Original Research Article

ANTAGONISTIC COMPATIBILITY OF Streptomyces griseorubens, Gliocladium virens, and Trichoderma harzianum AGAINTS Fusarium oxysporum CAUSE OF TOMATO WILT DESEASES.

6 7

1

2 3

4

5

9 10

11 ABSTRACT

12

This research was intended to discover the compatibility of biological control agents (BCAs) Strepromyces griseorubens, Gliocladium virens, and Trichoderma harzianum againts Fusarium oxysporum f.sp. capsici in vitro and in vivo. Study was done in rainy season 2012-2013 at east java-Indonesia. Study was a true experiment designed, using a completely randomized design (CRD), consist of tree stages research. First stage of study was the compatibility of the three biological agents in sintetis medium (PDA). In the next research phase, were antagonistic test of biological agents combinations S. griseorubens, Gliocladium virens, and Trichoderma harzianum capable inhibited microbial pathogens F. oxysporum f.sp. lycopersici in the laboratory and in the screenhouse. Mixture of BCAs S. griseorubens, G. virens, and T. harzianum were compatibel and effectively against F. oxysporum on Petri dishes and at screenhouse. Clear zona avarage of antibiosis of BCAs filtrat in Potato Glocose extrac shown the antibiosis of S. griseorubens and T. harzianum mixtured higher than the antibiosis of other BCAs treatment. Morewere plant infested with mixture of BCAs significantly protected plant tomato from F. oxysporum compared to the untreated control plants. Plant protection by BCAs mixture of T. harzianum with S. *griseorubens* was more pronounced than plant protection by mixture of *S. griseorubens* with G. virens and single BCAs. Soil filtrate that was inoculation with mixtured of biological agents G. virens (G), T. harzianum (T), and S. griseurubens (S), resulted antibiosis which could inhibit the growth of fungal pathogens F. oxysporum.

- 13
- 14

Keywords: Wilt deseases, Antagonitic, mix of Biological Agents (BCAs)

15 16

17 Fusarium oxysporum f.sp. lycopersici (FOL) is a highly destructive pathogen of both 18 greenhouse and field grown tomatoes in warm vegetable production areas. The disease 19 caused by this fungus is characterized by wilted plants, yellowed leaves and minimal or absent crop yield. There may be a 30 to 40% yield loss (5). Fungal phytopathogens are 20 21 cause of many plant diseases and much loss of crop yields, especially in tropical and 22 subtropical regions. Fusarium oxysporum is major soilborne fungal pathogens of both 23 greenhouse and field grown tomatoes in the warm vegetable growing areas of the world 24 (14).

25 Plant diseases induced by soil-borne plant pathogens are among the most difficult to 26 control. In the absence of effective chemical control methods, there is renewed interest in 27 biological control based on application of populations of antagonistic micro-organisms. (2). 28 Although lots of challenge occurs, the advance of soil microorganism technology these days 29 has come to the application on field, by hoping to be able to make efficient of natural 30 resources, conservation and the everlasting environment, also to produce cheaper and 31 healthier agriculture products (7). The research and the usage of Actinomycetes as an 32 biological agent is rarely conducted in agriculture field especially in Indonesia, meanwhile

the bacteria, fungus and virus had been through many researches. Trichoderma 33 is a saprofyt fungus that lives in the ground and able Decome hyperparasite to several 34 species of pathogen fungus. The growth of *Trichoderma* **sp** is very rapid and not becoming 35 36 disease for high level plants. The hypha threads of pathogenic fungus will be cut to pieces 37 because it winded by Trichoderma hypa (as antagonist fungus). Trichoderma eventually 38 release deadly antibiotics to harmful fungus which is Glicotoxin (9). G. virens control plant pathogen by several mechanisms such as parasitism, antibiosis, competition and cell 39 40 destruction. Gliocladium will grow around the pathogen and release enzyme that can destroy 41 pathogen cuticle. G. virens also producing glicotoxin antibiotics (11) S. griseorubens, G. virens and Trichoderma sp. as the single antagonist is able to control fusarium wilt disease 42 43 on tomato and melon in the greenhouse scale (10,20). This Research was intended to 44 discover the biological agents S. griseorubens f.sp, capsicum, Gliocladium virens, and Trichoderma harzianum capable inhibited microbial pathogens 45 F. oxysporum f.sp. 46 lycopersici in the laboratory and in the screenhouse. in vitro and in vivo.

⁴⁷ Isolation of biological agents used soil plating method by Dhingra and Sinclair (19): 1 gram ⁴⁸ of chilli and tomato soil weigh with an analytical balance, then made the suspension by ⁴⁹ dilution 10^4 . Subsequently 1 mL of *Streptomyces griseorubens* suspension was spreaded ⁵⁰ on Glucose Nutrienth Agar (GNA) and 1 mL of *T. harzianum* and *G. virens* suspension ⁵¹ (BPTPH Pandaan) also spreaded on Potato Dextrose Agar (PDA). Biological agents ⁵² obtained was purified and propagated on PDA in Petri dishes.

53 54

2.1 Isolation of *F. oxysporum* f.sp. lycopersici

55 Cut into the base of the stem of a diseased plant lengthwise to reveal the xylem just below 56 the epidermis. Trim off all the leaves and secondary roots leaving only the main stem and 57 the hypocotyls and main root. Surface sterilize the stem by soaking in 10% bleach solution 58 for 5 minutes. Dry the stem on paper towels. Using sterile technique, cut thin (2-4 mm thick) 59 wedges out of one side of the stem near the root/stem junction making sure to include xylem 60 tissue with each wedge. Place 5-6 wedges on PDA plates. Incubate the plates under 61 fluorescent lights. Once the fungus has grown sufficiently from the pieces, transfer isolates 62 onto fresh PDA plates. Incubate the plates for 10-14 days. Colonies of F. oxysporum are 63 pigmented with a reddish purple color and surmounted by a pinkish white aerial mycelium 64 (16).

65

66 2.2 Compatibility test67

68 Preparing 0,5 cm diameter colony of 10th days biological agents, 69 Strepton, s griseorubens T. harzianum, G. virens. Compatibility tests on the PDA media 70 plating in 20 diameter cm petri dishes were three types of biological agents colony S. 71 griseorubens., G. virens., a r derma harzianum (SGT) were placed on PDA medium, 72 each with the same distance of 5 cm, and were incubated for 14 days. Along with the 73 treatment of compatibility, also it was prepared control treatment as a comparison.

74 75

2.3 Antagonism compatibility in vitro test

76

85

6 mL suspension biological agents treatment was mixed up in 44 mL of sterile water 0.33 cc of this each suspension biological agents treatment was taken into holes (wells) wich have been made before. Then put the 0.5 cm colony of *F. oxysporum* 7 days age, in the presence of biological agents suspension that has been inoculated in the hole at a distance of 5 cm. Each treatment was randomly stored at room temperature for 8 days. The inhibition was calculated by the formula (8):

- 83 Dc Dt 84 DI = ----- x 100
 - Dt

DI is the percentage inhibition; Dc is the diameter of the control; Dc is the diameter of inhibition.

EER REVIEW HC.

86 87

88

89

2.4 Antibiosis in Potato Glucose Extrac medium

90 6 mL of BCAs treatment entered into 44 mL Potato Glucose Extrac solid medium in 91 Erlenmeyer tubes. BCAs suspension was shaken. Inhibition ability of antibiosis of BCAs filtrat againts *F. oxysporum* was done by Steinkelner method. 0,55 cc antibiosis was given 92 into 0,5 cm Whatman paper disc, then air drying them. These paper disc was inoculated on 93 94 PDA medium in Petri dish contains F. oxysporum suspension . Inhibition ability of antibiosis 95 was done by coump of clear zone diametre (24). 96

97 2.5 Antagonism in vivo test

98

99 Seedling was inoculated by soaking a solution of inoculum combination of biological 100 agents that have been prepared before. Furthermore, the seed was planted in the soil that 101 had been inoculated F. oxysporum for 14 days. It was prepared by filled 3 liters of sterile soil 102 in polybags and inoculated with spore inoculum mass suspension of F. oxysporum has 103 been sprayed with a hand sprayer at ground level (19). Stored for 14 days in the screen house and watering with sterile water every day. Data were collected for: (1) The period of 104 105 incubation, performed daily until symptoms of disease were yellowing of the leaves from the 106 bottom, where the control healthy plants not showing symptoms. (2) The severity of the 107 disease, was conducted with most of the leaves are vellow, wilt and dry every 7 days until harvest (3). 108

109

110 count of yellowing leaf KP =x 100% count of all leaf of plant 112

KP is the percentage of disease severity(3)3. RESULTS AND DISCUSSION

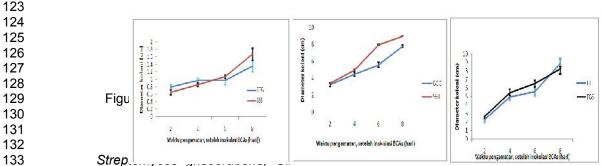
3.1 Compatibility test 114

115

116 117

Growth average of diameter colony S. griseorubens (S) G.virens (G)and Τ. harzianum (T) in compatibility test (SGT) on 2 nd days after inoculation showed no significantly different when compared with controls (SSS, GGG, TTT). However, on 4 th and 118 ¹days after inoculations, diameter growth of biological agents such colony in compatibility 119 testing (SGT) was larger and significantly different than the control. On 8 th days, the growth 120 121 of biological agents on a compatibility test was not different from the control of biological 122 agents (Fig.1)

113



biological agents could grow together on PDA and formed an association that does not harm 134 135 each other or not produced secondary metabolites that could inhibit the growth of biological 136 agents each other. The three biological agents is a saprophyte soil microbes, and produce 137 antibiosis mycoparasite only affect microbial pathogens. This opinion is supported by the 138 results of research, plant pathogenic fungi cell walls composed of chitinase which is a key

139 enzyme and responsible for the lysis of the cell wall. Streptomyces griseus degrade fungal 140 cell walls by lytic enzymes (8). Trichoderma sp. produced lytic enzyme that degraded chitin, 141 mycoparasit process and can improve the cell wall itself on the division process (15). 142 Gliocladium sp. and Trichoderma sp. produce chitinase enzymes that can cause parasitic on 143 plant pathogens, whereas less effective antibiosis produced degrades *F. oxysporum* (11). 144 The existence of microbes of different biological agents also induce the microbes to grow 145 faster. As noted by some researchers that Trichoderma sp. and Gliocladium sp. produce 146 fungal pathogen that work in synergy with the intracellular enzyme produced by G.virens. 147 Both of these biological agents was synergy in controlling pathogenic tomatoes until 57% 148 (21, 22).

149 3.2 Antagonism compatibility in unit 150

151 Giving a single biological agents S. griseorubens (S), G.virens (G), T. harzianum 152 (T), a mixture of two biological agents (SG, ST, GT) and a mixture of three biological agents 153 (SGT) to tomato seed, significantly inhibited the development of the colony diametre of F. 154 oxysporum. Giving only G.virens to tomato seed shown smaaler and significantly diffrent 155 average inhibition than the other single of biological agents S. griseorubens and T. 156 harzianum and the combination treatment (Table 1).

157

158 159

Tabel 1. Inhibitors average of biological agents S. griseorubens (S), G. virens (G) dan T. harzianum (T) againts F. oxysporum

Treatment of BCAs	Inhibitor average (%)
S. griseorubens, T. harzianum (ST)	63,97 ^a
G. virens, T. harzianum (GT)	59,29 ^{ab}
S. griseorubens, G. virens, T. harzianum (SGT)	59,07 ^{ab}
S. griseorubens, G. virens (SG)	58 ,41 ^{ab}
T. harzianum (T)	49,05 ^{ab}
S. griseorubens (S)	45,11 ^b
G. virens (G)	40,24 ^c

160

Explanation : The same letters in addition to numbers showed no significant difference in the Duncan level test.

161 162

163 Single biological agents S. griseurubens (S), G. virens (G), and T. harzianum have 164 Inhibition ability of microbial pathogens F. oxysporum (F) lower than the inhibition ability of 165 biological agents combination (TS, GS, TG and TGS). This is because G.virens serves only 166 as competitors and parasites, and did not produce antibiosis on PDA (11, 15, 21). Antibiosis 167 biological agents G.virens, T. harzianum and S. griseorubens f.sp. capsicum on Potato 168 Glucose Extrac (PGE) also proved that G.virens not generate antibiosis on observations 169 2nd,4th,6th days after inoculation biological agents (Table 2). 170

171

Table 2. Avarage of Inhibitor zone of biological agents crude extrac to F. oxysporum

PEER REVIEW h'R

No	Biological agents giving	Clear zone (halo)mm		
		2 dai	4 dai	6 dai
1	T. harzianum	0,2	0,2	0,3
2	G. virens	0	0	0
3	S. griseorubens	1,3	0,4	0
4	G. virens d . harzianum.	0	0	0
5	T. harzianum d. D. griseorubens	1,2	0,7	0,2
6	G. virens dan S. griseorubens	0,3	0,3	0
7	<i>G. virens , S. griseorubens</i> and <i>T. harzianuim</i>	0	0,4	0,2

172 173

174 This suggests that a mixture of all three biological agents still produced antibiosis 175 derived from T. harzianum and S. griseorubens f.sp. capsicum. Single biological agents 176 T. harzianum (T) and a mixture of biological agents (TG, TS, GS and SGT) can result 177 inhibition ability higher than inhibition ability of single biological agents S. griseorubens (S) 178 and G.virens (G). Several studies have shown that the fungus Trichoderma sp. is a saprophyte fungi that lives in the soil and becomes hyperparasite on some pathogenic fungi. 179 180 T. harzianum also inhibit the growth of F. oxysporum colonies growing very rapidly, and 181 producing antifungal namely Glicotoxin. *Gliocladium* sp., *Streptomyces* sp. *Trichoderma* sp. clasified as soil saprophyte fungi used as biological agents and active ingredients of fertilizer 182 183 biocompos (24).

184

185

3.4 Antagonistic compatibility in Green house (in vivo) 186

Tomato plants that were not given biological agents showed fusarium wilt symptoms 187 39-40th days after planting tomato seedlings. Meanwhile, the plants treated with 188 biological agents, appeared symptoms at 46-50th days after planting. Longer incubation occured because of biological agents mixture could inhibit the development of *F. oxysporum*. 189 190 Giving of biological agents with the pathogen in the soil for one week, the incubation period 191 192 occurred after 45 days (18). Single biological agents S. griseorubens (S) and a mixture 193 of biological agents, S. griseorubens with G.virens (SG), S. griseorubens, with T. harzianum 194 (ST), and a mixture of S. griseorubens, G.virens, T. harzianum (SGT) can inhibit disease 195 severity (Table 3). 196

197

Avarage of wilt deseases severity of tomato plant at 41th, 48th, 55th and 62th days Table 3 after planting (dap).

No	Giving of biological agents	Avarage of deseases severity (%)			
		41 dap	48 dap	56 dap	62 dap
1	Control (withaot bcas) (k)	3,80 ^a	7,60 ^a	17,86 ^a	44,64 ^a
2	S. griseorubens, T. harzianum. (st)	0,00 ^a	5,70 ^a	7,14 ^a	19,65 ^b
3	S. griseorubens (s)	0,00 ^a	1,90 ^a	15,42 ^a	16,07 ^b
4	S. griseorubens, G. virens (sg)	1,90 ^a	3,80 ^a	7,15 ^a	16,07 ^b
5.	S. griseorubens, G. virens, T. harzianum (sgt)	1,52 ^a	5,77 ^a	5,77 ^a	14,29 ^b

198 199 Explanation: The same alphabet beside the number in the same coloum shown insignificant Duncan test (p < 0.05)

200

201 Each of these biological agents inhibit the development of pathogens with multiple 202 mechanisms and can develop optimally in soils containing organic matter, so that they can 203 be complement each other. Based on several studies, the Trichoderma sp., Streptomyces 204 sp., and Gliocladium sp., were antagonistic to fungal and bacterial diseases of plant roots 205 (3,18,24). This three biological agents can flourish together on compost, manure and garden 206 soil . In synthetic medium, S. griseorubens did not develop optimal. but developed optimal in field conditions. The study also found that for 4 weeks, the average population of biological 207 208 agents S. griseorubens more higher than the avarage populations of biological agents T. 209 harzianum (23).

On 55th and 62th days after planting, combination of biological agents 210 T. harzianum and S. griseorubens f.sp. capsicum demonstrated lower power resistor ability 211 of (high disease severity) than power resistor ability of other biological agents combinations. 212 Several studies in the screenhouse and in the field proved that T. harzianum inhibit ability in 213 214 soil borne pathogens was lower than G.virens inhibit ability. Otherwise, Gliocladium virens 215 grew very fast and produced antibiotics gliovirin. Antibiotics worked in synergy with intracellular enzymes to inhibit the development of fungal plant pathogens (3,4). Providing a 216 217 mixture of biological agents T. harzianum (T), G.virens (G), S. griseorubens f.sp. capsicum 218 (S) can also enhance plant growth and provide organic material for plants as planting soil 219 decomposition Giving biological agents can also grow new roots more and replaces the root 220 of suffering discoloration and can repair the affected plant roots by F. oxysporum (17).

221 222

223 4. CONCLUSION

224

Biological agents *S. griseorubens*, *G.virens* and *T. harzianum* were compatible microbes to inhibit the development of *F. oxysporum* in vitro conditions. Mixture of two biological agents *S. griseorubens* and *G.virens* as well as *S. griseorubens* and *T. harzianum* as well as three biological agents *S. griseorubens f.sp. capsicum* f.sp. *capsicum*, *G.virens* and *T. harzianum* in inhibit ability of disease severity of tomato fusarium wilt caused by *F. oxysporum* f.sp. *lycopersici.*

231

Identification of *Streptomyces griseorubens* was done at Tropical Deseases Centre (TDC) of Airlangga University by squensing DNA 16 SRNA **CONSENT**All outhors declare that written informed consent was obtain from the approved of our research parties

242 **REFERENCES**

232 233

243

- Abeysinghe, S. 2007. Biological control of *Fusarium solani* f.spp. *phaseoli* the causal agents of root rot of bean using *Bacillus subtilis CA 32* and *Trichoderma harzianum* RU01. Ruhuna Journal of Science. 2: 62-88.
- 2472. Alabouvette, C., C. Olivain, Q. Migheli, C. Steinberg. 2009. Microbiological control of248soil-borne phytopathogenic fungi with special emphasis on wilt-249inducing *Fusarium oxysporum. On Line Journal* DOI: 10.1111/j.1469250-8137.2009.03014.x
- 3. Anitha, A. and M. Rabeeth. 2010a. Control of Fusarium wilt tomato by bioformulation of Streptomyces griseus in green house condition. African Journal of Basic & Aplied Sciences 1 (1-2): 9 – 14.
- 2544. ______, 2010 b. Degradation of fungal cell walls of phytopatogenic fungi by lytic255enzyme of Streptomyces griseus. African Journal of Plant Science . 4 (3):256061-066.
- 2575. Asha, B., C. Nayaka, U. Shankar, Srinivas, Nirjana. 2011. Biological control of258*F. oxysporum* f. sp. *lycopersici* causing wilt of tomato by *Pseudomonas*259*fluorescens.* International Journal of Microbiology Research. 3(2): 79-84
- 2606. Bollen, G.J. 1974. Fungal recolonization of heat-treated glasshouse soils. Agro261Ecosystems I: 139-155
- 262 7. Cook J. R. and K. F. Baker. 1996. The nature and practice of biological control of 263 plant pathogens. APS PRESS. The American Phytopathological Society. St. 264 Paul, Minnesota.
- 2658. Fahri, Y. and M. Dikilita. 2007. Control of fusarium wilt of tomato by combination of266Pseudomonas florescent, non patogen Fusarium and Trichoderma267harzianum T-22 in greenhouse conditions. Plant Pathology Journal 6(2) :268159-163.
- 269 9. Gruber, S. and V. Seiboth. 2012. Self versus non-self: fungal cell wall degradation In
 270 *Trichoderma. Microbiology* 158:26-34.
- 27110. Kaewchai, S., Soytong, K., and Hyde, K.D. 2009. Mycofungicides and fungal272biofertilizers.Fungal Diversity. 38: 25-50.
- 11. Kyeong, S. J., Hong, M. K., and Bong, K. C., 2000. Purification and antifungal activities
 of an antibiotic produced by *Gliocladium virens* G1 against plant patogen.
 Plant Patholohy Journal, J.17(1): 53-56.
- 12. Larkin, R.P. and D.R. Fravel. 1993. Biocontrol of Fusarium Wilt of tomato. Biocontrol of Plant Deseases Laboratory. Bestvile
- Menzies JG, Koch C, Seywerd F. 1990. Additions to the host range of *Fusarium* oxysporumf. sp. radicis-lycopersici. Plant Diseases. 74: 569–572.
- 14. Morid, B., S. Hajmansoor, N. Kakvan, 2012. Screening of resistance genes to fusarium root rot and fusarium wilt diseases in tomato (*Lycopersicon esculentum*) cultivars using RAPD and CAPs markers. European Journal of Experimental Biology. 2 (4):931-939

284 285 286 287 288	 Olivain C., C. Humbert, J. Nahalkova, J. Fatehi, FL. Haridon, and C. Alobouvete. 2006. Colonitation of tomato root by phatogenic and non patogenic <i>Fusarium</i> <i>oxysporums</i> strains inoculated together and separately into the soil. Aplaid and Enviromental Microbiology. 72 (2): 1523-1531. Reis A, Costa H, Boiteux LS, Lopes CA. 2005. First Report of <i>Fusarium oxysporum</i> f. sp.
289	lycopersici Race 3 on Tomato in Brazil. Fitopatol. Bras. 30(4): 426-428.
290	17. Siddiquee, Shafiquzzaman, Soon G. T., and Y. U. Kalsum. 2010. Isozyme Analisis and
291	Relationships Among Three Species in Malaysia Trichoderma Isolates.
292	Mycrobial Biotechnol. 20(9): 1266 – 1275
293	18. Singh R.,B.K. Singh, R.S. Upadhyay, B. Rai and Y. S. Lee. 2002 Biological control of
294 295	fusarium wilt disease of pigeonpea. Plant Pathology Journal 18(3) : 279-283.
295 296	
296 297	19. Singleton, J.D. Mihail and C.M. Rush. 1993. Methods for research on soilborne phytopatogenik fungi . APS Press. The American Phytophatological Society.
297	St. Paul Minesota.
299	20. Staniazsek M, Kozik EU, Marczewski W . 2007. A CAPS marker TAO1902 diagnostic
300	for the I-2 gene conferring resistance to <i>Fusarium oxysporum</i> f. sp.
301	lycopersici race 2 in tomato. Plant Breeding. 126(3): 331-333.
302	21. Stipanovic, R.D. and C.R. Howell. 1982. The Structure of <i>Gliovirin</i> , A new Antibiotic
303	from <i>Gliocladium virens</i> . The Journal of Antibioticsicotoxin.
304	22. Suharjono, Tri Handayani, Soejono, Susanti Dewi. 2008. Antagonis test of Trichoderma
305	sp. dan Gliocladium sp. againts Fusarium oxysporum cause of wilt
306	deseases of some variety of Purwodadi field banana in Vitro (In
307	Indinesian). Biologi Study, Mathemathic and Scient Faculty, Unibraw
308	Malang, e- <i>mail : <u>calistus@brawijaya.ac.id</u></i>
309	23. Suryaminarsih dan Mujoko. 2012. Growth population of multiantagonis Streptomyces
310	sp. Gliocladium sp and Trichoderma harzianum as biological agents of
311	fusarium wilt disease in natural and semi natural package pellet formula
312	(<i>In Indonesian</i>). Plumula, 1 (2): 202-210.
313	24. Steinkellner S., R. Mammerder, and H. Vierhellig. 2008. Germination of Fusarium
314	oxysporums in root exudates from tomato plants callenged with diffrent
315	Fusarium oxysporums strains. Plant pathology. 122: 395-401
316	24. Titus, A. and G.N. Pereira. 2008. The role of Actinomycetes in coffee plantation
317	ecology. Ineedcoffee.com