and every treatment was repeated three times. Data of percentage 85 inhibition was analyzed by Duncan test

86 suspension of biological agents treatment was made : 6 mL was mixed up in 44 mL
 87 of sterile water, 0.33 cc of this each suspension biological agents treatment was taken into
 88 holes (wells). Then put the 0.5 cm colony of *F. oxysporum* 7 days age, in the presence of

biological agents suspension that has been inoculated in the hole at a distance of 5 cm.
Each treatment was randomly stored at room temperature for 8 days. The inhibition was
calculated by the formula (8):

DI is the percentage inhibition;

Dc is the colony diameter of the control treatment; Dt is the colony diameter of BCAs treatment.

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Antibiosis test consist of 7 treatment placed in randomly, every treatment was done in triplicate.

104 Six mili liter of BCAs (every BCA 38-42 spore/cc) treatment entered into 44 mL 105 Potato Glucose Extract solid medium in Erlenmeyer flasks. BCAs suspension was shaken. 106 Inhibition ability of antibiosis filtrate of BCAs againts *F. oxysporum* was done by Steinkelner 107 method. 0.55 cc antibiosis was given into 0.5 cm Whatman paper disc, then air drying 108 them. These paper disc was inoculated on PDA medium in Petri dish contains *F. oxysporum* 109 suspension. Inhibition ability of antibiosis was done by counting of clear zone diameter (27).

110 111 **2.5 Antagonism** *in vivo* test

Dc - Dt

DI = ----- x 100

Dt

112 This research was done by Completely Randomized Design (CRD) with seven 113 treatmen and every treatment was done in triplicate. Data of percentage inhibition was 114 analyzed by Duncan's t-test.

115 Seedling was inoculated by soaking a solution of inoculum combination of biological agents that have been prepared before. Furthermore, the seed was planted in the soil that 116 had been inoculated *F. oxysporum* suspensiton (10⁹ spore/mL) for 14 days. It was prepared 117 by filled 3 liters of sterile soil in polybags and inoculated with spore inoculum mass 118 suspension of *F. oxysporum* has been sprayed with a hand sprayer at ground level (21). 119 Stored for 14 days in the screen house and watering with sterile water every day. Data were 120 collected for : (1) Phase of incubation, performed daily until symptoms of disease were 121 vellowing of the leaves from the bottom, where the control healthy plants not showing 122 123 symptoms. (2) Disease severity, was conducted with most of the leaves are yellow, wilt and 124 dry every 7 days until harvest (3).

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KP =	count of yellowing leaf per plant count of all leaf of plant	126 x 1 020% 128
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2.4 Antibiosis in Potato Glucose Extract medium

KP is the percentage of disease severity per plant (3)

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132 3. RESULTS AND DISCUSSION

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3.1 Compatibility test

Performance of three biological agents *S. griseorubens* (S) *G. virens* (G) and *T. harzianum* (T) showed none antagonistic, grew thicker than single biological agent (Fig. 1)

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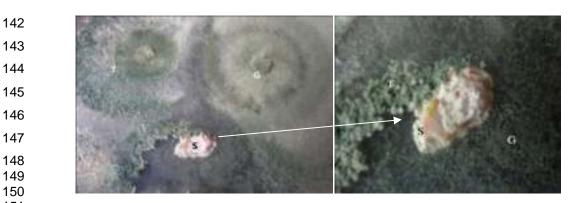
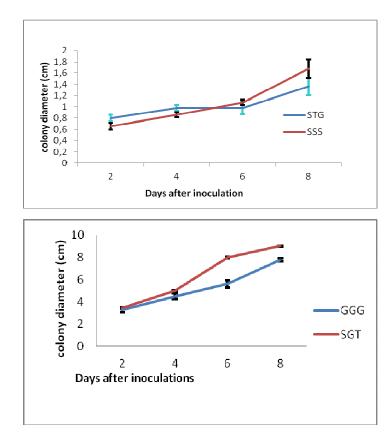
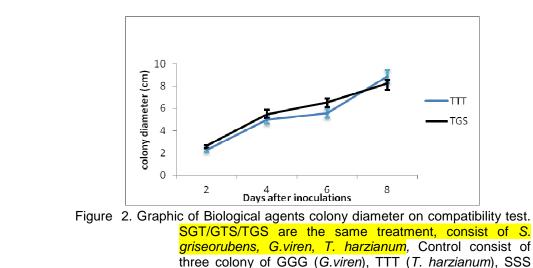


Figure 1. T, harzianum colony(T), S. griseorubens colony (S), G. virens colony (G) on PDA medium, 14th day after inoculations BCAs.

Growth average of colony diameter S. griseorubens (S) G. virens (G) and T. harzianum (T) in compatibility test (SGT) on 2^{nd} day after inoculation showed none significantly different when compared with controls (SSS, GGG, TTT). However, on 4^{th} and 6th day after inoculations, diameter growth of biological agents such colony in compatibility testing (SGT) was larger and significantly different than the control. On 8th day, the growth of biological agents on a compatibility test was not different from the control of biological agents (Fig.2)





(S. griseorubens.)

S. griseorubens, G. virens and T. harzianum as biological agents could grow on PDA media and formed an association that does not harm each other or not produced secondary metabolites that could inhibit the growth of biological agents each other. The three biological agents are saprophyte soil microbes, and produce antibiosis to mycoparasite only affect microbial pathogens. This opinion is supported by the results of some other research, plant pathogenic fungi cell walls composed of chitinase which is a key enzyme and responsible for the lysis of the cell wall. S. griseus degrade fungal cell walls by lytic enzymes (8). Trichoderma sp. produced lytic enzyme that degraded chitin, interfungus parasitism and can improve the cell wall itself on the division process (16). *Gliocladium* sp. and Trichoderma sp. produce chitinase enzymes that can cause parasitic on plant pathogens, whereas less effective antibiosis produced degrades F. oxysporum (11). The existence of microbes of different biological agents also induce the microbes to grow faster. As noted by some researchers that Trichoderma sp. and Gliocladium sp. produce fungal pathogen that work in synergy with the intracellular enzyme produced by G.virens. Both of these biological agents was synergy in controlling pathogenic tomatoes until 57% (23, 24).

227 3.2 Antagonism compatibility in vitro

Giving a single biological agents *S. griseorubens* (*S*), *G.virens* (*G*), *T. harzianum* (*T*), a mix of two biological agents (SG, ST, GT) and a mix of three biological agents (SGT) to tomato seed, significantly inhibited the development of the colony diameter of *F. oxysporum.* Giving only *G.virens* to tomato seed shown small and significantly diffrent average inhibition than the other single of biological agents *S. griseorubens* and *T. harzianum* and the combination treatment (Table 1).

Tabel 1. Inhibitors average of biological agents S. griseorubens (S), G. virens (G), T. harzianum (T) against F. oxysporum

Treatment of BCAs	Inhibitor average ± SE(%)
S. griseorubens, T. harzianum (ST)	<mark>63.97 ± 7.27</mark> ^a
G. virens, T. harzianum (GT)	<mark>59.29 ± 3.87 ^{ab}</mark>
S. griseorubens, G. virens, T. harzianum (SGT)	59.07 ± 1.82 ^{ab}
S. griseorubens, G. virens (SG)	58.41 ± 1.93 ^{ab}
T. harzianum (T)	49.05 ± 3.41 ^{ab}
S. griseorubens (S)	45.11 ± 5.37 ^b
G. virens (G)	40.24 ± 7.27 [°]

246 247 Explanation : The same letters which added behind the numbers indicate none significant difference in the Duncan's t-test (p < 0.05).

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249 Single biological agent *G. virens* (G), have ability to inhibit microbial pathogens *F* 250 oxysporum (F) lower than the inhibition ability of single biological agents T. harzianum, S. 251 griseorubens (T.S), and mix of biological agents (TS, GS, TG and TGS). G.virens serves 252 only as competitors and parasitic. They did not produce antibiosis on PDA media (in vitro) 253 (11,15,21). Antibiosis of biological agents G.virens, T. harzianum and S. griseorubens on Potato Glucose Extract (PGE) also proved that G. virens did not produce antibiosis on 254 observations 2nd,4th,6th day after the inoculation of biological agents. Single BCA T. 255 harzianum and S. griseorubens and combination of BCAs on observation produced 256 antibiosis (Table 2) 257

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Table 2. Avarage of Inhibition zone of biological agents crude extract to *F. oxysporum*

No	Biological agents giving	Clear zone (mm)			
		2 dai	4 dai	6 dai	
1	T. harzianum	0.2	0.2	0.3	
2	G. virens	0	0	0	
3	S. griseorubens	1.3	0.4	0	
4	G. virens, T. harzianum.	0	0	0	
5	T. harzianum, S. griseorubens	1.2	0.7	0.2	
6	G. virens. S. griseorubens	0.3	0.3	0	
7	<i>G. virens , S. griseorubens</i> and <i>T. harzianuim</i>	0	0.4	0.2	

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Notes : Dai is day after inoculations

262 263 This suggests that a mix of all three biological agents produced antibiosis derived 264 from T. harzianum and S. griseorubens f.sp. capsicum. Single biological agents Τ. 265 *harzianum* (T) and a mix of biological agents (TG, TS, GS and SGT) can result inhibition ability higher than inhibition ability of single biological agents S. griseorubens (S) and 266 G.virens (G). Several studies have shown that the fungus Trichoderma sp. is a saprophyte 267 268 fungus that lives in the soil and becomes hyper parasite on some pathogenic fungi. T. 269 harzianum also inhibit the growth of F. oxysporum colonies growing very rapidly, and 270 producing antifungal namely Glicotoxin. Gliocladium sp., Streptomyces sp. Trichoderma sp. clasified as soil saprophyte fungi and used as biological agents have multi antagonistic 271 272 mechanism and compatible againts *F. oxysporum* (27).

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274 3.4 Antagonistic compatibility *in vivo*

275 Tomato plants that were not given biological agents showed fusarium wilt symptoms 39-40th days after planting tomato seedlings. Meanwhile, the plants treated with 276 on biological agents, appeared symptoms at 46-50th days after planting. Longer incubation 277 278 occured because of biological agents mixture could inhibit the development of F. oxysporum. Giving of biological agents with the pathogen in the soil for one week, the incubation period 279 occurred after 45 days (20). Single biological agents S. griseorubens (S) and a mix of 280 biological agents, S. griseorubens with G.virens (SG), S. griseorubens, with T. harzianum 281 282 (ST), and a mixed of S. griseorubens, G.virens, T. harzianum (SGT) can inhibit disease 283 severity (Table 3).

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Avarage of wilt desease severity of tomato plant at 41th, 48th, 55th and 62th day Table 3 after planting (dap).

No	Giving of biological agents	Avarage of deseases severity (%)			
NO		41 dap	48 dap	56 dap	62 dap
1	Control (withaot bcas) (K)	<mark>3.80±2.19</mark> ^ª	<mark>7.60±0</mark> ^ª	17.86±1.57 [°]	44.64±3.30 ^a
2	S. griseorubens, T. harzianum. (ST)	0.00±0 ^ª	5.70± 1.9 ^ª	7.14± 5.91 ^a	19.65±3.41
3	S. griseorubens (S)	<mark>0.00±0</mark> ª	<mark>1.90±1.9</mark> ^a	<mark>15.42±2.90.</mark> ^a	<mark>16.07±7.46</mark> [▶]
4	S. griseorubens, G. virens (SG)	<mark>1.90± 1.9 ª</mark>	<mark>3.80±2.19</mark> ^ª	7.15± 4.13 [°]	<mark>16.07±7.36</mark> [▶]
5.	S. griseorubens, G. virens,T. harzianum (SGT)	<mark>1.52±1.9</mark> ^ª	<mark>5.77±5.77</mark> ^ª	<mark>5.77±3.77</mark> [°]	14.29±5.63 ^b

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Explanation: The same alphabet beside the number in the same coloum shown insignificant Duncan test (p < 0.05)

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Each of these biological agents inhibit the development of pathogens with multiple 291 mechanisms and can develop optimally in soils containing organic matter, so that they can 292 be complement each other. Based on several studies, the Trichoderma sp., Streptomyces 293 sp., and *Gliocladium* sp., were antagonistic to fungal and bacterial diseases of plant roots (3, 294 27). Three biological agents can flourish together on compost, manure and garden soil. 295 In synthetic medium, S. griseorubens did not develop optimal, but developed optimal in 296 field conditions. The study also found that for 4 weeks, the average population of biological 297 agents S. griseorubens more higher than the avarage populations of biological agents T. 298 harzianum (25).

On 55th and 62 day after planting, combination of biological agents 299 300 T. harzianum and S. griseorubens demonstrated lower power resistor ability of severity disease than power resistor ability of other biological agents combinations. Several studies 301 in the screenhouse and in the field proved that inhibit ability of T. harzianum to pathogens 302 was lower than inhibit ability of G.virens in soil. Otherwise, Gliocladium virens grew very 303 304 fast and produced antibiotics gliovirin. Antibiotics worked in synergy with intracellular 305 enzymes to inhibit the development of fungal plant pathogens (3,4). Providing a mixed of 306 biological agents T. harzianum (T), G.virens (G), S. griseorubens (S) can also enhance plant 307 growth and provide organic material for plants as planting soil decomposition. Biological 308 agents can also induce the growth of pheriphere new roots more and replaces the root of suffering discoloration, can repair the affected plant roots by *F. oxysporum* also(17,26). 309

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311 4. Conclusion

312 S. griseorubens, G. virens and T. harzianum as biological agents were compatible grow on PDA media and formed an association that does not harm 313 314 each other or not produced secondary metabolites that could inhibit the growth of biological agents each other. A single biological agents S. griseorubens (S), T. 315 harzianum (T), a mix of two biological agents (SG, ST, GT) and a mixed of three 316 317 biological agents (SGT) more inhibited the development of the colony diameter of 318 F. oxysporum than a single biological agent G. virens in vivo. Giving mix of two 319 biological agents S. griseorubens and G. virens as well as S. griseorubens and T. harzianum as well as three biological agents S. griseorubens, G. virens and 320 inhibit disease severity of tomato fusarium wilt caused by 321 T. harzianum to 322 T. oxysporum f.sp. lycopersici

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324 CONSENT

325 All author declare that written informed consent was obtain from the approved of our 326 research parties

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329 REFERENCES

- 331 1. Abeysinghe, S. 2007. Biological control of Fusarium solani f.spp. phaseoli the causal 332 agents of root rot of bean using Bacillus subtilis CA 32 and Trichoderma 333 harzianum RU01. Ruhuna Journal of Science. 2: 62-88.
- 334 2. Alabouvette, C., C. Olivain, Q. Migheli, C. Steinberg. 2009. Microbiological control of 335 soil-borne phytopathogenic fungi with special emphasis on wilt-336 oxysporum. On Line Journal DOI: 10.1111/j.1469 inducing Fusarium -8137.2009.03014.x 337
- 338 3. Anitha, A. and M. Rabeeth. 2010a. Control of Fusarium wilt tomato by bioformulation of 339 Streptomyces griseus in green house condition. African Journal of Basic & 340 Aplied Sciences 1 (1-2): 9 – 14.
- 4. Anitha, A. and M. Rabeeth, 2010 ^b. Degradation of fungal cell walls of phytopatogenic 341 fungi by lytic enzyme of Streptomyces griseus. African Journal of Plant 342 343 Science . 4 (3): 061-066.

344 345 346	5. Asha, B., C. Nayaka, U. Shankar, Srinivas, Nirjana. 2011. Biological control of <i>F. oxysporum</i> f. sp. <i>lycopersici</i> causing wilt of tomato by <i>Pseudomonas</i> <i>fluorescens. International Journal of Microbiology Research.</i> 3(2): 79-84
347 348	6. Bollen, G.J. 1974. Fungal recolonization of heat-treated glasshouse soils. Agro Ecosystems I: 139-155
349 350 351	7. Cook J. R. and K. F. Baker. 1996. The nature and practice of biological control of plant pathogens. APS PRESS. <i>The American Phytopathological Society</i> . St. Paul, Minnesota.
352 353 354	8. Fahri, Y. and M. Dikilita. 2007. Control of fusarium wilt of tomato by combination of <i>Pseudomonas florescent</i> , non <i>patogen Fusarium</i> and <i>Trichoderma</i> <i>harzianum</i> T-22 in greenhouse conditions. <i>Plant Pathology Journal</i> 6(2) :
355	159-163.
356 357	9. Gruber, S. and V. Seiboth. 2012. Self versus non-self: fungal cell wall degradation In <i>Trichoderma. Microbiology</i> 158:26-34.
358 359	10.Kaewchai, S., Soytong, K., and Hyde, K.D. 2009. Mycofungicides and fungal biofertilizers. <i>Fungal Diversity</i> . 38: 25-50.
360 361 362	11. Kyeong, S. J., Hong, M. K., and Bong, K. C., 2000. Purification and antifungal activities of an antibiotic produced by <i>Gliocladium virens</i> G1 against plant patogen. Plant <i>Patholohy Journal</i> , J.17(1) : 53-56.
363 364	 Kusriningrum, R.S. 2008. <i>Perancangan Percobaan</i>. Airlangga University Press. Larkin, R.P. and D.R. Fravel. 1993. Biocontrol of Fusarium Wilt of tomato. Biocontrol of
365 366	Plant Deseases Laboratory. Bestvile 14. Menzies JG, Koch C, Seywerd F. 1990. Additions to the host range of <i>Fusarium</i>
367	oxysporumf. sp. radicis-lycopersici. Plant Diseases. 74: 569–572.
368 369 370	15. Morid, B., S. Hajmansoor, N. Kakvan, 2012. Screening of resistance genes to fusarium root rot and fusarium wilt diseases in tomato (<i>Lycopersicon esculentum</i>) cultivars using RAPD and CAPs markers. <i>European Journal of</i>
371 372	<i>Experimental Biology</i> . 2 (4):931-939 16. Nourozian J., H. R. Etebarian, and G. Khodakaramian, 2006. Biological control of
373 374	Fusarium grameniarum on wheat by antagonistic bacteria. Songklanakarin Journal, Sci Technol 28 :29-38.
375 376 377 378	17a. Olivain C., C. Humbert, J. Nahalkova, J. Fatehi, FL. Haridon, and C. Alobouvete. 2006. Colonitation of tomato root by phatogenic and non patogenic <i>Fusarium</i> <i>oxysporums</i> strains inoculated together and separately into the soil. <i>Aplaid and Enviromental Microbiology.</i> 72 (2): 1523-1531.
379	18. Reis A, Costa H, Boiteux LS, Lopes CA. 2005. First Report of Fusarium oxysporum f. sp.
380 381 382	<i>lycopersici</i> Race 3 on Tomato in Brazil. <i>Fitopatology</i> . Bras. 30(4): 426-428. 19. Siddiquee, Shafiquzzaman, Soon G. T., and Y. U. Kalsum. 2010. Isozyme Analisis and Relationships, Among Three, Species in Malaysia, <i>Triphadarma</i> , Isolatas
382 383	Relationships Among Three Species in Malaysia <i>Trichoderma</i> Isolates. <i>Mycrobial Biotechnol.</i> 20(9): 1266 – 1275
384 385 386	20. Singh R.,B.K. Singh, R.S. Upadhyay, B. Rai and Y. S. Lee. 2002 Biological control of fusarium wilt disease of pigeonpea. <i>Plant Pathology Journal</i> 18(3) : 279-283.
387 388 389	21. Singleton, J.D. Mihail and C.M. Rush. 1993. Methods for research on soilborne phytopatogenik fungi . APS Press. <i>The American Phytophatological Society</i> . St. Paul Minesota.
390 391 392	22. Staniazsek M, Kozik EU, Marczewski W . 2007. A CAPS marker TAO1902 diagnostic for the I-2 gene conferring resistance to <i>Fusarium oxysporum</i> f. sp. <i>lycopersici</i> race 2 in tomato. <i>Plant Breeding</i> . 126(3): 331-333.
393	23. Stipanovic, R.D. and C.R. Howell. 1982. The Structure of Gliovirin, A new Antibiotic
394 395 396	from <i>Gliocladium virens. The Journal of Antibioticsicotoxin.</i> 24. Suharjono, Tri Handayani, Soejono, Susanti Dewi. 2008. Antagonis test of <i>Trichoderma</i> sp. dan <i>Gliocladium</i> sp. againts <i>Fusarium oxysporum</i> cause of wilt

- 397 deseases of some variety of Purwodadi field banana in Vitro (In 398 Mathemathic and Scient Faculty, Unibraw Indinesian). Biologi Study, Malang, 399 400 25. Suryaminarsih dan Mujoko. 2012. Growth population of multiantagonis Streptomyces 401 Gliocladium sp and Trichoderma harzianum as biological agents of sp. 402 fusarium wilt disease in natural and semi natural package pellet formula 403 (In Indonesian). Plumula, 1 (2): 202-210. 26 Suryaminarsih, Kusriningrum, Ni'matuzahroh, Surtiningsih, 2014. Plant Resistance with 404 405 pheriphere new roots by BCAs Gliocladium sp and T. harzianum againts F. 406 oxysporum on sprout of tomato. Prossiding of Plant Protection national 407 Seminar (In Indonesian). 408 27. Steinkellner S., R. Mammerder, and H. Vierhellig. 2008. Germination of Fusarium 409 oxysporums in root exudates from tomato plants callenged with diffrent Fusarium oxysporums strains. Plant pathology. 122: 395-401 410
- 411 28. Titus, A. and G.N. Pereira. 2008. The role of Actinomycetes in coffee plantation 412 ecology. *Ineedcoffee.com*