

Dry bean response to fertilization using Minjingu phosphate rock and composted Tughutu (*Vernonia subligera* O. Hoffn)

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ABSTRACT

Aims: The objectives of the study were to evaluate the effects of organic materials from Tughutu (*Vernonia subligera* O.Hoffn) and locally available Minjingu phosphate rock or commercial Triple super phosphate fertilizer on the plant growth and amount of phosphorus (P) and calcium (Ca) content in bean shoots (*Phaseolus vulgaris* L.) and their residual effects in the soil after harvest.

Study design: The experiments were set up as a randomized complete block design

Place and Duration of Study: Field experiments were conducted in famers' fields in the Western Usambara Mountains in Tanzania.

Methodology: Each farmer's field was assigned as a replicate. The experimental treatments consisted of six plots as follows: control, Minjingu Phosphate Rock (MPR) or Triple Super Phosphate (TSP) each at 26 kg P ha⁻¹, Tughutu at 2.5 t dry matter ha⁻¹ and Tughutu at 2.5 t dry matter ha⁻¹ combined with Minjingu Phosphate Rock or Triple Super Phosphate at 26 kg P ha⁻¹.

Results: The dry matter yield and amount of P and Ca in bean shoots significantly increased by supplying the organic and inorganic fertilizers above the control treatment. Addition of P fertilizers alone or in combination with Tughutu also modified the soil pH and significantly increased the concentration of P and Ca in the soil at harvest.

Conclusion: Overall, combination of Minjingu phosphate rock or Triple super phosphate with Tughutu enhanced the effectiveness of these fertilizers and resulted in better growth and greater P and Ca amount in the shoots as well as their concentration in the soil at harvest.

Keywords: dry matter, pH, triple super phosphate, Minjingu Phosphate Rock

1. INTRODUCTION

The densely populated areas of the Western Usambara Mountains (WUM) in northern Tanzania suffer from low soil nutrient levels [1,2] owing to intensive cultivation of crops without the application of external inputs. This practice has of late resulted in the depletion of essential plant nutrients, including P and Ca that are important for plant growth [2]. These constraints, amongst others, have been reported to be the cause of poor growth of bean and a major threat to the sustainability of the small-scale production systems in Western Usambara Mountains [2,3]. Phosphorus deficiency is known to impair early nodule functioning [4], retard plant growth and reduce the ability of the plant roots to release protons that may be useful in the solubilization of organic and inorganic P sources in the soil [5, 6]. Calcium deficiency in legumes is also known to impair N₂ fixation and retard plant growth [7, 8, 9, 10, 11, 12].

The majorities of small-scale farmers living in Western Usambara Mountains are poor and cannot afford to purchase expensive agricultural inputs such as Triple super phosphate or lime that could supply phosphorus and calcium to the commonly grown crops in the area, including beans [13]. Therefore, there is a great need to explore the available indigenous sources of nutrients that will ultimately be affordable to the poorly resourced farmers.

Incorporating some organic materials in the soil may contribute significantly in providing mineral nutrients needed by the plants [14, 15] and hence increase the crop yields. The organic matter may consist of a wide range of mineral nutrients, upon the process of decomposition and mineralization, they may be made readily available for plant uptake and growth [16], [17]. Additionally, the use organic plant materials may improve other important soil characteristics such as their water retention and the microbiological activities in the soil [18].

The exploratory work by [17] revealed that traditional shrub Tughutu contained more P and Ca than *Tithonia diversifolia*, the most researched shrub species in East Africa [18, 19, 20, 21]. A number of studies also reported that Minjingu phosphate rock fertilizer has higher contents of water soluble P and Ca [22, 23,24]. From this background, it is evident that these two indigenous resources could be used to boost agricultural production and improve people's livelihood in poor soils of the Western Usambara Mountains.

A few studies conducted in East Africa have shown improved growth and increased concentration of P and Ca nutrition in plants following the application of organic materials and phosphate rocks as fertilizers [1,15,16,24,25,26,27,28,29,30,31,32,33]. Significant synergistic effects were reported in growth or yield by combining the organic and inorganic P fertilizers such as rock phosphates or Triple Super Phosphate [1, 14, 15, 16, 29, 30, 31, 34].

This study was conducted with the objective of evaluating the effects of supplying Tughutu alone, or in combination with locally available Minjingu phosphate rock or commercial Triple Super Phosphate fertilizer on plant growth, and amounts of P and Ca in bean shoots. Furthermore, soil chemical properties such as pH, P and Ca availability were assessed at harvesting.

2. MATERIAL AND METHODS

2.1 Site location and description

Six sites in farmers' fields were planted to beans in Western Usambara Mountains (04°_47.976 S, 38°_17.623' E) during the 2000 - 01 cropping season. The Western

Usambara Mountains are located in an altitude ranging from 900 to 2300 m. a.s.l. and the major soils are classified as Humic and Chromic Acrisols, Luvisols and Lixisols [28]. Before planting, soil samples were collected from each site, bulked and analysed separately for pH, P, and Ca content for each site. Details of soil collection and analysis are described in [2].

2.2 Field trials

The fields were ploughed by a hand hoe. The experiments were set up as a randomized complete block design. Each farmer's field was assigned as a block. The experimental treatments consisted of six plots assigned to each block as follows: control, Minjingu phosphate rock or Triple Super Phosphate each at 26 kg P ha⁻¹, Tughutu at 2.5 t dry matter ha⁻¹ and Tughutu at 2.5 t dry matter ha⁻¹ combined with Minjingu phosphate rock or Triple Super Phosphate at 26 kg P ha⁻¹. P levels from Minjingu phosphate rock and Triple Super Phosphate were calculated considering the water soluble P in each fertilizer. Each treatment was planted in 25 m² plot. Inter-row and intra-row spacing of 50 cm and 20 cm respectively were used. Tughutu used in this study was collected from the Magamba forest reserve in Western Usambara Mountains. Fresh Tughutu leaves were surface applied and incorporated into the soil to 15 cm depth by hand hoe one week before sowing and allowed to decompose. [17] Recommended that Minjingu phosphate rock be broadcasted and incorporated into the soil by a hand hoe at sowing. Triple Super Phosphate was applied into the rows at planting. Two bean seeds coated with *Rhizobium tropici* strain CIAT 899 (10⁹ g cells seed⁻¹) purchased from the Nairobi MICERN, University of Nairobi, Kenya, were planted in each hole and allowed to grow to maturity. The inoculants packets were supplied with gum arabic for sticking as many cells as possible onto the seeds. Bean variety Lyamungu 90 used in this study was supplied by the breeder of the Selian Agricultural Research Institute, Arusha, Tanzania.

2.3 Plant harvest and analysis

Twenty plants were sampled 45 days after planting Di Ammonium Phosphate (DAP) from the two inner rows (1 m²) in each plot for the above-ground biomass assessment. Bean yields from this study are reported elsewhere by [1]. The above ground shoots were cut with a knife at about 1 cm above the soil surface. Plant samples were dried to constant weight at 70 °C in an oven for 48 h and ground into fine powder for the analysis of P by dry ashing method [35]. Phosphorus in the digest was measured by the molybdenum blue method as described by [36]. Calcium concentrations were determined by atomic absorption spectrophotometer as described by [37]. The measured concentrations of nutrients were multiplied by plant dry matter to obtain the above ground biomass amounts of the mineral. At harvest, soil adhering to the roots was collected from around 10 bean plants from each plot and analysed for pH, P and calcium as described above.

2.4 Statistical analysis

Dry matter yield, P and Ca concentration and amounts in the tissues were analyzed using one-way analysis of variance (ANOVA). These computations were done with the software program STATISTICA [38]. The Fisher least significance difference (LSD) was used to compare treatment means at $P \leq 0.05$.

3. RESULTS AND DISCUSSION

3.1 Soil analysis

Results of soil analysis are shown in Table 1 and reported elsewhere by [2]. Generally, P was low at all the sites with reference to optimum level of 11 mg P kg⁻¹ soil as proposed by [39] for bean production. The exchangeable Ca ranged from 3.39 - 5.59 cmol kg⁻¹ (Table 1). The proposed optimum level of Ca for the majority of crops is 5.0 cmol kg⁻¹ [40] Four out of the six sites contained Ca levels lower than the proposed optimum levels and hence should receive some form of amelioration.

Table 1. **Selected chemical properties of soils collected from the Western Usambara mountains in the Lushoto District of the Tanga Region, Tanzania**

Site	Chemical properties of soils*		
	pH H ₂ O	Bray 1 P	Ca
		mg kg ⁻¹	cmol kg ⁻¹
Magamba	4.9	4.20	3.39
Ubiri	5.8	3.16	3.99
Ngulwii	5.5	2.80	5.59
Soni	5.9	1.68	5.59
Mbuzii	5.9	2.80	4.79
Mkuzi	5.2	4.20	4.79
Average	5.5	3.14	4.59

*Source: Ndakidemi and Semoka (2006).

3.2 Dry matter yield

There was a small variation in dry matter yield across sites (Table 2) suggesting that to a greater extent, the six farmers' fields were fairly uniform. The dry matter (DM) yield of bean responded strongly ($P \leq 0.001$) to all amendments used in this study (Table 3) indicating that P was among the limiting nutrients in the study area. Dry matter yields also correlated significantly with their shoot amounts P (Table 3; $r = 0.97$, $P = .05$) and Ca ($r = 0.86$, $P = .05$) suggesting that the **dry matter** was related to their amounts. Moreover, there was significant correlation between the amendment treatments and **dry matter** ($r = 0.83$, $P = .05$) indicating insufficient P and probably Ca in soils used in the study (Table 1). The dry matter increased by more than twofold by adding **Triple Super Phosphate** alone or Tughutu combined with **Minjingu phosphate rock** as compared with the control treatment. Addition of Tughutu or **Minjingu phosphate rock** alone increased the **dry matter** by more than 50% as compared to the control treatment. However, combining Tughutu and **Triple super phosphate** gave the best results by increasing the **dry matter** more than threefold as compared with the control. In general, **Minjingu phosphate rock** or Tughutu supplied alone produced comparable **dry matter** yield, whereas there was no significant **dry matter** yield difference between **Triple Super Phosphate** or Tughutu combined with **Minjingu phosphate rock**. (Table 3). The observed variations in **dry matter** are partly explained by the chemical composition of the materials used in this study [28, 23]. Yield from **Minjingu phosphate rock** or Tughutu treatment were generally lower than those of **Triple Super Phosphate** owing to low P contents in these materials [28, 23]. Generally there was a positive effect on **dry matter** yield by combining **Minjingu phosphate rock** or **Triple Super Phosphate** with Tughutu. This work is consistent with the work done by [34,15,29] that reported yield increases in maize by combining *Tithonia* or *Tephrosia vogelii* with **Triple Super Phosphate** or **Minjingu phosphate rock**.

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162 Table 2. Estimates of micro-site variations on dry matter yield of common bean in
 163 WUM, northern Tanzania
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Site	Dry matter yield** g plant ⁻¹
Magamba	17.95
Ubiri	18.24
Ngulwii	17.55
Soni	19.17
Mbuzii	16.95
Mkuzi	20.94
Average	18.47

165 **Dry matter was determined by taking the average yield across all treatments from
 166 each site.
 167

168 3.3 Phosphorus and Calcium amount in shoots

169 Significant variation on P and Ca amounts in shoots were observed ($P \leq 0.001$) in all plots
 170 that were supplied with the amendments (Table 3). The amount of P and Ca in this study
 171 almost paralleled the dry matter yield (Table 3). This suggests that P and Ca were among
 172 the major limiting elements for bean growth in the Western Usambara Mountains (Table 3;
 173 [2,3]). Combining Tughutu and Triple Super Phosphate resulted in significantly higher P
 174 amount compared with all other treatments tested and was followed by Tughutu combined
 175 with Minjingu phosphate rock (Table 3). The good response of Triple Super Phosphate as
 176 compared with Minjingu phosphate rock could be due to the higher and faster dissolution
 177 rate of Triple Super Phosphate than Minjingu phosphate rock that improved their
 178 concentration in the soil (Table 4), in shoots (data not shown) and made them available for
 179 plant uptake (Table 3). Similar results have also been reported by [24,33] in maize.

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181 Calcium amount in shoots almost followed the trend of P, but combining Tughutu with
 182 Minjingu phosphate rock or Triple Super Phosphate produced the best results that were not
 183 significantly different (Table 2). A comparative study on the properties of Tughutu and other
 184 organic sources from Western Usambara Mountains revealed that Tughutu was superior to
 185 most of them in N, P and K content [17] Furthermore, mineralization patterns of Tughutu
 186 were faster and higher for N, P and K as compared with those of Tithonia and other useful
 187 shrubs. Therefore, high qualities of this plant render it a good source of nutrient for bean
 188 growers in Western Usambara Mountains.

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In this study, combining organic material with Triple Super Phosphate or Minjingu phosphate rock significantly promoted the uptake of P and Ca in bean. This was also reflected in the dry matter yield (Table 2). Research evidence suggests that Ca plays a crucial role in plant growth and development. This is responsible for structural rigidity of the cell wall, controlling the cell membrane structures and functions, and in many different signal transduction processes [41], which ultimately may contribute significantly in regulating plant growth and development as observed in this study (Table 2). Results from this study showed that the amount of P and dry matter yield in beans was significantly enhanced with the addition of Tuguthu, Minjingu phosphate rock and the combination of Tuguthu with Minjingu phosphate rock or Triple Super Phosphate (Table 2 & 3). Phosphorus plays a key role in a plant's energy transfer mechanism (high energy phosphate), an essential intermediary in the metabolism of carbohydrate, protein and phospholipid transformations in plants and final plant growth [6]. Taken together, these results suggest that small-scale farmers in Western Usambara Mountains have the potential to benefit from the biomass transfer technology combined with a cheap inorganic resource such as Minjingu phosphate rock in improving their crop production.

Table 3. Effects of Tuguthu application, Minjingu phosphate rock or Triple Super Phosphate supply on dry matter yield (DMY), P and Ca amount in shoots of Phaseolus vulgaris L measured at 45 days after planting.

Treatments	Dry matter yield g.plant ⁻¹	P mg.plant ⁻¹	Ca mg.plant ⁻¹	P kg.ha ⁻¹	Ca kg.ha ⁻¹
P ₀ +T ₀ (Control)	9.68±0.26d	12.31±0.80d	116.81±4.02d	2.46±0.16d	23.36±0.80d
T ₀ + MPR	15.58±0.66c	26.66±2.12c	302.38±23.34bc	5.33±0.42c	60.48±4.67bc
T ₀ + TSP	19.88±0.69b	38.86±2.46b	326.34±21.21b	7.77±0.49b	65.27±4.24b
T	15.27±0.81c	24.06±1.88c	216.34±18.97c	4.81±0.38c	43.27±3.79c
T+ MPR	22.03±1.13b	45.07±3.75b	557.51±61.05a	9.01±0.75b	111.50±12.21a
T+ TSP	28.35±1.92a	66.80±6.86a	529.80±41.23a	13.36±1.37a	105.92±8.25a
F - Statistics	37.84***	28.97***	26.48***	28.97***	26.48***

*** Significant at P≤ 0.001.

a) P₀ = zero level of phosphorous; T₀ = zero level of Tuguthu; Minjingu phosphate rock 26 kg P ha⁻¹ = 26 kg P ha⁻¹ of Minjingu phosphate rock; Triple Super Phosphate 26 kg P ha⁻¹ = 26 kg P ha⁻¹ of Triple Super Phosphate; T2.5 t ha⁻¹ = 2.5 t of Tuguthu ha⁻¹

b) Mean values within a column followed by different letter(s) differ significantly at P=0.05 according to Fishers least significance difference.

3.4 Soil pH, residual P and Ca at harvest

The treatments supplied significantly (P<.05) changed the soil pH, P and Ca concentration in the soil solution at harvest (Table 4). Compared with control, Triple Super Phosphate supplied alone significantly decreased the soil pH more than with all other treatments. [42] and [43] also reported significant pH decreases with Triple Super Phosphate supply. However, the addition of Tuguthu, Minjingu phosphate rock and the combination of Tuguthu with Minjingu phosphate rock or Triple Super Phosphate increased the soil pH as compared with Triple Super Phosphate and the control treatment (Table 4). The increase in soil pH owing to Minjingu phosphate rock can be owing to reasonably higher Ca content in this material [23]. Minjingu phosphate rock in this context provided a liming effect to the soil and hence raised its pH. Similar increases in soil pH with Minjingu phosphate rock applications and use of organic amendments have been reported by [34, 16, 15, 30, 31]

231 The effect of Tuguthu, Minjingu phosphate rock, Triple Super Phosphate and/or their
 232 combination on residual P in the soil at harvest is shown in Table 4. Their application in the
 233 soil significantly ($P=0.05$) increased the residual P in the soil. In this study, the use of Tuguthu
 234 combined with Triple Super Phosphate or Minjingu phosphate rock or Triple Super
 235 Phosphate applied alone elevated the residual P above the optimum level of 11 mg P kg^{-1}
 236 soil for bean production [39]. Addition of Tuguthu alone doubled the residual P in the soil as
 237 compared with control, but the level was still low. The observed small increase in soil P with
 238 Tuguthu application was probably from the decay and the release into the soil of the
 239 organically bound P held in this material. These results suggest that Tuguthu supplied alone
 240 has lower P input in the soil. Addition of Minjingu phosphate rock alone was not sufficient to
 241 raise the residual P value to the proposed optimum level (Table 4). It is possible that soil
 242 conditions were not favourable for Minjingu phosphate rock solubility and release of enough
 243 P into the soil in the first year. Amorphous rock phosphate could be weathered and released
 244 for plant growth over time [44, 45]. Combining Minjingu phosphate rock with Tuguthu
 245 released P above the proposed optimum level of 11 mg P kg^{-1} soil for bean production
 246 (Table 4). Studies have shown that organic acids released during the decomposition of
 247 organic materials improved the availability of soluble P from rock phosphates [14, 16, 6 15,
 248 30, 31). These findings suggest that repeated addition of inorganic phosphorus applied
 249 alone or in combination with Tuguthu could build soil P reserves to reasonable levels and
 250 sustain crop productivity in Western Usambara Mountains.

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252 The addition of Tuguthu, Minjingu phosphate rock, Triple Super Phosphate and/or their
 253 combination significantly ($P \leq 0.05$) influenced the residual exchangeable Ca in the soil (Table
 254 4). The proposed optimum level of exchangeable Ca in the soil is 5.0 cmol kg^{-1} [40]. Except
 255 Triple Super Phosphate and control, other treatments increased the exchangeable Ca above
 256 the proposed optimum levels in the following order: Tuguthu + Minjingu phosphate rock >
 257 Minjingu phosphate rock > Tuguthu + Triple Super Phosphate > Tuguthu > Triple Super
 258 Phosphate > control (Table 4). All treatments involving Minjingu phosphate rock were
 259 superior in increasing residual soil Ca as compared with others. Similar observations in
 260 which the addition of Minjingu phosphate rock in soils increased the residual Ca as
 261 compared with Triple Super Phosphate have been reported by [32]. Minjingu phosphate rock
 262 has a reasonably higher Ca content than Triple Super Phosphate [23] and Tuguthu [17].

263

264 Table 4. Effects of Tuguthu application, Minjingu phosphate rock or Triple Super Phosphate
 265 supply on pH, residual P and Ca in the soil collected from rhizosphere of *Phaseolus vulgaris*
 266 L. plants at harvest.

Treatments	pH H ₂ O	P mg.kg ⁻¹	Ca cmol.kg ⁻¹
P ₀ + T ₀ (Control)	5.32±0.16bc	3.28±0.41d	4.52±0.40b
T ₀ + MPR	5.86±0.18a	10.59±0.47b	5.99±0.45a
T ₀ + TSP	5.23±0.16c	14.76±0.80a	4.72±0.41b
T	5.79±0.20ab	6.93±0.51c	5.01±0.41ab
T + MPR	5.98±0.20a	14.61±1.23a	6.14±0.41a
T + TSP	5.71±0.17abc	16.79±1.06a	5.05±0.37ab
F - Statistics	2.92*	41.77*	2.68*

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* Significant at $P=0.05$.

a) P_0 = zero level of phosphorous; T_0 = zero level of Tughutu; Minjingu phosphate rock 26 kg P ha⁻¹ = 26 kg P ha⁻¹ of Minjingu phosphate rock; Triple Super Phosphate 26 kg P ha⁻¹ = 26 kg P ha⁻¹ of Triple Super Phosphate; T2.5 t.ha⁻¹ = 2.5 t of Tughutu ha⁻¹
b) Mean values within a column followed by different letter(s) differ significantly at $P=0.05$ according to Fishers least significance difference.

4. CONCLUSION

The inherently low soil fertility status in most parts of the Western Usambara Mountains makes sustaining agricultural production difficult. This is the first work to evaluate Tughutu alone and/or in combination with phosphate rock or Triple super phosphate on their influence on growth, amount of phosphorous and calcium content in bean shoots. The application of Tughutu, Minjingu Phosphate Rock or triple super phosphate and their combination showed significant synergistic effects in plant growth, amount of phosphorous and calcium and improvement in selected chemical soil properties measured at the end of the crop season. In conclusion, the locally available shrub (Tughutu) and Minjingu Phosphate Rock can constitute a very valuable and viable supplement to the industrial fertilizer sources in the Western Usambara Mountains. Future research could also focus on the relative cost (collecting, purchasing, supply of the products, transport and farm labour) of applying Tughutu and Minjingu Phosphate Rock in relation to that of triple super phosphate. There would appear to be no negative effects of Tughutu and Minjingu Phosphate Rock application on these soils or the bean crop.

ACKNOWLEDGEMENTS

This work was supported by the Directorate of Research and Development in the Ministry of Agriculture, Food Security and Cooperatives of Tanzania, through TARP II World Bank project.

AUTHORS' CONTRIBUTIONS

The Author is responsible for the whole study.

REFERENCES

- [1]Ndakidemi, P.A. Agronomic and Economic Potential of Tughutu (*Vernonia subligera* O.Hoffn) and Minjingu Phosphate Rock as Alternative P Sources for Bean Growers. *Pedosphere*. 2007;17 (6) ;732-738
- [2]Ndakidemi, P.A. Semoka, J.M.R.,. Soil fertility survey in western Usambara Mountains, northern Tanzania. *Pedosphere*. 2006;16; 237-244.
- [3]Ndakidemi, P.A. & Semoka, J.M.R., 1993. Soil fertility constraints to bean production in Lushoto District and their amelioration by application of lime, phosphorous and potassium. In Msumali, G.P. and Mabagala, R.B. (eds.), *Bean Research*, Vol. 8. Proceedings of 12th Bean Research Workshop, Sokoine University of Agriculture, Morogoro, Tanzania, 15-17 September, 1993, pp 32-50.

- 315 [4]Vance, C.P.Symbiotic Nitrogen Fixation and Phosphorus Acquisition. Plant Nutrition in a
316 World of Declining Renewable Resources. Plant Physiol. 2001;127; 390-377.
- 317 [5]Tang, C., Hinsinger, P., Drevon, J.J. & Jaillard, B. Phosphorus Deficiency Impairs Early
318 Nodule Functioning and Enhances Proton Release in Roots of *Medicago truncatula* L.
319 Ann. Bot. 2001;88; 131-138.
- 320 [6]Vance, C.P., Uhde-Stone, C. & Allan, D.L. Phosphorus acquisition and use: critical
321 adaptations by plants for securing a nonrenewable resource. New Phytol. 2003;157; 423-
322 447.
- 323 [7]Greenwood, E.A.N. & Hallsworth, E.G.Studies on the nutrition of forage legumes. II. Some
324 interactions of Ca, P, Cu and Mo on the growth and chemical composition of *Trifolium*
325 *subterraneum* L. Plant Soil. 1960;12;97-127.
- 326 [8]Banath, C.L., Greenwood, E.A.N. Loneragan, J.F. Effects of calcium deficiency on
327 symbiotic nitrogen fixation. Plant Physiol. 1966;41;760-763.
- 328 [9]Lowter, W.L. Loneragan, J.F. Calcium and nodulation in subterranean clover (*Trifolium*
329 *subterraneum* L.). Plant Physiol. 1968;43;1362-1366. on P availability. Soil Biol. Biochem.
330 2003c ;35;207-216.
- 331 [10]Muns, D.N.Nodulation of *Medicago sativa* in solution culture. V. Calcium and pH
332 requirements during infection. Plant Soil. 1970;32; 90-102.
- 333 [11]Blevins, D.G., Barnett, N.M. & Bottino, P.J. The effects of calcium and the ionophore
334 A23187 on nodulation, nitrogen fixation and growth of soybeans. Physiol. Plant.
335 1977;41;235-238.
- 336 [12]Streeter, J.G.,. Effect of elevated calcium concentration in infected cells of soybean
337 (*Glycine max* L. Merr.) nodules on nitrogenase activity and N input to the plant. J. Exp.
338 Bot. 1998;49;997-1003.
- 339 [13]Lyamchai, C.J., Lyiom, S.D., Ndondi, R.V., Owenya, M.Z., Ndakidemi, P.A. Massawe,
340 N.F. Participatory Rural Appraisal for Kwalei Catchment, Lushoto District. Selian
341 Agricultural Research Institute, Arusha, Tanzania. 1998; 71pp.
- 342 [14]Ikerra, T.W.D, Mnkeni P.N.S. & Singh, B.R. Effects of added compost and farmyard
343 manure on P release from Minjingu phosphate rock and its uptake by maize. Norw. J.
344 Agr. Sci. 1994;8;13-23.
- 345 [15]Ikerra, S.T. Use of Minjingu phosphate rock combined with different organic inputs in
346 improving phosphorus availability and maize yields on a Chromic Acrisol in Morogoro,
347 Tanzania, Ph.D Thesis. Sokoine University of Agriculture, Morogoro, Tanzania.
348 2004.262p.
- 349 [16]Savini, I. The effect of organic and inorganic amendments on phosphorous release and
350 availability from two phosphate rocks and triple super phosphate in phosphorous fixing
351 soils, MSc Thesis. University of Nairobi, Kenya. 2000;104 pp
- 352 [17]Wickama, J. M. & Mowo, J. G.Using local resources to improve soil fertility in Tanzania.
353 Managing Africa's Soils Working Paper No. 21. International Institute for Environment and
354 Development (IIED). London. 2001;24pp.

- 355 [18]Kwabiah, A.B., Palm, C.A., Stoskopf, N.C. & Voroney, R.P. Response of soil microbial
356 biomass dynamics to quality of plant materials with emphasis
- 357 [19]George, T.S., Gregory, P.J., Wood, M., Read D. & Buresh, R.J. Phosphatase activity and
358 organic acids in the rhizosphere of potential agroforestry
- 359 [20]Kwabiah, A.B., Stoskopf, N.C., Palm, C.A. VOoroney, R.P. Soil Availability as affected
360 by the chemical composition of plant materials: implications for P-limiting agriculture in
361 tropical Africa. Agr. Ecosyst. Environ. 2003a;100; 53-61.
- 362 [21]Kwabiah, A.B., Stoskopf, N.C., Palm, C.A., Voroney, R.P., Rao, M.R. Gacheru, E.
363 Phosphorus availability and maize response to organic and inorganic fertilizer inputs in a
364 short-term study in western Kenya. Agr. Ecosyst. Environ. 2003b; 95; 49-59.
- 365 [22]Weil, R.R. Soil and Plant Influences on Crop Response to Two African Phosphate
366 Rocks. Agron. J. 2000;92;1167-1175.
- 367 [23]Szilas, C. The Tanzanian Minjingu phosphate rock – possibilities and limitations for direct
368 application. PhD Thesis, Royal Veterinary and Agricultural University, Copenhagen
369 (<http://www.igv.kvl.dk/-casz>) .2002
- 370 [24]Msolla, M.M. Agronomic evaluation of maize performance using hard and soft Minjingu
371 phosphate rocks and in mixtures with triple phosphate on selected Tanzanian soils.
- 372 [25]Mnkeni P.N.S., Semoka, J.M.R. & Buganga B.B.S. Agronomic effectiveness of Minjingu
373 phosphate rock as a source of phosphorus for maize in some soils of Morogoro District,
374 Tanzania. Zimbabwe J. Agric. Res. 1991;30; 27-37.
- 375 [26]Semoka J.M.R., Mnkeni, P.N.S. & Ringo, H.D. Effectiveness of Tanzanian phosphate
376 rocks of igneous and sedimentary origin as sources of phosphorus for maize. Zimbabwe
377 J. Agric. Res. 1992;30;127-136.
- 378 [27]Haque, I., Lupwayi, N.Z. & Ssali, H., Agronomic effectiveness of unacidulated and
379 partially acidulated Minjingu rock phosphate on *Stylosanthes guianensis*. Trop.
380 Grasslands. 1999 ;33;159-164.
- 381 [28]Mowo, J.G. Effectiveness of phosphate rock on Ferralsols in Tanzania and the influence
382 of within field variability, Ph.D Thesis. Wageningen Agricultural University, The
383 Netherlands, 2000;164 pp.
- 384 [29]Mkangwa, C.Z. Influence of Minjingu Phosphite Rock Application on *Tephrosia vogelii*
385 Fallow Productivity and Subsequent Maize Response on Ferralsol in Morogoro,
386 Tanzania. PhD Thesis, Sokoine University of Agriculture, Morogoro, Tanzania,
387 2004;227pp.
- 388 [30]Savini, I., Smithson, P. & Karanja, N. Effects of added biomass, soil pH and calcium on
389 the solubility of Minjingu phosphate rock in a Kenyan Oxisol. Arch. Agron. Soil Sci. 2006a;
390 52, 19-36.
- 391 [31]Savini, I., Smithson, P.C., Karanja, N.K., Yamasaki, H. Influence of *Tithonia diversifolia*
392 and triple superphosphate on dissolution and effectiveness of phosphate rock in acidic
393 soil. J. Plant Nutr. Soil Sci. 2006b;169;593-604.

- 394 [32]Szilas, C., Semoka, J.M.R. Borggaard, O.K. Can local Minjingu phosphate rock replace
395 superphosphate on acid soils in Tanzania? Nutr. Cycl. Agroecosys. 2006;77;257-268.
- 396 [33]Msolla, M.M., Semoka, J.M.R., Szilas, C. & Borggaard, O.K. Crop (Maize) Response to
397 Direct Application of Local Phosphate Rock on Selected Acid Soils of Tanzania. Comm.
398 Soil Sci. Plant Anal. 2007;38;93-106.
- 399 [34]Osunde, A.O., Zapata, F. & Sangiga, N., 1992. Agronomic evaluation of rock Phosphate
400 as a phosphorus sources for *Leucaena leucocephala* grown on an altisol. In Proceedings
401 of the African Association of Biological Nitrogen Fixation (AABNF) held at the
402 International Institute of Tropical Agriculture (IITA), Ibadan Nigeria, 24-28 September,
403 1990.
- 404 [35]Chapman, H.D. & Pratt, P.F. Methods of Analysis for Soils, Plants and
- 405 [36]Murphy, J. & Riley, J.P. A modified single solution method for the determination of
406 phosphate in natural waters. Anal. Chim. Acta. 1962;27; 31-36.
- 407 [37]Hesse, P.R. A text book of soil chemistry analysis. John Murray Ltd. London. 1971 pp
408 120-121 and 309.
- 409 [38]STATISTICA. StatSoft, Inc. 2300 East 14th Street, Tulsa, OK 74104, USA.2008
- 410 [39]Schwatz, H.F. & Corrales, M.A.P. Nutritional disorders in beans. In Schwatz, H. F., and
411 Corrales, M. A. P. (eds.) Bean Production Problems in the Tropics. CIAT, Cali, Columbia.
412 1989; 57-604.
- 413 [40]Marx, E.S., Hart, J. & Steven, R.G. Soil Test Interpretation Guide. Oregon State
414 University extension services. Oregon State University, Corvallis. 1996;7pp.
- 415 [41]Helper, P.K.,. Calcium: A Central Regulator of Plant Growth and Development. Plant
416 Cell. 2005;17;2142-2155.
- 417 [42]Robinson, J.B.D. The influence of fertilizer and manure on the pH reaction of a coffee
418 soil. E. Afr. Agric. and For. J. 1956;22;76-80.
- 419 [43]Huffman E O. Reaction of phosphate in soils: recent research by TVA. Proceeding 71.
420 The Fertilizer Society, London. 1962.
- 421 [44]Chieh, S.H. & Menon, R.G. Factors affecting the agronomic effectiveness of phosphate
422 rock for direct application. Fert. Res. 1995 ;41, 227-234.
- 423 [45]Smalbarger, S.A. Singh, U., Chien, S.H., Henao, J. & Wilkens, P.W.,. Development and
424 validation of a phosphate rock decision support system. Agron. J. 2006;98, 471-483