# <u>Original Research Article</u> Effects of pesticide use in farmland under with inte nsive soil management use in Southern Brazil

## ABSTRACT

Poor soil management and intensive use of pesticides causes serious damage to soil and water gu ality in Brazil. To confirm this, two studies were conducted in an area with intensive farming in Southern Brazil with the following objectives: i) objectives to evaluate the level of pestici des in the river adjacent to the farmland during different seasons (river study) and to know; ii) to analyze the potential contamination resulting from surface runoff (runoff study). The riv er study was performed with samples from river water and riverbed sediment obtained over a one year period with three months sampling period intervals every three months (differen t seasons) on dry days (base flow effect). The runoff study was performed in the laboratory with simulated rainfall after recent pesticide application. The pesticides analyzed were Teb uconazole, Metalaxyl, Deltamethrin, Chlorothalonil, Glyphosate and its Metabolite-aminome thylphosphonic acid. They represented the most commonly used pesticides in the studied r egion. None of the pesticides tested were found in the riverwater river water or riverbed-se diment riverbed sediment samples at any sampling period time point. The detection limit in river water samples for Glyphosate and its metabolite was5  $\mu$ g L<sup>-1</sup>while it was 1  $\mu$ g L<sup>-1</sup> and f or the other pesticides was 1  $\mu$ g L<sup>-1</sup>. On The other hand, The runoff study (one hour rainfall) demonstrated that all the pesticides were present at high levels. (It was 36 µg L<sup>-1</sup> for Tebuc onazole, 3.24 µg L<sup>-1</sup> for Metalaxyl, and 5.74 µg L<sup>-1</sup> for Chlorothalonil in runoff samples,) sug gesting a high probability of contamination in downstream environments during intense rain fall events after pesticides application. Our data The results highlights the importance of go od management practices to prevent pesticides contamination from agricultural runoff to of downstream environments due to runoff from agricultural lands such as rivers.

Keywords: Agrochemical, catchment, land use, riverbed sediment, runoff, water quality, land use, c atchment, runoff, water quality, riverbed-sediment

#### **1 INTRODUCTION**

Increased in intensive agricultural production has led to a rise in the use of pesticides worldwide. Br azil is now the second largest consumer of pesticides in the world and the eighth per cultivated are a [1]. Pesticides usage can cause environmental damage as well as affects human and animal heal th depending on its toxicity level, time of exposure, quantity amount applied and persistence [2, 3].

Pesticides applied on farmland can reach the water bodies bodiy by surface runoff, leaching (matrix flow) and preferential flow [4]. The fate of pesticides is strongly affected by the natural affinity of th e chemical with the environmental solid, liquid, gaseous and biota states compartments (solid, liquid, gaseous, biota), and this behavior is usually expressed by the soil organic carbon sorption coefficient ( $K_{oc}$ ), water solubility, Henry's constant ( $K_H$ ) and Octanol-water partition coefficient ( $K_{ow}$ )[3, 4, 5]. Movement of pesticides from soil to water depends on factors such as soil texture, soil organic matter [3, 4, 6, 7], topography, and rainfall [8, 9]. Pesticides that are highly adsorbed by soil mineral and organic particles have a lower leaching potential and consequently a high potential for being tr ansported (along with the sediment) by surface runoff along with the sediments [10].

The Water quality standard is set according to risk assessments for environment, and animal and h uman health. This is encoded by environmental laws which define the maximum limits of biological,

chemical and physical elements. In Brazil, normative as ...... [11; 12; 13] [11], [12] and [13] estab lish the maximum limits for pollutants in superficial and ground waters and in soil as ...... [14] in s oil. Also, the Brazilian Health Department, by resolution ....... [15] established limits for drinking w ater as ..... [15]. However, not all pollutant groups are described in the Brazilian legislations, so int ernational legislations, such as the United States Environmental Protection Agency [16, 17] and Eu ropean Union legislations [18] should also be considered.

The Campestrecatchment is located in Colombo, Paraná State, south of Brazil, occupied by family f armers that who produces mainly vegetables to supply Curitiba and the Metropolitan market. In this catchment, most of the arable areas are in conflict with the land use capacity, with very high slope and shallow soils [19]. The conventional system of vegetable production includes intensive soil use as well as an intensive use of pesticides and fertilizers thereby increasing the potential for rivers, la kes, and groundwater contamination. Colombo city plays an important role in for domestic water su pply because of the surface drainage network and the presence of the Karst aquifer [20].

This study therefore Our study included two objectives. Firstly, we assessed the level of pesticides in the river water (base flow) and riverbed-sediment affected by land use in different seasons. It als o investigated Second, under laboratory conditions, we simulated rainfall to analyze runoff potential contamination in events of intense precipitation after immediate recent applications of pesticides of pesticides.

#### 2 MATERIAL AND METHODS

#### 2.1 Physicoal-chemical properties and transport potential of the studied pesticides

The physical – and chemical properties of the studied pesticides are described in Table 1, and the potential for leaching or runoff transport estimated by threemethods three methods which includes GUS, EPA and GOSS (GUS, EPA and GOSS) in (Table 2).

TheGUSindex The GUS index, created by[21] by ...... [21] which is based isbased on thehalf-life t he half-life in soil and the soil organic carbonsorption carbon sorption coefficient(Koc) coefficient (K oc) ([GUS=( loghalf-life log half-life in soil) X (4- logK<sub>oc</sub>)]) [22].Valuesgreater Values greater than 2. 8indicate a highpotential high potential for leaching, while valuesbelow 1.8indicate values below 1.8 indicate thatthis that this pesticide will be lost byrunoff[23] by runoff [23].According to the GUS crite ria(Table 2),metalaxylhas metalaxyl has a highleaching high leaching potential followed by tebucon azole. Conversely, deltamethrin,chorotalonil deltamethrin, chorotalonil and glyphosate have a very I ow leaching potential.

TheEPAmethod The EPA method evaluates the pesticidesaccording pesticides according to the foll owingphysical-chemical following physical-chemical properties: water solubility, soilorganiccarbon s oil organic carbon sorption coefficient (Koc), Henry's constant (K<sub>H</sub>), half-life in soil, half-life inwater and in water and annual rainfall. According to EPAthe EPA the pesticide leaching potential is high w henwater when water solubility > 30 mgL<sup>-1</sup>, Koc<300-500 gmL<sup>-1</sup>, K<sub>H</sub><10<sup>-2</sup> Pa m<sup>3</sup>mol<sup>-1</sup>, half-life in soil >14 to 21 days, half-life inwater days, half-life in water > 175 days and annual rainfall > 250 mm [24]. According to the EPA criteria (Table 2), metalaxylandtebuconazolehave metalaxyl and tebuconazole have a highleaching high leaching potential, while chlorothalonil,glyphosate chlorot halonil, glyphosate and deltamethrin have noleaching no leaching potential.

The GOSS method evaluates the potential transport associated with the sediment as follows: a) hig h potentialassociated potential associated withsedimenttransport(half-life with sediment ransport (half-life in soil  $\geq$  40 days 40 days and K<sub>oc</sub> and K<sub>oc</sub>= 1,000 or half-life 1,000 or half-life in soil  $\geq$ 40 days and K<sub>oc</sub> $\geq$  500 and solubility in water $\leq$ 0.5mg L<sup>-1</sup>; b) low potentialassociated potential associated with the esediment transport (half-life in soil < 1 day orhalf-life or half-life in soil  $\leq$ 2 days and K<sub>oc</sub> and K<sub>oc</sub>  $\leq$  500 or half-life in soil < 4 days and K<sub>oc</sub> $\leq$ 900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility in water  $\geq$  0.5mg L<sup>-1</sup> or half-life in soil  $\leq$ 40 days and K<sub>oc</sub> $\leq$  900 and solubility i

d in water transport (half-life in soil > 35 and  $K_{oc}$ < 1,000,000 and solubility in water ≥ 1 mgL<sup>-1</sup> or  $K_{oc}$  ≤700 and solubility and solubility in water between 10 and 100mg L<sup>-1</sup>); d) low potential dissolved pote ntial dissolved in water transport ( $K_{oc}$ ≥1,000,000 or half-life 000 or half-life in soil ≤ 1 day and  $K_{oc}$ ≤100 or half-life or half-life in soil <35 days and solubility in water<0.5 mg L<sup>-1</sup>); e) substances that substan ces that do not fit not fit into anyof any of the above criteria are considered to have an average pote ntial to pollute surface water [25]. Following these criteria these criteria (Ta ble 2), tebuconazole and metalaxyl have low potential associat ed with sediment transport and high potential dissolved in water. Chlorothalonil and deltamethrin are in a transitio n zone between low and high potential associated with sediment transport while glyphosate had a low potential fortransport dissolved inwater for transport dissolved in water.

Pesticides											
	Tebuconazole Metalaxyl Chlorothalonil Deltamethrin Glyphos										
M (g mol <sup>-1</sup> )	307.82	279.33	265.9	505.2	168.07						
S (mg L <sup>-1</sup> )	36	7100	0.81	0.0002	10500						
V. P. (mPa)	0.0013	0.75	0.076	0.0000124	0.0131						
M.P. (°C) K <sub>ow</sub>	105 5011.87	67.9 44.66	256.1 758.57	101 39810.71	189.5 0.001						
K <sub>oc</sub>	769	500	850	10240000	21699						
K <sub>H</sub> (Pa m <sup>3</sup> mol⁻¹)	1 10 <sup>-5</sup>	1.60 10 <sup>-5</sup>	2.50 10 <sup>-2</sup>	3.10 10 <sup>-2</sup>	2.10 10 <sup>-7</sup>						
DT50 soil (days)	62	42	22	13	12						
DT50 water (days)	356	56	22	65	87						

Table 1. Physical-chemical properties of the pesticides [26].

M- Molecular mass, S- Solubilityin water, V.P.-Vapor pressure, M.P. - Melting point,  $K_{ow}$ -Octanol-wat er partition coefficient,  $K_{oc}$ -Soil organic carbon sorption coefficient,  $K_H$ . Henry's constant, DT50 soil - Half-life in soil, DT50 water - Half-life in water.

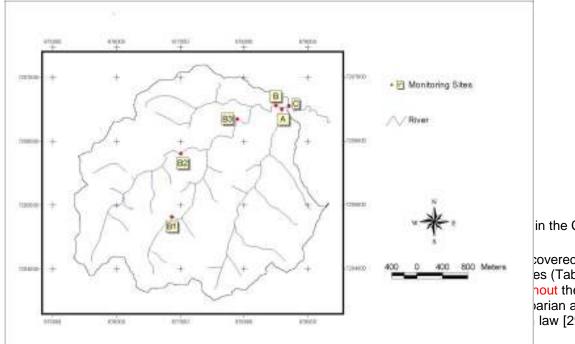
Table 2. Leaching and runoff potential according to GUS, EPA and GOSS EPA and GOSS criteria.

Pesticides									
	Tebuconazole	Metalaxyl	Chlorothalonil	Deltamethrin	Glyphosate				
GUS EPA	2.00 (high leaching potential) High leaching potential	2.11 (high leaching potential) High leaching potential	1.44 (no leaching potenti al) Nol eaching potential	-3.35 (no leaching potenti al) No leaching potential	-0.36 (no leaching pote ntial) No leaching potential				
GOSS		low potential with sediment and high potential dissolved in water		between low and high pot ential with sediment	low potential dissolved i n water				

# y 1 – Pesticides in the river

#### a characterization

v was carried out in Colombo, Metropolitan region of Curitiba, Paraná state, Southern Brazil (Figure 1). The Campestre catchmentbelongs of o the Capivari river catchment. The climate is mesothermal humid subtropical (Cfb) by Köppen with average annual rainfall of 1400 to 160 Cambisol is the predominant soil, with Leptsol mainly on the top of the hills[28].





covered by native vegetation (57%) (Table 3). However, 19% is es (Table 4) cropped by small family farmers with several kinds nout the entire year (winter and summer cultivar; using the conv parian area that should be preserved by law is not covered with law [29], the drainage network should have 30 m each side po

		Land use										
Monitoring sites	Area	Native Forest		Refores	station	Agriculture		Grassfield		Other		
	ha	ha	%	ha	%	ha	%	ha	%	ha	%	
А	331	164	50	89	27	28	8	48	15	2	0.6	
В	675	274	41	144	21	163	24	90	13	5	0.7	
С	1010	440	44	234	23	192	19	138	14	6	0.6	

Table 3. Land use (ha and %) in the Campestre catchment, Colombo, Paraná, Brazil.

Table 4. Slope classes and land use (ha and %) in the Campestre the Campe stre catchment, Colombo, Para ná, Brazil.

Slope classes (%)

	Total	2.5	0.4	43.5	0.4	104.1	15.4	170.4	20.Z	204.4	42.1	50.0	0.4	14.1	2.1
Agricultu	Agriculture	0.6	0.3	15.2	7.9	40.4	21,0	58.9	30.5	68.8	35.7	7.2	3.7	1.6	0.8
C	Total	3.1	0.3	50.4	5.0	141.0	14.0	239.5	23.7	450.0	44.6	100.1	9.9	26.2	2.6

Monitoring sites	Land use of the riparian area											
	Native Forest		Reforestation		Agriculture		Grassfield		Other		Total	
	ha	%	ha	%	ha	%	ha	%	ha	%	ha	%
А	27	60	10	22	3	6	5	10	0.4	0.9	44	100
В	49	55	10	11	17	19	13	14	0.4	0.4	89	100
С	77	57	20	15	20	15	18	13	0.8	0.6	135	100

Table 5. Land use of the riparian zone in the Campestre the Campe stre catchment, Colombo, Paraná, Brazil.

#### 2.2.2 Monitoring sites and sampling

Six monitoring sites were selected for water analysis. Site C represents the entire study area (Figure 1) and s ite A and B represent the sub-basins.

The river water sampling was carried out from September 2008 to September 2009 every three months inter val. On September 9<sup>th</sup>, 2008 (spring) and June 3<sup>rd</sup>, 2009 (winter) riverbed-sediment wasalso was also sample ed. Soil (0-10 cm and 10-20 cm)from field cropped with vegetable wasalso was also sampled on September 9<sup>th</sup>, 2008 (spring) and June 3<sup>rd</sup>, 2009 (winter) in a conventional management system in the experimental area conducted by [30]. On the sampling of March 3<sup>th</sup>, 2008 (autumn), December 15<sup>th</sup>, 2008 (summer) and September 15<sup>th</sup>, 2009 (spring), only water from the river was sampled. All river samples were collected on dry days in polyethylene bottles, transported in ice boxes to the Food Processing Research Center at the Federal Uni versity of Paraná and kept under refrigeration at a temperature of 5° C (was that the temperature of the river site????) until further pending laboratory the time for analysis.

#### 2.2.3 Pesticide analysis

A survey of the most applied pesticides in the region was carried out. As a result, tebuconazole, metalaxyl, d eltamethrin, chlorothalonil and glyphosate were chosen for analysis in the present study.

The extraction of pesticides (tebuconazole, metalaxyl, deltamethrin, chlorothalonil)from chlorothalonil) from river water samples was performed using decantation funnels as described inApha[31]. An aliquot of 1 literof liter of the sample was added to a 60 mL beaker containing 15% of the solvent hexane/dichloromethane (v/v) (15% analytic grade) and agitated for three minutes. The sample was then drained from the solvent with a se parating funnel. This procedure was repeated three times. The extracted solvent (180 mL) was dried in a vap orizer and the remaining (1 mL) was injected in to a gas chromatographer and mass spectrophotometer with an electron detector [31]. Forthis method, the detectionlimitfor eachpesticidewas For this method, the detection n limit for each pesticide was 1  $\mu$ g L<sup>-1</sup>.

Glyphosate and its metabolite (aminomethylphosphonic acid - AMPA) in the river water samples was analyze d by the Chelex – 100 column[32] after filtering the acidified sample (pH 2.0  $\pm$  0.4 with HCl 6 M) through a gla ss filter. The detection limitforglyphosatein waterwas limit for glyphosatein water was 5  $\mu$ g L<sup>-1</sup>.

For the analysis of the pesticides (tebuconazole, metalaxyl, deltamethrin, chlorothalonil)in chlorothalonil) in riverbed -sediment and soil, 30 g of the sample was added to 20 mL of the solvent ethyl acetate and agitated f

remained. After that it was applied to the column with resin AG<sup>a</sup> 50W-X2. The chromatographic column (4 x 1 50 mm) Glyphosate (Pickering) with guard Glyphosate (Pickering) column (3 x 20 mm) was used with 0.4 mL min<sup>-1</sup> flow of mobile phase, oven temperature of 55 °C, and post column oven. A fluorescence detector was used with emission 430 nm and excitation of 340 nm. The injection volume was 50  $\mu$ L and the retention time for the glyphosate was 13.60 and 26.49 minutes for its metabolite[34]. The detectionlimitof glyphosateinriverb ed-sediment and soil was 0.1 mg kg<sup>-1</sup>.

#### 2.3 Study 2 – Pesticides in the runoff

#### 2.3.1 Preparation of erosion boxes

#### 2.3.1 Preparation of erosion boxes

This study was carried out in erosion boxes with a rainfall simulator. Runoff samples were analyzed by the Br azilian Agricultural Research Corporation – EmbrapaForestry. Embrapa Forestry. The top soil (0-5 cm)was (0 -5 cm) was collected from the Campestre catchment area, Colombo, Parana State, at the same field of the ri ver study. Before filling the erosion box (30 cm wide, 40 cm long and 10 cm deep, with small roles on the bott om for drainage) the soil was sieved through a 5 mm mesh and dried. The boxes were filled with 7.5 cm of dr ied fine sand (washed with HCl 3% and deionized water to eliminate any contamination). The upper 2.5 cm w as filled with soil using a field bulk density of 0.92 g cm<sup>-3</sup> [30]. Some physical and chemical attributes of the so il (0-20 cm) [30]: organic carbon (30.5gkg<sup>-1</sup>); clay (280gkg<sup>-1</sup>), silt (370 gkg<sup>-1</sup>) and sand (350 gkg<sup>-1</sup>). Six boxes were used per pesticide. The erosion boxes were protected with a 5 cm high galvanized plate to avoid lateral losses and the runoff was collected in a bucket by a covered funnel placed at the end of the erosion boxes.

#### 2.3.2 Pesticide application and rainfall simulation

#### 2.3.2 Pesticide application and rainfall simulation

Three commercial products were used for the experiment. For Tebuconazole the Folicur<sup>®</sup> 200 EC (Bayer; 20 0 g L<sup>-1</sup> of Tebuconazole) was used following the recommendation for beetroot (1 L of the commercial product per hectare). For chlorothalonil and metalaxyl the Folia Gold<sup>®</sup> (Syngenta Gold<sup>®</sup> (Syngenta; 675 g kg<sup>-1</sup> of Chlor othalonil and 67.5 g kg<sup>-1</sup> of Metalaxyl) was used following the recommendation for tomatoes (1.5 kg ha<sup>-1</sup>). For Deltamethrin the K-Othrine<sup>®</sup> SC 25 (Bayer; 25 g L<sup>-1</sup> of Deltamethrin) was used following the recommendation of 500 L per hectare).

To simulate rainfall, a programmable simulator equipped with a nozzle (Veejet 80-100) was used with deioniz ed de-ionized water. The simulator was placed 2.4 m from the ground and the erosion boxes inclined 12%, si mulating the field hillside slope. To obtain moisture uniformity, a rainfall of 20 mm h<sup>-1</sup> was simulated for 10 mi nutes. After that, a rainfall intensity of 60 mm h<sup>-1</sup> was applied for one hour. The runoff was collected twice (30 and 60 minutes). The runoff volumes were recorded and a representative sample was refrigerated for further analysis.

Pesticides were applied in 100 mL of deionized de-ionized water, according to recommendations per hectare and using a spray bottle for better product distribution and moisture uniformity. The pesticides were applied at night to avoid higher temperatures, thus preventing chemical breakdown. Rainfall was simulated 12 hours after pesticide application.

#### 2.3.3 Pesticide analysis

#### 2.3.3 Pesticide analysis

Prior to pesticide extraction, samples were passed through a 0.45 μmcellulose ester membrane. The extraction of the pesticides was carried out as in Study 1. The bromatographic analysis was performed by performed

s operated with spectrometer was operated with an electron impactof70eV impact of 70 eV. To quantifythe quantify the pesticides the following fragments: m/z265 for chlorothalonilm/z205 formetalaxyl,m/z250 Chlorothal onil m/z205 Formetalaxyl m/z250 for tebuconazoleand Tebuconazole and the m/z 180 Fordeltamethrin were used. Quantification was performed against an external standard using a calibration curve.

To validate this method, the amount of agrochemical recovered from 1 liter of ultrapure water with 0.8  $\mu$ g L<sup>-1</sup> of the standard pesticide was measured. The recovered value (40 to 120%) was within the values recommen ded by [35].

The detection limit was determined based on the standard deviation and inclination of the calibration curve wi th the formula: LOQ = 10 (SD / S), where LOQ is the detection limit; SD is standard deviation and S isinclinat ion is inclination of calibration curve [36]. The detection limit obtained for Metalaxyl was the lowest, 1.92 ng L<sup>-1</sup>, and the highest value was for Deltamethrin, 23.59 ng L<sup>-1</sup>.

#### **3 RESULTS AND DISCUSSION**

## 3.1 Study 1 – Pesticides in the river

None of the analyzed pesticides (Metalaxyl, Chlorothalonil, Deltamethrin, Tebuconazole, Glyphosate and AM PA) were detected in any of our riverwater the river water samples above the detection limits (1  $\mu$ g L<sup>-1</sup> for Met alaxyl, Chlorothalonil, Deltamethrin and Tebuconazole and 5  $\mu$ g L<sup>-1</sup> for glyphosate and its metabolite). The de tection limit for glyphosate andtebuconazolewere muchlower and Tebuconazole were much lower than them aximum the maximum value allowed fordrinking for drinking water according to the Brazilian Ministry of Heal th (500 mgL<sup>-1</sup> and 180 $\mu$ g L<sup>-1</sup>, respectively) [15]. This was also lower than the limit for glyphosate (65  $\mu$ g L<sup>-1</sup>) in fresh water established by the Brazilian Environmental Council [12].For the other pesticides there are were n o maximum values defined by the Brazilian laws. USEPA [37] has a higher maximum limit for glyphosate indr inking in drinking water (700  $\mu$ g L<sup>-1</sup>). However, the maximum limit established by [18] is 0.1  $\mu$ g L<sup>-1</sup> for any pesticide and pesticide [18] and the sum of the pesticides should not be higher than 0.5  $\mu$ g L<sup>-1</sup>.

Due to the soil type (low depth), steep slopes, intensive soil and agrochemicals used, we expected to find pe sticides were expected to be found in the river water. The [38] Authors have analyzed the water quality of the Campestre catchment area for one year and also found a very low concentration of nitrogen, phosphorus, a nd carbon [38].

Low pesticide levels in the river water can be explained by both the catchment land cover and the sampling ti me. the fact that most of the catchment area is covered by forest (41% of native and 24% of planted forest), r esulting in buffering the effect of agriculture (arable land), which represents only 19% of the catchment. Rete ntion of effect on pesticides in native vegetation has been demonstrated with the due to major contributor bei ng adsorption by soil organic matter [39, 40].

In addition, all samples were collected during dry days. At these sampling days, there would be there was littl e contamination by runoff which was against the normal trend that should normally follows intense rainfall. Th e sampling in days without precipitation, on the other hand, suggests showed that the subsurface water whic h supplies of the river is was not contaminated. However, it is important to note that the detection limits in the present study (1  $\mu$ g L<sup>-1</sup>) were above the concentration obtained in rivers by several some authors [41, 42]. In The [43] in a study carried out in the Mediterranean Sea, it was found that contamination levels of metalaxyl and chlorothalonil in the River Rhône (France) and River Pó (Italy) were below 2 and 1 ng L<sup>-1</sup>, respectively. T herefore, in the Campestre catchment, the pesticides might be present in the samples analyzed, but with a c oncentration below the detection limit 1  $\mu$ g L<sup>-1</sup> (1  $\mu$ g L<sup>-1</sup>).

sorption and persistence of Glyphosate in the soil makes the presence of its metabolite in the sediment highly likely.

## 3.2 Study 2 – Pesticides in simulated runoff

For all pesticides, the highest concentrations in runoff water for all pesticides were detected in the first 30 minutes and it decreased decreasing with rainfall duration (Figure 2). These results confirmed the hypotheses t hat intense precipitation may increase river contamination (Authority). In this study, only the dissolved fraction no f the pesticides (which passed through a 0.45  $\mu$ m cellulose membrane) was analyzed and so chemicals tr apped in the particulate fraction were not extracted. Very high concentrations of pesticides in the dissolved fr action (3.24  $\mu$ g L<sup>-1</sup> for metalaxyl, 36  $\mu$ g L<sup>-1</sup> for tebuconazole, and 5.74  $\mu$ g L<sup>-1</sup> for chlorothalonil) were obtained after one hour of rainfall (Figure 2). Deltamethrin was not detected during the last 30 minutes of rain, showin g the low potential for being transported in a dissolved fraction in the surface runoff (in the surface runoff). Ev en With a greater runoff volume in the final 30 minutes of rainfall, with values of ~2.52 L against 1.69 L in the first 30 minutes), pesticides loss was greater in the first 30 minutes (Table 6). This was however, the total amount of pesticides lost by runoff were very low compared with t he total amount applied (Table 6). We saw However, there was greater losses in the pesticide Tebuconazole with 0.71 % of the total applied was lost in the one hour runoff (0.71 % of the total applied was lost in the one hour runoff).

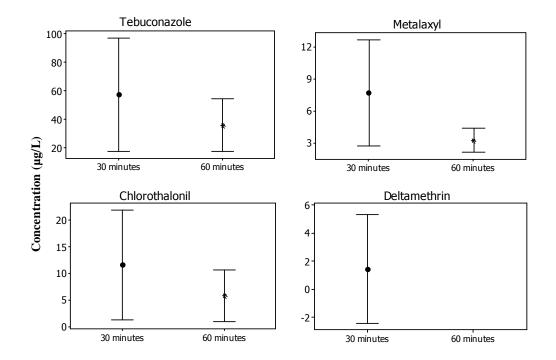


Fig.2. Mean concentration ( $\pm$  standard deviation) of Chlorothalonil, Metalaxyl, Tebuconazole and Deltamethri n dissolved in surface runoff (30 and 60 minutes) under simulated rainfall at 60 mm h<sup>-1.</sup>

Table 6..Pesticide losses through surface runoff under rainfall simulation Losses of pesticides by surface run off under rainfall simulation.

Following the GUS method (Table 2), we expected loss of tebuconazole and metalaxyl in the dissolved fracti on of the runoff was expected. This was observed with Tebuconazole, but not with Metalaxyl (Table 6). Simil ar The [45] also observed high levels of dissolved Tebuconazole in surface water has been reported [45]. Te buconazole has also been detected in streams, wastewaters and lakes [46, 47, 48, 49], and hence this fungi cide poses a risk of runoff transport.

Chlorothalonil and Deltamethrin were was expressed at low levels in runoff (dissolved fraction), which is in a greement with the GUS, EPA and GOSS GUS, EPA and GOSS models (Table 2). These are expected to stron gly adsorb to soil organic particles due to its high Koc (Table 1) [50]. Chlorothalonil was applied at a higher c oncentration (Table 6) and was detected at lower levels. Some authors have The [51] observed small losses of chlorothalonil by leaching [51], supporting the fact concluding that this agrochemicals has a greater potenti al for loss by runoff in the particulate fraction. The Chlorothalonil was developed to degrade in less than four weeks in water, however, it was found in most of the Greek estuarines [52] suggesting its persistence in the r iverbed sediments.

Deltamethrin is degraded in one to two weeks [53], which may explain the fact that we could not find this che mical in soil or river water (after how long?). The results of [54] analyzed contamination of waters in the Pant anal and found no Deltamethrin in the environment [54], attributing this to its physical-chemical properties an d low use.

However, even with a small percentage of the applied pesticides being lost by runoff, the concentrations can could be high enough to cause serious environmental and human health problems. To avoid contamination i n the river waters, pesticides use should be carefully managed.

## **4 CONCLUSION**

The pesticides Tebuconazole, Metalaxyl, Chlorothalonil, Deltamethrin, Glyphosate and its their metabolites w ere not found in any of the riverwater river watet or riverbed-sediment samples from the Campestre catchme nt area. However, it must be considered that all sampling was carried out on dry days (base flow effect) with no influence of agricultural runoff from intense rainfall storms. On the other hand, our simulated rainfall study demonstrated a high potential for pesticide contamination by surface runoff (dissolved fraction<  $0.45 \mu$ m). In addition to pesticide management it is also important to perform soil management to prevent pollutants conta ined in agricultural runoff from reaching river waters.

# REFERENCES

1. Anvisa - Agência Nacional de Vigilância Sanitária. Resíduos de agrotóxicos em alimentos. RevistaSaúdePú blica. 2006; 40:361-363.

2. Ecobichon DJ. Pesticide use in developing countries. Toxicology.2001; 160:27-33.

3. Fao – Food Agriculture Organization of the United Nations. Control of water pollution from agriculture. GE MS/Water Collaborating Centre- Canadá Centre for Inland Waters, Burlington, Canada, Paper n. 55. 1996.

4. Pierzynski GM, Sims JT and Vance GF Soils and environment quality. 2 ed. CRC press, Boca Raton; 200 0.

5. Gramatica P and Guardo AD. Screening of pesticides for environmental partitioning tendency. Chemosphe re.2002; 47:947–956.

6. Filizola HF, Ferracini VL, SansLMA, Gomes MAF and FerreiraCJA. Monitoramento e avaliação do risco de contaminação por pesticidas em água superficial e subterrânea na região de Guairá. PesquisaAgropecuáriaB rasileira.2002: 37:659-667.

des in Malaysian agricultural soils. Pesticide Science. 1997; 50:53-63.

11. Brasil - Conselho Nacional do Meio Ambiente (CONAMA). Resolução Conama nº. 357. Diário Oficial da U nião de 17/03/2005. Brasília;2005.

12. Brasil - Conselho Nacional do Meio Ambiente (CONAMA). Resolução Conama nº. 396. Diário Oficial da U nião de 30/04/2008. Brasília; 2008.

13. Cetesb - Companhia de Tecnologia de Saneamento Ambiental. Relatório de estabelecimento de valores orientadores para solos e águas subterrâneas no Estado de São Paulo. Cetesb, São Paulo, Brasil; 2014.

14. Brasil - Conselho Nacional do Meio Ambiente (CONAMA). Resolução Conama nº. 460. Diário Oficial da U nião de 30/12/2013. Brasília; 2013.

15. Brasil - Ministério da Saúde. Portaria do Ministério da Saúde nº 2914. Diário Oficial da União de 14/12/201 1. Brasília; 2011.

16. Usepa – United State Environmental Protection Agency.Water quality standards handbook. Usepa, Wash ington; 1995.

17. Usepa – United State Environmental Protection Agency. Code of federal regulations 40. Part 131 – Water quality standards. Usepa, Washington; 1995.

18.European Union.Council Directive 98/83/EC of 3 November 1998 on the quality of water intended for hum an consumption. Official Journal of the European Communities. Bruxelas; 1998.

19. Soares MGJ, Mellek JE, Orrutéa AG, Kummer L, Nunes T, BarrosYJ, Andretta R, Favaretto N and Souza LCP.Potencial de uso agrícola e fragilidade ambiental da microbacia do rio Campestre, Colombo – PR. Scie ntia Agraria.2008; 9:587-596.

20. AlmeidaL de. Mudanças Técnicas na Agricultura: Perspectivas da transição Agroambiental em Colombo-PR. Ph.D Thesis, UniversidadeFederal do Paraná. Curitiba, Brazil; 2003.

21. Gustafson DI. Groundwater ubiquity score: a simple method for assessing pesticide leachability. Environ mental Toxicology and Chemistry.1989; 8:339–357.

22. Andrade AIASS andStigter TY. Multi-method assessment of nitrate and pesticide contamination in shallo w alluvial groundwater as a function of hydrogeological setting and land use. Agricultural Water Managemen. 2009; 96:1751–1765.

23. Wilson SC, Duarte-Davidson R and Jones KC. Screening the environmental fate of organic contaminants in sewage sludgesapplied to agricultural soils: The potential for downward movement to groundwaters. The Science of the Total Environment.1996; 185:45-57.

24. Cohen SZ, Wauchope RD, Klein AW, Eadsforth CV and Graney R. Offsite transport of pesticides in water mathematical models of pesticide leaching and runoff. International Union of Pure and Applied Chemistry.19 95: 67:2109-2148.

25. Goss DW. Screening procedure for soils and pesticides for potential water quality impacts. Weed Techno logy. 1992; 6:701-708.

26. University of Hertfordshire. Eletronic publishing at footprint: Description of fungicides and insecticides. 20 10. Acessed. 09 June 2010. Available at http://sitem.herts.ac.uk/aeru/footprint/en/index.htm.

27. lapar - Fundação Instituto Agronômico do Paraná Cartas climáticas do estado do Paraná. 2011. Accessed 29 april 2010. Available at http://www.iapar.br.

28. Waltrick JCN. Aplicação da metodologia P-index na bacia hidrográfica do campestre – Colombo, PR. Mas ter Thesis, Universidade Federal do Paraná, Curitiba, Brazil; 2011.

29. Brasil - Conselho Nacional do Meio Ambiente (Conama) Resolução Conama nº. 302. DiárioOficial daUniã o de 13/05/2002.Brasília; 2002.

30. Ramos MR, Favaretto N, Dieckow J, Dedeck RA, Vezzani FM, Almeida L, Sperrin M. Soil, water and nutr ient loss under conventional and organic vegetable production managed in small farms versus forest system. Journal of Agriculture and Rural Development in the Tropics and Subtropics.2014; 115:31-40.

31. Apha -American Public Health Association. Standard methods for the examination of water and wastewat er. 19ed., American Public Health Association, Washington; 1995.

32. Cowell JE, Kunstman JL, Nord PJ, Steinmetz JR and Wilson GR. Validation of an analytical residue meth od for analysis of glyphosate and metabolite. Journal of Agricultural and Food Chemistry. 1986; 34:955-960.

33. Peres TB, PapiniS, Marchetti M, Nakagawa LE, Marcondes MA, Andréa MM andLuchiniLC.Métodos de e xtração de agrotóxicos de diversas matrizes. Revista Arquivos do Instituto Biológico. 2002; 69:87-94.

34. Spann KP and Hargreaves PA. The determination of glyphosate in soils with moderate to high clay conte

pril 2010. Available at http://extoxnet.orst.edu/pips/glyphosa.htm

38. Ribeiro K, Favaretto N, Dieckow J, Souza ICP, Minella JPG, Almeida L, Ramos MR. Quality of surface wat er related to land use: a case study in a catchment with small farms and intensive vegetable crop production in southern Brazil. Revista Brasileira de Ciência do Solo. 2014; 38:656-668.

39. BicalhoSTT, Langenbach T, Rodrigues, RR, Correia FV, Hagler AN, Matallo MB and Luchini LC.Herbicid e distribution in soils of a riparian forest and neighboring sugar cane field. Geoderma.2010; 158:392-397.

40. Pinho AP, Matos AT, Costa LM, Morris LA, Jackson RC, White Wand Martinez MA.Retenção de atrazina, picloran e caulinita em zona ripária localizada em área de silvicultura. EngenhariaAgrícola. 2004; 12:260–27 0.

41. Mañosa S, Mateo S and Guitar R. A review of the effects of agricultural and industrial contamination on t he Ebro Delta biota and wildlife. Environmental Monitoring and Assessment. 2001; 71:187–205.

42. Malik A, Ojha P and Singh KP. Levels and distribution of persistent organochlorine pesticide residues in water and sediments of Gomti river (India)—a tributary of the Ganges river. Environmental Monitoring and As sessment. 2009; 148:421–435.

43. Readman JW, Albanis TA, Barcelo D andGalassi J. Fungicide contamination of mediterranean estuarine waters: results from a med pol pilot survey. Marine Pollution Bulletin. 1997; 34:259-263.

44.Usepa – United State Environmental Protection Agency. Ground Water & Drinking Water: Drinking Water Contaminants. Usepa, Washington; 2000.

45. Ferracini VL, Pessoa MCYP, Silva A and Spadotto CA. Análise de risco de contaminação das águas subt errâneas e superficiais da região de Petrolina (PE) e Juazeiro (BA). Pesticida:Revista de Ecotoxologia e Mei oAmbiente. 2001; 11:1-16.

46. Berenzen N, Lentzen-Godding A, Probst M, Schulz H, Schulz R and Liess M. A comparison of predicted and measured levels of runoff-related pesticide concentrations in small lowland streams on a landscape level . Chemosphere. 2005; 58:683–691.

47. Kahle M, Buerge IJ, Hauser A, Müller MD andPoiger T. Azole fungicides: occurrence and fate in wastewa ter and surfacewaters. Environmental Science & Technology.2008; 42:7193–7200.

48. Komárek M, Čadková E, Chrastný V, Bordas F and Bollinger JC. Contamination of vineyard soils with fun gicides: A review of environmental and toxicological aspects. Environment International.2009; 36:138-151.

49. Herrero-Hernández E, Andrades MS, Marín-Benito JM, Sánchez-Martín M and Rodríguez-Cruz MS. Field -scale dissipation of tebuconazole in a vineyard soil amended with spent mushroom substrate and its potenti al environmental impact. Ecotoxicology and Environmental Safety.2011; 74:1480–1488.

50. Regitano JB, Prata F, Dias NM, Lavorenti A and Tornisielo VL.Sorção-Dessorção do fungicida clorotalonil em solos com diferentes teores de matéria orgânica. RevistaBrasileira de Ciência do Solo.2002; 26:267-274.

51. Kahl G, Ingwersen J, Nutniyom P, Totrakool S, Pansombat K, ThavornyutikarnP andStreck T. Loss of pe sticides from a litchi orchard to an adjacent stream in northern Thailand. European Journal of Soil Science.20 08; 59:71–81.

52. Albanis TA, Lambropoulou DA, Sakkas VA andKonstantinou IK.Antifouling paint booster biocide contami nation in Greek marine sediments. Chemosphere.2002; 48:475-485.

53. Extoxnet- Extension Toxicology Network. Pesticide information profiles: Delthamethrin. 1996. Accessed 2 9 april 2010. Available at http://extoxnet.orst.edu/pips/dicofol.htm.

54. Laabs V, Amelung W, Pinto AA, Wantzen M, Silva CJ andZech W. Pesticides in surface water, sediment, and rainfall of the northeastern Pantanal basin, Brazil. Journal of Environmental Quality. 2002; 31:1636-164 8.